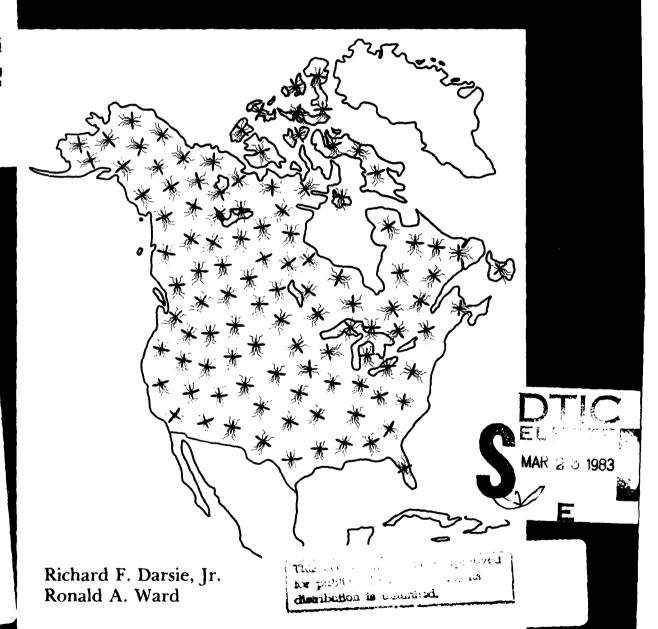


MICROCOPY RESOLUTION TEST CHART NATIONAL BUREAU OF STANDARDS-1963-A

Mosquitoes of North America,

North of Mexico



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ABBREVIATIONS OF THE STATES OF THE UNITED STATES OF AMERICA AND THE PROVINCES OF CANADA

United States

	* "				
AL —	Alabama	MI —	Michigan	UT —	Utah
AK —	Alaska	MN —	Minnesota	VA —	Virginia
AR —	Arkansas	MO —	Missouri	VT —	Vermont
AZ	Arizona	MS —	Mississippi	WA	Washington
CA —	California	MT —	Montana	WI —	Wisconsin
co —	Colorado	NE —	Nebraska	wv —	West Virginia
CT —	Connecticut	NC —	North Carolina	WY —	Wyoming
DC —	District of	ND —	North Dakota	Canada	
	Columbia	NH —	New Hampshire		– Alberta
DE —	Delaware	NJ —	New Jersey	BC —	British Columbia
FL —	Florida	NM —	New Mexico		Labrador*
GA —	Georgia	NV —	Nevada		- Manitoba
IA —	Iowa	NY —	New York		
ID —	Idaho	OH —	Ohio		New Brunswick
IL —	Illinois	OK —	Oklahoma		- Newfoundland
IN —	Indiana	OR —	Oregon	NS —	Nova Scotia
KS —	Kansas	PA —	Pennsylvania	NWT —	- Northwest Territories
KY —	Kentucky	RI —	Rhode Island	ONT	Ontario
LA —	Louisiana	SC —	South Carolina	PEI —	Prince Edward
MA —	Massachusetts		South Caronna South Dakota	14.1 —	Island
MD —	Maryland	SD —		PQ —	Quebec
ME —	Maine	TN —	Tennessee	SASK —	- Saskatchewan
		TX —	Texas	YUK —	
					

^{*}Although Labrador is now part of Newfoundland, mosquito records are here listed separately.

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19. KEY WORDS (Continue on reverse side if necessary and identity by block number)

MOSQUITOES, CULICIDAE, NORTH AMERICA



20. ABSTRACT (Continue on reverse side if necessary and identify by block number)

CORRECTIONS AND ADDITIONS ARE PROVIDED AS A SUPPLEMENT TO THE PUBLICATION, IDENTIFICATION AND GEOGRAPHICAL DISTRIBUTION OF THE MOSQUITOES OF NORTH AMERICA, NORTH OF MEXICO.

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Corrections and Additions to the Publication, <u>Identification</u>

and <u>Geographical Distribution of the Mosquitoes</u>

of North America, North of Mexico

Ronald A. Ward and Richard F. Darsie, Jr. 2

ABSTRACT. Corrections and additions are provided as a supplement to the publication, <u>Identification and Geographical Distribution of the Mosquitoes</u> of North America, North of Mexico.

INTRODUCTION

Following the publication of Darsie and Ward (1981), certain errors and corrections have been brought to our attention and are appended below in an Errata section. As the chapters, Bibliography of Mosquito Taxonomy and Geographical Distribution, and Addendum to Bibliography in Darsie and Ward (1981) surveyed the relevant literature through most of 1979, we are providing annotated references from 1979 to mid-1982. In addition, several pre-1979 publications are cited that were earlier omitted. Certain species merit comment and are noted along with new distribution records at the state (U.S.A.) and province (Canada) level.

We wish to thank Peter Belton, Alain Maire and Lewis T. Nielsen who have provided data for our use and Bruce Harrison for his advice on an earlier draft.

ERRATA

Page 7, line 19
For Interocular setae (IS), read Interocular setae (ISe)

Page 8, lines 39, 43, 48
For meskatepisternum, read mesokatepisternum

Page 9, line 39 For Fig. 15, read Fig. 38

Ronald A. Ward, Department of Entomology, Walter Reed Army Institute of Research, Washington, D. C. 20012

²Richard F. Darsie, Jr., Division of Parasitic Diseases, Chamblee 23, Center for Infectious Diseases, Centers for Disease Control, Atlanta, Georgia 30333.

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Page 9, line 41
     For Fig. 221, read Fig. 222
Page 10, 1ine 49
     For the, read The
Page 14, In list of abbreviations for Plate 4, add PMe-pleural membrane
Page 27, Couplet 1, lines 1 and 2
     For hindtarsomere, read hindtarsomeres
Page 30, Figs. 59, 61
     For Hindtarus, read Hindtarsus
Page 44, Fig. 140
     For candensis, read canadensis
Page 51, line 2
     For posterior, read anterior
Page 115, line 44
     For Fig. 789, read Fig. 815
Page 115, !ine 45
     For Fig. 815, read Fig. 784
Page 116, line 25
     For Figs. 514, 516, read Figs. 514, 516, 855
Page 119, 1ine 7
     For mesurement, read measurement
Page 132, Couplet 11
     Should read:
       11 (10).
                Saddle completely encircling segment X (Fig. 538),
                or if not, siphon aciculate (Fig. 824) . . . . . . . . .
                Saddle not completely encircling segment X (Fig. 539);
                                                               (in part) Aedes
                Page 133, Couplet 14, lines 2 and 3
     For tergum on VII, read segment VII
Page 182, Figs. 790, 792
     For 1-I-IV, read 1-IV
Page 199, Figs. 878, 880
     For Ventral view, read Dorsal view
Page 202, Fig. 895
     For inormator, read inormata
```

Page 214, Couplet 8
For johnstonni, read johnstonii

Page 215, Fig. 958
For abdomen Ps., read abdomen - Ps.

Page 220, Fig. 983
For thorax Wy., read thorax - Wy.

Pages 226 and 233, column 1
Heading should read Mosquito Species

Page 241, line 8
For MAN (470), read MAN (70, as Ae. triseriatus)

Page 258, line 5 For NE (164), read NE (373)

Page 260, Plate 35, line 2 For NM 3/8K_!(, read (502)

Page 260, Plate 35, line 3 For MQ, read MO

Page 287, ref. 311
For silvertris, read silvestris

Page 290, ref. 398 For Schyler, read Schuyler

Page 297, line 12 For 57, 60, read 57-60

Page 305, Fig. 670 For Pina, read Pima

Page 312, line 46
For mulrennai, read mulrennani

Page 313, line 4
For s. spenceri, read s. spencerii

COMMENTS ON CERTAIN SPECIES

Aedes (Aedes) hemiteleus Dyar. Bickley (1980) studied the adult morphological characters of the North American members of the Cinereus Group (sensu Peus 1972) and found them too variable to discriminate Ae. hemiteleus from Ae. cinereus Meigen, or other members of the group. However, in the absence of adequate North American specimens with associated immature skins, we feel it is not advisable to synonymize Ae. hemiteleus with Ae. cinereus.

Aedes (Finlaya) togoi (Theobald). Belton (1980) presented conclusive evidence that Ae. togoi is now established south of Anacortes, Washington, in rock pools along the coastline. He indicated that Ae. togoi is probably a recent introduction from Asia (within the past 40 years) and may have been dispersed through the agency of ferry traffic among islands. Since this species is already included in Darsie and Ward (1981), no modifications to the keys are required.

Culex (Melanoconion) taeniopus Dyar and Knab. The new synonomy of Cx. (Mel.) opisthopus Komp with taeniopus by Sirivanakarn and Belkin (1980) will undoubtedly cause consternation among some North American mosquito workers. Through the proper examination of type-specimens and the application of the principle of priority, Sirivanakarn and Belkin (1.c.) have resolved a difficult problem of nomenclature.

Culex (Tinolestes) latisquama (Coquillett). The presence of this species in the U. S. is based on one male supposedly collected in 1906 from Estero, Lee county, Florida (Stone 1968). Since additional specimens have not been collected north of Honduras, Berlin and Belkin (1980) believed the Florida record is erroneous. They attribute this to an incorrect label on specimens collected for the "Mosquitoes of North and Central America and the West Indies" (Howard, Dyar and Knab 1915). As we do not wish to perpetuate the error, Cx. latisquama is removed from the list for the subject area. This reduces the list of species and subspecies known to occur in North America, north of Mexico, from 167 to 166.

Culiseta (Culiseta) annulata (Schrank). Faran and Bailey (1980) collected a single adult female Cs. annulata at Fort McHenry, Baltimore, Maryland, from the inner walls of an old munitions bunker. This female was collected in association with overwintering Cx. pipiens Linn. females and survived 20 days in the laboratory. Due to the proximity of Fort McHenry to Baltimore harbor, it is probable that this specimen or its ancestors were introduced into the area by a ship travelling from Europe. Since further specimens have not been reported, Cs. annulata is not yet an established faunal component.

Wyeomyia (Wyeomyia) smithii (Coquillett) and Wy. (Wyo.) haynei Dodge. Bradshaw and Lounibos (1977) examined the effects of latitude, altitude and longitude on photoperiodicity, morphology of larval anal papillae, stage of dormancy and response of F_1 hybrids of pitcher-plant mosquitoes of the genus Wyeomyia from eastern North America. They conclude that Wy. smithii is probably a polytypic species which includes Wy. haynei as a geographic subspecies. Their observation that Wy. haynei populations from the Gulf Coast possess four long anal papillae is of interest and indicates the need for a rigorous taxonomic analysis of extensive series of reared specimens of both species throughout the entire range. These should be complemented by a scries of cross-breeding experiments from various critical populations. Until these studies have been accomplished, haynei and smithii should be retained as discrete species.

CHANGES TO-STATE AND PROVINCE DISTRIBUTION RECORDS

The following new United States and Canada distribution records have come to our attention since Darsie and Ward (1981) was submitted to the printer. Changes in distribution such as county records are included in the Annotated References.

SPECIES	POLITICAL UNIT	REFERENCE
Ae. atropalpus	IN	Restifo and Lanzaro (1980)
Ae. atropalpus	NFLD	Nielsen and Mokry (1982b)
Ae. cinereus	WV	Butler and Amrine (1980)
Ae. decticus	NFLD	Nielsen and Mokry (1982b)
Ae. diataeus	NFLD	Nielsen and Mokry (1982b)
Ae. dorsalis	NH	Burger (1981)
Ae. euedes	WY	Nielsen (1982)
Ae. grossbecki	MI	Cassani and Newson (1980)
Ae. hendersoni	FL	Zavortink and Belkin (1979)
Ae. hexodontus	NFLD	Nielsen and Mokry (1982b)
Ae. nevadensis	ВС	Belton and Belton (1981)
Ae. pionips	NFLD	Nielsen and Mokry (1982b)
Ae. sollicitans	WV	Butler and Amrine (1980)
Ae. sticticus	NFLD	Nielsen and Mokry (1982b)
Ae. togoi	WA	Belton (1980)
Cs. alaskaensis	UT	Bickley (1979)
Cs. melanura	NFLD	Nielsen and Mokry (1982a)
Or. alba	WV	Heaps (1980)
Ps. cyanescens	IA	Ritchie and Rowley (1980)
Ps. mathesoni	MI	Cassani and Newson (1980)
Ur. sapphirina	CO	Maloney (1980)

Four species were deleted from the list of Mebraska species by Lunt and Rapp (1981) as they were unable to verify their presence within the state on the basis of valid collection records. These include: Aedes fitchii (Felt and Young), Ae. implicatus Vockeroth, Culex quinquefasciatus Say and Culiseta melanura (Coquillett).

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IDENTIFICATION AND GEOGRAPHICAL DISTRIBUTION OF THE MOSQUITOES OF NORTH AMERICA, NORTH OF MEXICO

by

Richard F. Darsie, Jr.

Medical Entomology Research and Training Unit/Guatemala Bureau of Tropical Diseases Center for Disease Control Atlanta, Georgia, 30333

and

Ronald A. Ward

Department of Entomology Walter Reed Army Institute of Research Washington, D. C. 20012

With illustrations by

Chien C. Chang



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PREFACE

This publication was conceived by Harold D. Chapman in 1975 while he was serving as President of the American Mosquito Control Association. Since that time it has been supported by Presidents D. Bruce Francy, Lewis T. Nielsen, Paul A. Hunt, Glenn W. Stokes and Robert K. Washino, their respective Boards of Directors, the publications committees and Executive Directors Thomas A. Mulhern and W. Donald Murray. The publication's Editorial Board was composed of William E. Bickley, John D. Edman, Lewis T. Nielsen and the authors.

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INTRODUCTION

In 1955 Carpenter and LaCasse published a monograph entitled *Mosquitoes of North America*, *North of Mexico* (106).* They included 143 species and subspecies in 11 genera and 19 subgenera, identification keys to genera and species, and descriptions of the known adult female and larval stages. Their geographical distribution information consisted of lists of the states of the United States of America (USA) and provinces of Canada in which each taxon had been collected, with substantiating references.

There are now known from the same region 167 species and subspecies in 13 genera and 27 subgenera. The additions and changes in the names of the North American mosquito fauna have been reviewed by Carpenter (91, 95, 104) and Darsie (140, 142). The principal objective of this volume purports to be the revision of the identification keys to adult female and larval stages which incorporate all 167 taxa. Each key is preceded by a detailed description of the morphology of that stage, which is needed to use the key successfully. In addition, each couplet of the two keys is illustrated to assist the user in interpreting the characters employed.

A second purpose is to present up-to-date information on the geographical distribution of the mosquito taxa. We are continuing the arrangement used by Carpenter and LaCasse (106), listing the states and provinces from which each taxon has been reported with substantiating references. In addition, we are depicting the distribution on maps; actually the captions of the map plates (Plates 9-49) contain the specific states/provinces in which each taxon is found. Using Carpenter and LaCasse's monograph (106) as a starting point and listing the state/province data given by them, we are adding a total of 574 new state/province records which encompasses the 16 new species described since 1955. Detailed also are 37 instances in which species once reported as occurring in particular states/provinces are deleted.

The morphological terms employed in this volume are substantially changed from those used by Carpenter and LaCasse (106). In 1970 Kenneth L. Knight and the late Jean L. Laffoon started an extensive study of mosquito morphology, the ultimate aim of which was to produce a standardized set of morphological terms, adequately defined. Knight and his associates prepared 16 scientific articles in the "mosquito taxonomic glossary" series, i.e., Harbach & Knight (1977 A, B, C, D; 1978 A, B, C; 1980), Knight (1970, 1971 A), Knight & Laffoon (1970 A, B, C; 1971 A, B) and Laffoon & Knight (1971, 1973). The terms proposed by them and used in this volume take into consideration homology, phylogeny, and their use generally among the dipterous insects. There is one exception: we continue to use "claw" instead of "unguis" as proffered. To assist in the transition, the older terms have been given after the new ones in the sections on morphology.

Another modification from the 1955 monograph is the adoption of the chaetotaxical nomenclature espoused by John N. Belkin (1950, 1952, 1953, 1954, 1960, 1962) and the abbreviations he used to designate parts of the body and setae borne on them, especially in the immature stages, e.g., T for metathorax and 6-T for seta 6 on that segment. This practice has been used by Belkin and his associates (1, 6, 7, 8, 34, 305, 306, 508, 511, 514) and by many other taxonomists (167, 174, 405, 501, 524).

This volume contains illustrated keys to adult females and fourth stage larvae. We strongly recommend that the user study the sections on morphology before starting to identify specimens. Wherever possible we have used adult characters which are the least disturbed by the mechanical light trapping process; but in some couplets, especially in the genus *Aedes*, the use of traits disrupted by trapping was unavoidable. The user should be familiar with the proper method of preserving mosquito larvae because the presence of a full complement of the appendages and setae is essential for their identification in our larval keys. We have also tried to quantify insofar as practicable all characters to reduce the guesswork in dealing with "relative" terms.

Below each species when it is named in the keys will be found the plate number on which its distribution is shown. The user can immediately ascertain if a determined species has been reported from the locality where it was collected. One taxon shown on a map (Plate 46) is not included in the keys, i.e., *Toxorhynchutes* sp., see discussion below.

An appendix provides locality data for the voucher specimens selected for illustration in the keys. These mosquitoes (or slides) have a "Fig. ___" label and are largely from the U.S. National Museum collection.

^{*}References cited by year are found in the Selected Bibliography of Mosquito Morphology at the end of the Morphology of Adult Female section; those cited by numbers are found in the Bibliography of Mosquito Taxonomy and Geographical Distribution at the end of the volume.

SYSTEMATICS

Mosquitoes belong to the phylum Arthropoda, class Insecta, order Diptera. They are bilaterally symmetrical insects, adults of which are covered with an exoskeleton, bearing jointed legs and two functional wings. A second pair of wings is represented by knobbed halteres. Mosquitoes may be distinguished from other dipterous insects by the presence of scales on the wing veins and by their mouthparts in the form of an elongate proboscis, adapted for piercing and sucking. They are holometabolous; therefore they have four dissimilar stages in their life cycle, i.e., egg, larva, pupa and adult. This volume deals with the adult female and fourth stage larva, which are so different in appearance that they seem not to be related.

It is assumed that the user can already recognize species belonging to the order Diptera and family Culicidae. If not, general references such as Borror et al. (1976) should be consulted.

In this volume we follow the classification of the family Culicidae as given by Knight and Stone (519) and Knight (518). We do not deal with suprageneric categories except to relate certain morphological structures as belonging to anophelines, referring to members of the subfamily Anophelinae, or culicine, meaning members of the subfamilies Culicinae and Toxorhynchitinae, as interpreted by Knight and Stone (*loc. cit.*). Also no infrasubspecies are considered in this work.

In Table 1 is the systematic index of the species of Culicidae now known from North America, north of Mexico, and after each taxon is given the zoogeographical region, area or specific country in which each is found outside the region being considered, if applicable. Those marked as indigenous are confined to the region.

Table 1. Systematic Index of the Culicidae of North America, North of Mexico and Distribution in other Regions, Areas or Specific Countries

Taxon	Extralimital Distribution	Taxon	Extralimital Distribution
Genus AEDES Meigen		canadensis canadensis (Theobald)	Mexico
Subgenus Abraedes Zavortink papago Zavortink	Indigenous*	canadensis mathesoni Middlekauff	Indigenous
Subgenus Aedes Meigen cinereus Meigen hemiteleus Dyar Subgenus Aedimorphus Theobald vexans (Meigen) Subgenus Finlaya Theobald togoi (Theobald)	Palearctic Indigenous Worldwide Palearctic Oriental	cantator (Coquillett) cataphylla (Dyar) churchillensis Ellis & Brust communis (De Geer) decticus Howard, Dyar & Knab deserticola Zavortink diantaeus Howard, Dyar & Knab dorsalis (Meigen)	Indigenous Palearctic Indigenous Palearctic Indigenous Indigenous Palearctic Palearctic
Subgenus Kompia Aitken purpureipes Aitken Subgenus Ochlerotatus Lynch Arribalzaga aboriginis Dyar	Mexico Indigenous	dupreci (Coquillett) epactius Dyar & Knab euedes Howard, Dyar & Knab exerucians (Walker) fitchir (Felt & Young)	Mexico Mexico Neotropical Palearctic Palearctic Palearctic
abserratus (Felt & Young) aloponotum Dyar atlanticus Dyar & Knab atropalpus (Coquillett) aurifer (Coquillett) bicristatus Thurman & Winkler bimaculatus (Coquillett) campestris Dyar & Knab	Indigenous Indigenous Indigenous Indigenous Indigenous Indigenous Accordopical Mexico	flavescens (Müller) fulvus pallens Ross grossbecki Dyar & Knab hexodontus Dyar impiger (Walker) implicatus Vockeroth increpitus Dyar onfirmatus Dyar	Palearctic Cuba Indigenous Palearctic Palearctic Palearctic Indigenous Mexico

^{*} Indigenous means no extralimital distribution.

Taxon	Extralimital Distribution	Taxon	Extralimital Distribution
intrudens Dyar	Palearctic	barberi Coquillett	Indigenous
melanimon Dyar	Indigenous	bradleyi King	Mexico
mercurator Dyar	Palearctic	crucians Wiedemann	Neotropical
mitchellae (Dyar)	Mexico	earlei Vargas	Indigenous
monticola Belkin & McDonald	Mexico	franciscanus McCracken	Mexico
muelleri Dyar	Mexico	freeborni Aitken	Mexico
nevadensis Chapman & Barr	Indigenous	georgianus King	Indigenous
nigripes (Zellerstedt)	Palearctic	<i>judithae</i> Zavortink	Mexico
nigromaculis (Ludlow)	Mexico	occidentalis Dyar & Knab	Indigenous
niphadopsis Dyar & Knab	Indigenous	perplexens Ludlow	Indigenous
pionips Dyar	Palearctic	pseudopunctipennis Theobald	Neotropical
provocans (Walker)	Indigenous	punctipennis (Say)	Mexico
pullatus (Coquillett)	Palearctic	quadrimaculatus Say	Mexico
punctodes Dyar	Palearctic	walkeri Theobald	Mexico
punctor (Kirby)	Palearctic	Subgenus Nyssorhynchus	
rempeli Vockeroth	Palearctic	Blanchard	
riparius Dyar & Knab	Palearctic	albimanus Wiedemann	Neotropical
scapularis (Rondani)	Neotropical		•
schizopinax Dyar	Indigenous	Genus COQUILLETTIDIA Dyar	
sierrensis (Ludlow)	Indigenous	Subgenus Coquillettidia Dyar	
sollicitans (Walker)	Caribbean, Mexico	perturbans (Walker)	Mexico
spencerii idahoensis (Theobald)	Indigenous	Genus CULEX Linnaeus	
spencerii spencerii (Theobald)	Indigenous	Subgenus Culex Linnaeus	
squamiger (Coquillett)	Mexico	bahamensis Dyar & Knab	Caribbean
sticticus (Meigen)	Palearctic,	chidesteri Dyar	Neotropical
	Mexico	coronator Dyar & Knab	Neotropical
stimulans (Walker)	Indigenous	declarator Dyar & Knab	Neotropical
taeniorhynchus (Wiedemann)	Neotropical	erythrothorax Dyar	Mexico
theleter Dyar	Mexico	interrogator Dyar & Knab	Neotropical
thibaulti Dyar & Knab	Palearctic	nigripalpus Theobald	Neotropical
tormentor Dyar & Knab	Neotropical	peus Speiser	Neotropical
tortilis (Theobald)	Neotropical	pipiens Linnacus	Palearctic, S.
trivittatus (Coquillett)	Mexico		Neotropical, S.
varipalpus (Coquillett)	Indigenous		Ethiopian
ventrovittis Dyar	Indigenous	quinquefasciatus Say	Cosmotropical
Subgenus Protomacleaya Theobald		restuans Theobald	Mexico
brelandi Zavortink	Indigenous	salinarius Coquillett	Mexico
burgeri Zavortink	Mexico	tarsalis Coquillett	Mexico
hendersoni Cockerell	Indigenous	thriambus Dyar	Neotropical
triscriatus (Sav)	Mexico	Subgenus Melanoconion Theobald	
zoosophus Dyar & Knab	Mexico	abominator Dyar & Knab	Indigenous
Subgenus Stegomyia Theobald		anips Dyar	Mexico
aegypti (Linnaeus)	Cosmotropical	atratus Theobald	Neotropical
Genus ANOPHELES Meigen		erraticus (Dyar & Knab) iolambdis Dyar	Neotropical Neotropical
Subgenus Anopheles Meigen		mulrennam Basham	- Caribbean
atropos Dvar & Knab	Caribbean	opisthopus Komp	Neotropical
arrigari Triai & Bilair	Samona an	damdar maili	. историсы

Taxon	Extralimital Distribution	Taxon	Extralimital Distribution
peccator Dyar & Knab	Caribbean, Mexico	Genus PSOROPHORA Robineau-Desvoidy	
pilosus (Dyar & Knab)	Neotropical	Subgenus <i>Grabhamia</i> Theobald	
Subgenus Neoculex Dyar		columbiae (Dyar & Knab)	Caribbean,
apicalis Adams	Mexico	(25) 253	Mexico
arizonensis Bohart	Mexico	confinnis (Lynch Arribalzaga)	Neotropical
boharti Brookman & Reeves	Indigenous	discolor (Coquillett)	Mexico
reevesi Wirth	Mexico	pygmaea (Theobald)	Caribbean
territans Walker	Palearctic	signipennis (Coquillett)	Mexico
Subgenus Tinolestes Coquillett		Subgenus Janthinosoma Lynch	
latisquama (Coquillett)	Neotropical	Arribalzaga	
, , ,	1	cyanescens (Coquillett)	Neotropical
Genus CULISETA Felt		ferωx (von Humboldt)	Neotropical
Subgenus Climacura Howard,		horrida (Dyar & Knab)	Indigenous
Dyar & Knab		johnstonii (Grabham)	Caribbean
melanura (Coquillett)	Indigenous	longipalpus Randolph & O'Neill	Indigenous
Subgenus Culicella Felt		mathesoni Belkin & Heinemann	Indigenous
morsitans (Theobald)	Palearctic	mexicana (Bellardi)	Mexico
minnesotae Barr	Indigenous	varipes (Coquillett)	Neotropical
Subgenus <i>Culiseta</i> Felt		Subgenus <i>Psorophora</i> Robineau-Desvoidy	
alaskaensis (Ludlow)	Palearctic	ciliata (Fabricius)	Neotropical
impatiens (Walker)	Indigenous	howardii Coquillett	Neotropical
incidens (Thomson)	Mexico	·	•
inornata (Williston)	Mexico	Genus TOXORHYNCHITES	
particeps (Adams)	Neotropical	Theobald	
Z NEISZZZEBETE ZI I II	ļ	Subgenus Lynchiella Lahille	
Genus DEINOCERITES Theobald		rutilus rutilus (Coquillett)	Indigenous
cancer Theobald	Neotropical	rutilus septentrionalis (Dyar & Knab)	1di.marray
mathesoni Belkin & Hogue pseudes Dvar & Knab	Mexico Neotropical	· ·	Indigenous Indigenous
pseudes Dyar & Khan	Neotropical	sp.	ritalgelious
Genus HAEMAGOGUS Williston		Genus URANOTAENIA Lynch Arribalzaga	
Subgenus Haemagogus Williston		Subgenus Pseudoficalbia Theobald	
cquinus Theobald	Neotropical	anhydor anhydor Dyar	Mexico
ryamas Piksmans	Acoropical	anhydor syntheta Dyar & Shannon	Mexico
Genus MANSONIA Blanchard		Subgenus <i>Uranotaenia Ty</i> nch Arribalzaga	
Subgenus Mansonia Blanchard		lowii Theobald	Neotropical
<i>dyari</i> Belkin, Heinemann &		sapphirina (Osten Sacken)	Mexico
Page	Neotropical		W XICV
titillans (Walker)	Neotropical	Genus WYEOMYIA Theobald	
Genus OR I HOPODOMYIA	j	Subgenus Wycomym Theobald	
I heobald	1	haynei Dodge	Indigenous
alha Baker	Indigenous	mitchellii (Theobald)	Caribbean,
kummi Edwards	Neotropical	, , , , , , , , , , , , , , , , , , ,	Mexico
signifera (Coquillett)	Caribbean.	smithii (Coquillett)	Indigenous
	Mexico	canduzeer Dyar & Knab	Cambbean

In order for the user to have a better understanding of our position on certain taxa included herein, the following comments are offered.

Aedes—We recognize Ae. hemiteleus Dyar as distinct from Ae. cinereus Meigen which was proposed by Bohart and Washino (54). However, they point out that at present only the adult males can be differentiated with certainty; so in our keys the two species are grouped together. Peus (356) and Bohart and Washino (loc. cit.) have called attention to the presence of two subdorsal setae, in addition to setae la-S and 2-S on the siphon, not mentioned by Carpenter and LaCasse (106) nor in many other mosquito publications (e.g., 192, 212, 245, 298, 339, 350, 444). Their presence enables the larvae of these two species to be linked in the larval identification key with Ae. bicristatus and Ae. provocans, the other two North American aedines with two or more setae on the siphon. These two extra siphonal setae in cinereus and hemiteleus are very tiny and require a compound microscope with 400X magnification to see them clearly.

Only Ae, togoi (Theobald) is placed in the subgenus Finlaya here. This Asian disease vector has apparently been recently introduced into coastal British Columbia. Those species formerly assigned to subgenus Finlaya by Carpenter and LaCasse (106) were transferred to subgenus Protomacleaya, except for Ae, atropalpus and Ae, varipalpus, which were placed in subgenus Ochlerotatus; see Zavortink (514) and Arnell & Nielsen (9).

We agree with and so treat here the three new names of *Aedes* species proposed by Wood (504): *Ae. barri* Rueger = *Ae. enedes* Howard, Dyar & Knab, *Ae. trichurus* (Dyar) = *Ae. provocans* (Walker) and western North American populations of *Ae. stimulans* (Walker) = *Ae. mercurator* Dyar.

Ar. dorsalis (Meigen) was reduced to a subspecies of Ar. caspius (Pallas) by Gutsevich et al. (205) and so listed by Knight (518). The characters employed for separating the two taxa appear to us sufficient to retain dorsalis as a full species.

Following Nielsen and Rees (338) we recognize two subspecies under *Ae. spencerii* (Theobald), the typical subspecies which inhabits the central plains of North America, and subspecies *idahoensis* (Theobald), a more westerly and northwesterly form, also reported from southern British Columbia (135).

Coquillettidia—The elevation of the subgenus Coquillettidia Dyar to generic rank by Ronderos & Bachmann (389) has not been universally accepted, but we do recognize it here.

Culey—The taxonomic status of the important disease bearing and pestiferous taxa, Cx. pipiens Linnaeus and Cx. quinquefasciatus Say has been highly controversial. Cx. quinquefasciatus has been considered a subspecies of Cx. pipiens because the only reliable characters for separating them are structures of the male palpi and genitalia. Yet they do maintain themselves as recognizable taxa through their behavior and geographical distribution, although intergrades are known at least in parts of the USA where their ranges overlap (Barr, 14). We are adopting the positions of Sirivanakarn (524) and Knight (518) by considering them as separate, full species.

Culiseta Cs. minnesotae Barr was described in 1957 (15) and used by that name until 1964 when Maslov (295) reduced it to a subspecies of silvestris Shingarev. In this new status it appears to have been first used in North American literature by Siverly and DeFoliart (420) and subsequently in references 50, 121, 126, 198, 308, 352, 421, 519. Means & Thompson (198) referred to it simply as Cs. silvestris. In 1979 Wood et al. (505) presented good reasons for returning minnesotae to full specific rank. They pointed out that Maslov's decision was based on the examination of but a single male and that the validity of the name silvestris was in question. Therefore we are using here the name Cs. minnesotae Barr.

Likewise, the name for *Cs. morsitans* (Theobald) in North America was changed to *Cs. m. dyari* Coquillett by Maslov (295), and widely used that way in the literature of the region, e.g., 50, 68, 121, 140, 249, 301, 330, 352, 362, 468, 519, 529. Wood et al. (505) believed that this designation was unwarranted because the characters used to separate subsp. *morsitans* from subsp. *dyari* were inconsequential. Following him we have dropped the trinomiat and recognize only *Cs. morsitans* (Theobald).

Psorophora—Belkin et al. (34) proposed changing the name of the common pest, Ps. confinnis Exnch Arribalzaga, to Ps. columbiae (Dvar & Knab) and applied it to the populations in the eastern and southern USA. That left the name for the populations found in the southwestern states of New Mexico, Arizona and California in doubt. Subsequently, Bohart and Washino (54) have called the California "confinnis", Ps. columbiae. We are calling the Ps. confinnis of Carpenter and LaCasse (106), Ps. columbiae in all states except New Mexico and Arizona; these are simply designated as belonging to the Ps. confinnis complex.

Similarly, Ps. varipes (Coquillett) populations of southeastern USA have been renamed Ps. mathesoni Belkin and Heinemann (33). But these authors are uncertain about those occurring in central and southwestern USA. Since their geographical distribution extends over a contiguous area ranging from New Jersey to southern Illinois and from northern Florida to Oklahoma and Texas (see Plate 46), it appears more likely to constitute a single species, i.e., Ps. mathesoni, than if it were more widespread or discontinuous in its distribution. Therefore, it is so considered here, although Ps. varipes remains in our systematic index awaiting further study.

Toxorhynchites—Zavortink (512) reported finding a third taxon of this genus in southeastern Arizona. He believes it to be either Tx. theobaldi (Dyar & Knab) or its synonym, Tx. moctezuma (Dyar & Knab). He stated that it definitely is not one of the subspecies of Tx. rutilus (Coquillett), the common species of the region. We are listing it as Tx. sp. and not including it in the identification keys as specimens were not available for study.

MORPHOLOGY OF ADULT FEMALE

The morphological descriptions below deal mostly with the structures used in the keys. For a more detailed account of mosquito anatomy, consult the references listed in the bibliography at the end of this section.

Basic Structures

The body of the adult mosquito is composed of hardened plates, called **sclerites**, separated from each other by lines, known as **sutures**, or by membranes of various sizes. These structures comprise the integument, or outer covering of the body and those important in identification of the female will be discussed below.

Since scales are common on adult females and indeed constitute one of the principal structures of recognition, they must be distinguished from setae. Setae (hairs, hair tufts, bristles and spiniforms) are usually round in cross section, tapering from base to apex, and arise in a relatively large, movable socket, called an **alveolus** (pl. alveoli). Scales, on the other hand, are flat in cross-section, usually widening from base to apex, with longitudinal ridges, attached to minute alveoli on the integument. They occur in three basic forms, broad and flat, narrow and curved and erect and apically forked. The scales on the fringe of the mosquito wing are fusiform in shape (see Harbach and Knight, 1978C).

The color of scales varies from black and brown to golden, shades of yellow, such as dingy yellow in *Cx. salinarius*, to white and silvery. The white color can be brownish white, as in *Cs. minnesotae*, to grayish white. The colors tend to fade somewhat as the pinned adult ages, so in the keys herein, pale has been used to mean shades of white and dark, black or brown.

The body of the adult female is divided into three principal regions, the head, thorax and abdomen, Plate 1. Each will be discussed in detail.

HEAD

The structure of the head is shown in Plates 1, 2C. It is ovoid in shape and a large proportion is occupied by the **compound eyes** (CE). They are composed of circular, morphological units called **corneal facets** (CoF). The **antennae** (A) arise between the eyes. The sclerite ventrad to their bases is the convex **clypeus** (Clp). Dorsad is a sclerite between and above the antennae, the **frons** (Fr), above which is the dorsum of the head, made up of the **vertex** (V) anteriorly and the **occiput** (Occ) posteriorly. Since there is no dividing suture between them, it is customary to refer to the whole dorsum simply as the occiput. The anterior border along the dorsal edge of the compound eye is known as the **ocular line** (OL).

The head bears the following five appendages: two antennae, two palpi and the proboscis (Plate 2A, B). The two antennae are composed of a narrow, basal ring, the **scape** (Sc), the bulbous **pedicel** (=torus) (Pe), and the **flagellum** (Fl), which contains 13,14 **flagellomeres** (=flagellar segments) (Flm), each bearing a whorl of setae. A pair of **maxillary palpi** (MPlp), called simply palpi (sing. palpus), is located ventrolateral to the clypeus and each consists of five **palpomeres** (Plp); however, in some females the basal palpomere is small or rudimentary so that the palpi appear to be 4-segmented. The **proboscis** (P) extends forward from the anteroventral base of the head.

Normally, only the outer scaled covering of the proboscis, known as the **labium** (Lb), and the two terminal lobes, the **labella** (La) (sing. labellum), can be seen. Inside the labium are thin stylets for piercing the host's skin.

Nine characters of the head are used in the keys as follows: (1) Shape of proboscis—it is usually nearly straight, but in genus Toxorhynchites, it is decidedly curved downward (Fig. 1). (2) Scales on proboscis—sometimes the proboscis has a definite pale-scaled ring near the middle, as in Ae. sollicitans (Fig. 49), or it is variously marked with pale scales; however in most species it is dark-scaled throughout. (3) Length of palpi—this character is used to differentiate anopheline and culicine females. In the former, the palpi are as long as the proboscis while in the latter, they are not more than 0.4 as long as that organ. Within the culicine species, Ps. longipalpus (Fig. 493) has rather long palpi, i.e., more than 0.33 as long as the proboscis; and in some species of subgenus Neoculex, the length is compared to the length of flagellomere 4 of the antenna (Fig. 396). (4) Scales of palpi—apices of some or all of segments 2-5 may have pale-scaled rings, as in An. walkeri (Fig. 341), scattered pale among dark scales or only dark scales. (5) Scales on antennal pedicel—the numbers and color are diagnostic, e.g., Ae. fitchii (Fig. 111). (6) Length of antenna and flagellomere 1—flagellomere 1 is unusually long in genus Deinocerites (Fig. 39), and also the entire antenna is longer than the proboscis. (7) Width of frons—the width of the frons medially between the eyes, called the interocular distance, can be measured by comparing the distance with the diameter of a corneal facet, e.g., Ae. epactius (Fig. 153). (8) Interocular setae (1S)—they are located on the dorsal part of the frons and medioanterior area of the vertex and are long and usually dark, but in some species they are pale, e.g., An. freeborni (Fig. 339). (9) Scales on dorsum of head—posteriorly the scales are erect, usually forked, while anteriorly and laterally they are decumbent and either narrow and curved, e.g., subgenus Culex (Fig. 352) or broad and flat, e.g., subgenus Melanoconion (Fig. 354).

THORAX

The thorax (Plates 3,4), the body region between the head and abdomen, is divided into three segments, the prothorax, mesothorax, and metathorax. Each bears a pair of legs; in addition, the mesothorax has a pair of functional wings, and the metathorax, a pair of knobbed **halteres** (Hl). The dipterant mesothorax is typically greatly enlarged to accommodate the flight muscles associated with the mesothoracic wings. The pro-and metathorax are correspondingly reduced in size.

In dorsal view (Plate 3A,B) and proceeding from anterior to posterior, the **antepronota** (=anterior pronotal lobes) (Ap), parts of the prothorax, are found laterally just posterior to the head. The size and scalation of this structure are used in the keys. Two genera, *Haemogogus* and *Wycomyia*, have enlarged antepronota, approaching each other middorsally (Fig.31).

The next three structures are mesothoracic, starting with the **scutum** (Scu), the largest sclerite of the mosquito body and rather spheroid. The anterolateral depressions in the sphere are known as the **scutal fossae** (SF) and the slightly depressed, usually unscaled, area posteromedially, is the **prescutellar area** (PrA). The scutum has setae arranged in three, somewhat irregular, rows in the middle 0.33. The central one is composed of the acrostichal setae (AcS), and the row on either side, of the **dorsocentral setae** (DS). In addition, there is a group in front of and superior to the wing root, the supraalar setae (SaS). Those anterolateral setae occurring around and in the scutal fossa are the scutal fossal setae (SFS, Plates 3A, 4A). To some species the scutal setae are quite numerous and long, e.g., An. barberi (Fig. 334), while in others they are shorter and fewer. In the subgenus Melanoconion (Fig. 353) the acrostichal setae are absent, and in some species the acrostichal and dorsocentral setae are absent anteriorly, a condition which has been termed the "acrostichal gap" and the "dorsocentral gap" by Lunt and Nielsen (1971, p. 103). The color of some of these setae, particularly the supraalars, is diagnostic for several species, e.g., Ae. hexodontus (Fig. 305). The scutal integument may have spots or be a distinctive color e.g., reddish brown in Cx. crythrothorax (Fig. 371). The patterns made by the scutal scales are extensively employed in culicine mosquito identification, see Ar. atlanticus, Fig. 175, and usually have the same names as the setae just described when they occur in the same location. One difficulty commonly encountered is rubbed specimens in which the scutum is devoid of scales and setae. This is particularly true of those collected in mechanical light traps. By examining such specimens under the high power of the stereoscopic microscope the color of some few scales still attached may give a clue about the pattern of that species. Likewise the presence of alveoli will indicate the presence of setae in the specimen.

Posterior to the scutum is a transverse, linear sclerite, the **scutellum** (Stm). In the subfamily Anophelinae (Fig. 5) it is arcuate and bears an even row of setae, the **scutellar setae** (MSS, LSS). In the subfamily Culicinae the scutellum is trilobate with a group of setae on each lobe (Fig. 7). Also, the kind and color of scales and setae on this sclerite may be important.

The shiny, dome shaped structure posterior to the scutellum is the **mesopostnotum** (Mpn). In most species it is nude, but in the sabethine mosquitoes (Wyeomyia) a group of setae occurs near its attachment to the **metanotum** (Mtn) and **abdominal tergum I** (Ab-I) (Fig. 9), known as the **mesopostnotal setae** (MpnS).

Posteriorly is the **metanotum** (Mtn), a thin sclerite which enlarges laterally and there bears the halteres, the organs of balance. Next the intersegmental cleft separates the thorax from abdominal segment I, then there is a second, very thin, metathoracic element, the **metapostnotum** (Mtpn). It actually adheres to the first abdominal tergum, but extends lateroventrally as a thin strip to touch the metameron, see Plate 4A. The halteres are usually dark-scaled, but generally have pale scales in *An. walkeri* (Fig. 345).

The three thoracic segments are also represented in the structures of the thoracic pleuron, Plate 4A. Two of the sclerites visible laterally, the **antepronotum** (Ap) and the **postpronotum** (Ppn), are components of the tergum of the prothorax, not of its pleuron. Starting anteriorly, the prothoracic elements consist of the antepronotum (Ap) which is connected ventrally by a straplike piece to the **proepisternum** (Ps); both of these bear setae, i.e., **antepronotal setae** (ApS) and **upper proepisternal setae** (PeSU), and sometimes scales. The proepisternum bends around medially to cover the ventroanterior face of the thorax below the head and neck, see Plate 3A, and lobes from each side extend ventrally between the two forecoxae. This anterior face of the proepisternum is sometimes covered with scales, the **lower proepisternal scales** (PScl), e.g., Ae. hexodontus (Fig. 296). The last prothoracic sclerite, the postpronotum (Ppn) is found posterior to the antepronotum and lateral to the scutum at the level of the scutal fossa. It bears scales, which sometimes have a distinctive pattern; and a number of setae (PpS), usually confined to the posterior margin, but sometimes scattered over the posterior 0.5, e.g., Ae. impiger (Fig. 284).

The mesothoracic pleuron has five, large and important sclerites. Just posterior to the postpronotum is an opening in the thorax. This is the mesothoracic spiracle (MS) and it is surrounded by a large sclerite, the anterior mesanepisternum (AMas). It is subdivided into four areas: (1) The **prespiracular area** (PsA) is the small triangle dorsoanterior to the spiracle. It adjoins the posterior border of the postpronotum, and sometimes bears setae, the prespiracular setae (PsS), e.g., genus Culiseta (Fig. 18), (2) The postspiracular area (PA) is a rather large expanse posterior to the spiracle with or without setae and scales; when present, they are the **postspiracular setae** (PS), e.g., genus Psorophora (Fig. 17), and postspiracular scales (PoSc), e.g., Ac, brelandi (Fig. 186), (3) The hypostigmal area (HvA) is immediately ventral to the spiracle and at times has scales, the hypostigmal scales (HySc), e.g., Ae. pullatus (Fig. 227), or a dark integumental spot, as in Ae. fulcus pallens (Fig. 169), (4) The subspiracular area (SA) is that portion ventral to the hypostigmal area, adjoining the meskatepisternum ventrally, with or without subspiracular setae (SuS) and scales (SSc), e.g., Ar. varipalpus (Fig. 161). The largest of the mesopleural sclerites, the **mesokatepisternum** (= sternopleuron) (Mks) is rather pear-shaped, bulging ventroanteriorly. It is united with a dorsal, narrow, linear area, the **posterior mesanepisternum** (PMas), which bears a dense group of setae. the prealar setae (PaS). The meskatepisternum has two groups of setae, the upper (MkSU) and **lower** (MkSL) mesokatepisternal setae. These are often combined into a single line of setae, the mesokatepisternal setae (MkS). The mesokatepisternal scales (MkSc) are sometimes arranged in distinctive patterns, e.g., narrow lines of scales, as in Ac. papago (Fig. 64), or more frequently an extensive scale patch which may or may not reach the anterior angle, as in Ar. prococurs (Fig. 239). Between the forecoxa and the ventroanterior border of the meskatepisternum there is a membrane, the **postprocoxal membrane** (PM). In some species of Acides it bears a small patch of scales the postprocoxal scales (PSc) e.g., Ar. punctor (Fig. 254).

The rectangular sclerite just posterior to the meskatepisternum and ventral to the root of the wing (W) is the **mesanepimeron** (Mam). It bears a group of setac in the dorsoposterior corner, the **upper mesanepimeral setae** (MeSU). Sometimes, another group, usually with not more than 1-6 setac in a single row occurs along the anteroventral border, the **lower mesanepimeral setae** (MeSL). They are often used to separate groups of species in the genus *Acdes*, e.g., *Te. riparus* (Fig. 91) from *Ae. stimulans* (Fig. 92). It may also have varying amounts of scaling. In some species of the subgenus *Melanoconion* the mesanepimeron has a definite pale spot or light- and dark-colored

integumental areas, which provide specific differentiation (Fig. 408). Just ventral to the mesanepimeron is the fifth and smallest, mesopleural sclerite, the **mesomeron** (Msm). It is triangular and is situated between mid - (C-II) and hindcoxae (C-III). The relation of the base of the mesomeron to the base of the hindcoxa is a generic character. Usually the base of the hindcoxa is distinctly ventral to the base of the mesomeron, but in the sabethine females the base of the hindcoxa is about even with the base of the mesomeron, see Figs. 10, 12.

The metathoracic pleuron is much reduced, (Plate 4A). The largest element is the **metepisternum** (Mts) and is located just posterior to the mesanepimeron. It is strapshaped, has dorsoventral axis and surrounds in its dorsal half the **metathoracic spiracle** (MtS), the other opening in the thorax. Posteriorly below the halter is the **metepimeron**, (Mtm), another narrow sclerite. Ventral to the metepisternum is a small sclerite, the **metameron** (Mem) articulating with the hindcoxa posteriorly and with the ventroposterior border of the mesanepimeron. Rarely it bears scales (see Fig. 265). Dorsoposterior to the metepimeron is the metanotum, already discussed.

The sternal elements of the thorax are not included in this discussion since they have not been used as identifying characters, except for one, the intersegmental membrane connecting the metasternum with abdominal sternum I. It sometimes bears **postmetasternal scales** (MScP), e.g., *Ar. pionips* (Fig. 304).

APPENDAGES OF THE THORAX

Wings- The two functional wings (W) of adult mosquitoes are attached to the mesothorax, see Plate 3C. Each is composed of a network of longitudinal thickenings, called **veins**. Between the veins are stretched transparent membranes, known as **cells**. The veins are clothed with scales dorsally and ventrally. The apical and posterior margin of the wing is bordered by long, fusiform scales, the **wing fringe** (FS). It may have pale and dark sections, best exemplified in *Ps. signipennis* (Fig. 465), or there may be a coppery or silvery, apical spot, e.g., *An. varlei* (Fig. 314).

The veins and cells have names, as shown in Plate 3C. The system of nomenclature used here is the Comstock-Needham system. There are six major longitudinal veins, i.e., costa (C), subcosta (Sc), radius (R), media (M), cubitus (Cu) and anal (A). If the veins are traced from base to apex, several of them have one or more subdivisions. For example, the radius has the basal vein R, with primary branches, R_1 and radial sector R_s . The latter further divides into R_{2+3} and R_{4+5} . The R_{2+3} separates into R_2 and R_3 apically. There are several crossveins, short connectors between major veins. The humeral crossvein (h) joins the costa with the subcosta, the radiomedial crossvein (r-m), the radius with the media, and the mediocubital crossvein (m-cu), the media with the cubital veins.

The cells likewise have names, per Plate 3C (letters in italics). An important one to know is cell R_2 because it is shortened in the genus Uranotaenia (Fig. 13). In the key character its length is compared to the length of the vein R_{2+3} , a short portion of vein R_s between the branching of R_{4+5} and the junction of veins R_g and R_g . This section of vein is called the "petiole" by some authors.

The wing scales provide many useful key characters. They can be broad and numerous, e.g., Cq. perturbans (Fig. 37), triangular shaped, e.g., Ae. grossbecki (Fig. 77), or narrow and filiform, e.g., Cx. pipiens (Fig. L5). Colors are important, too. Many species have the wing scales entirely dark, or they may vary in number of pale scales from a small patch at the base of the costa, e.g., Ae. atropalpus (Fig. 127), to scattered pale scales on the anterior veins, e.g., Ae. cataphylla (Fig. 221), to generally intermixed pale and dark scales, e.g., Ae. solludians (Fig. 54), to alternating mostly dark with mostly pale-scaled veins, e.g., Ae. s. alahoensis (Fig. 207), to mainly pale-scaled, e.g., Ae. dorsalis (Fig. 125). Furthermore, there are wings with unicolorous spots produced by the occurrence of dense clusters of scales along some veins, e.g., An. quadrimaculatus (Fig. 313). The costa, subcosta and radial veins in some anophelines possess spots of pale scales which are named. The area of pale scales at or near the apex of the wing is called the apical spot, and the subcostal spot is found where the subcostal vein joins the costal vein. Although they are called "spots," they are really patches of pale scales extending over several veins, e.g., An. punctipennis (Fig. 318). Most mosquito wings do not bear prominent setae, but in the genus Culiscia (Fig. 28), a row occurs ventrally near the base of the subcosta.

Legs- There are 3 pairs of legs, one attached to each thoracic segment. The leg consists of five main parts: **coxa** (C-I, C-III, C-III), **trochanter** (11), **femur** (Fe), **tibia** (Ti) and **tarsus** (Ta): Plate 2D. The tarsus is composed of five segments, known as **tarsomeres**. The fifth tarsomere (Ta₅)

bears two **claws** (= unguis, U) (Cl) which, in most species, have a secondary element, the tooth. The tarsal claws are used frequently in the *Aedes* key, e.g., *Ae. excrucians* (Fig. 89). They can best be studied under the stereoscopic microscope by shining the light on the stage below the specimen and viewing the claws in silhouette. Tarsomere $4(Ta_4)$ is unusually small in the fore- and midlegs of the genus *Orthopodomyia* (Fig. 34).

Scale patterns on the various segments of the legs are extensively employed as key characters. The scales on coxa 1 can be brown or pale, e.g., Ae. cinereus (Fig. 261). The femora may have the basal half all pale, e.g., Ae. zoosophus (Fig. 69); or with subapical pale rings, e.g., Ps. columbiae (Fig. 456); or with apical pale rings (=knee spots), e.g., Ae. implicatus (Fig. 238). The foretibiae sometimes bear a line of pale scales, e.g., Cx. tarsalis (Fig. 363). The femora and tibiae of some Psorophora species have long, erect scales apically, giving them a shaggy appearance (Fig. 467). The tarsomeres, especially on the hindleg, may have basal, pale rings, which are narrow, as in Ae. vexans (Fig. 71), or broad as in Ae. excrucians (Fig. 45), both apical and basal pale rings, as in, Ae. canadensis (Fig. 48), or with tarsomeres 4, 5 and part of 3 all pale, as in Ps. Jerox (Fig. 469).

ABDOMEN

The abdomen is composed of 10 segments, of which the first seven are quite similar in external structure. The three terminal segments are specialized for reproduction and excretion. It has become customary to refer to the abdominal segments by Roman numerals, e.g., abdominal segment 111.

Each of the first seven segments has a dorsal sclerite, the **tergum** (Te) and a ventral sclerite, the **sternum** (S); see Plate 4B. Laterally, they are connected by expandable, elastic tissue, the **pleural membrane** (PMe). A similar intersegmental membrane separates the terga dorsally and the sterna ventrally. These membranes permit the abdomen to distend during blood feeding and when the female becomes gravid.

Segments VIII-X are shortened and modified. In some genera, e.g., Culex, Culexta and Mansonia (Fig. 19), these segments are mostly telescoped inside the terminal segments making the apex of the abdomen appear bluntly rounded. In other genera, e.g., Irdes (Fig. 21) and Psorophora, parts of these segments protrude posteriorly, giving the abdominal terminus a pointed appearance. Also in those with blunt abdomens, segment VII is almost the same width as VI, but in the pointed abdomens, VII is decidedly smaller than VI. Abdominal segment VIII usually has a larger sternum than tergum. Posterior to tergum VIII can be seen two elongated lobes, the **cerci** (sing, cercus). These structures are long, straight and visible in the genera with pointed abdomens, but are shorter, usually curved medially and not so visible in the genera with blunt abdomens. Ventrally, posterior to sternum VIII is a smaller lobe lying ventral to the cerci, the **postgenital lobe** (PGL). Both of these terminal organs are parts of the female genitalia.

No attempt will be made to describe completely the female genitalia, since their parts are rarely used in the key, but some elements are described above because they should be recognized. For an account of the female genitalia, consult Laffoon and Knight (1971) and Reinert (1974).

The anopheline abdomen, with the exception of An. albimanus, is devoid of scales although it bears a number of tergal and sternal setae. In the other genera, both setae and scales are present on the abdomen. The patterns of dark and pale scales are very important in identification. Sometimes the pale scales are located basally on the tergum, i.e., on the part nearest the base of the abdomen, where it is attached to the thorax, e.g., Ae. intrudeus (Fig. 242), and sometimes on the apical part, i.e., nearest the free distal end of the abdomen, e.g., Cx. territans (Fig. 356). Likewise, the scales on the sterna may be unicolorous or have distinctive patterns, e.g., Cx. tarselis (Fig. 364). In some cases it is necessary to distinguish shades of the pale scales, for example, the pale band on hindrarsomere 1 m. In solluctans is vellow-scaled, while in Ae. ingromaculis, when present, is white-scaled; see Figs. 59, 61. In Mansonia there are special spiniforms on the posterior border of tergum VII in Ma. titillans (Fig. 441), and thick, peglike spiniforms on tergum VIII of all species, the cerci of December (Fig. 439) have specialized spatulate setae.

The following list indicates the changes made in names of adult structures in this publication, adopted from the mosquito taxonomic glossary; see Knight (1970), Knight and Laffoon (1970B, 1970C, 1971A) and Harbach and Knight (1980).

Old Name

anterior pronotum

flagellar segment

meron

mesane pister num

mesepimeron

mesonotum

ommitidium palpal segment

postcoxal area

postnotum

prealar area

propleuron

prosternum

sternopleuron, mesepisternum

tarsal segment

New Name

antepronotum

flagellomere

mesomeron

anterior mesanepisternum

mesanepimeron

scutum

corneal facet

palpomere

postprocoxal area

mesopostnotum

posterior mesanepisternum

proepisternum

anterior part of proepisternum

mesokatepisternum

tarsomere

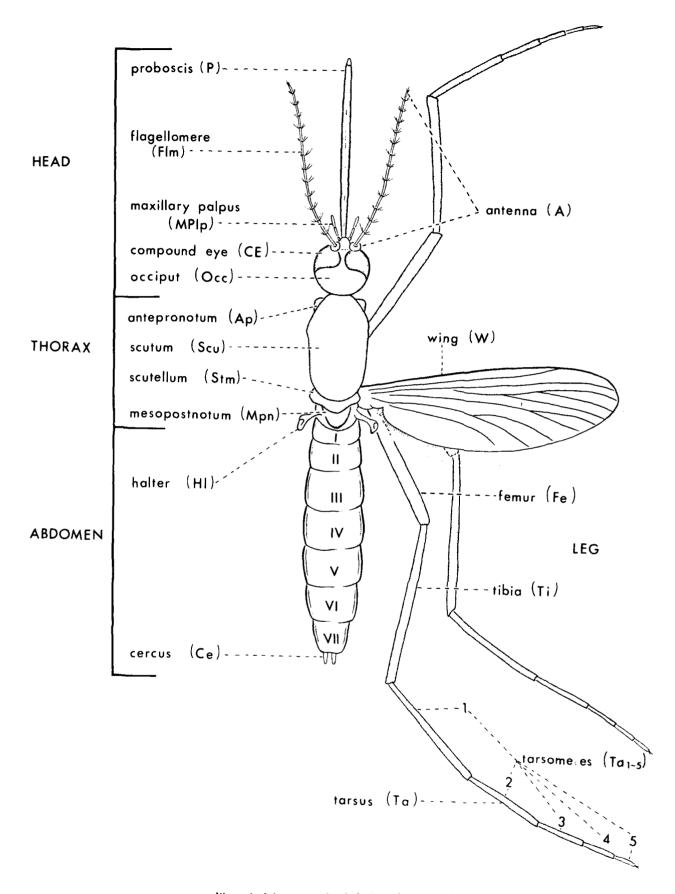


Plate 1. Diagram of adult female mosquito.

ABBREVIATIONS OF ADULT FEMALE MORPHOLOGY IN PLATES

Plate 2

Lb - labium A - antenna C - coxa MPlp - maxillary palpus CE - compound eve Occ - occiput OL - ocular line Cl - claw P - proboscis Clp - clypeus CoF - corneal facet Pe - pedicel Fe - femur Plp - palpomere Sc - scape Fl - flagellum Ta - tarsus Flm - flagellomere Fr - frons Ta_{1.5} - tarsomere

> Ti - tibia Tr - trochanter V - vertex

> > W - wing.

Plate 3

Illustrations A and B

La - labellum

IS - interocular space

AcS - acrostichal setae PeSU - upper proepisternal setae Ap - antepronotum Ppn - postpronotum ApS - antepronotal setae PpS - postpronotal setae C-I - forecoxa PrA - prescutellar area Cv - cervix Ps - proepisternum DS - dorsocentral setae SaS - supraalar setae LSS - lateral scutellar setae Scu - scutum Mpn - mesopostnotum SF - scutal fossa SFS - scutal fossal setae MSS - median scutellar setae Stm - scutellum

Mtn - metanotum

Illustration C (Wing)

 M_{1+2} - anterior branch of A - anal vein A - anal cell medial vein C - costal vein M_2 - medial₂ cell C - Costal cell M_{3+4} - posterior branch Cu - cubital vein of medial vein Cu₁ - anterior branch of M_4 - medial₄ cell

cubital vein m-cu - mediocubital crossvein

 Cu_i - cubital cell R - radial vein Cu2 - posterior branch of R - radial cell

cubital vein \mathbf{R}_1 - anteriormost branch of Gu₂ - cubital₃ cell radial vein FS - fringe scales

 R_i - radial, cell h - humeral crossvein R_s - radial sector vein M - medial vein R₂ - anterior branch of M - medial cell radial sector vein

 R_2 - radial₂ cell

 R_{2+3} - connector vein (stem) of radial sector vein

Ra - median branch of radial sector vein

 R_3 - radial₃ cell

R₄₊₅ - posterior branch of radial sector vein

R, - radial₅ cell

r-m - radiomedial crossvein

Sc - subcostal vein

Sc - subcostal cell

Plate 4

Ab-I - abdominal segment I

AMas - anterior mesanepisternum

Ap - antepronotum

ApS - antepronotal setae

C-I - forecoxa

C-II - midcoxa

C-III - hindcoxa

Ce - cercus

Cv - cervix

DS - dorsocentral setae

H- head

Hl - halter

HvA - hypostigmal area

LSS - lateral scutellar setae

Mam - mesanepimeron

Mem - metameron

MeSL - lower mesanepimeral setae

MeSU - upper mesanepimeral setae

Mks - mesokatepisternum

MkSL - lower mesokatepisternal setae

MkSU - upper mesokatepisternal setae

Mpn - mesopostnotum

MS - mesothoracic spiracle

Msm - mesomeron

MSS - medial scutellar setae

Mtm - metepimeron

Mtn - metanotum

Mtpn - metapostnotum

Mts - metepisternum

MtS - metathoracic spiracle

PA - postspiracular area

PaS - prealar setae

PeSU - upper proepisternal setae

PGL - postgenital lobe

PM - postprocoxal membrane

PMas - posterior mesanepisternum

Ppn - postpronotum

PpS - postpronotal setae

Ps - proepisternum

PS - postspiracular setae

PsS - prespiracular setae

PsA - prespiracular area

S - sternum of abdomen

SA - subspiracular area

SaS - supraalar setae

Scu - scutum

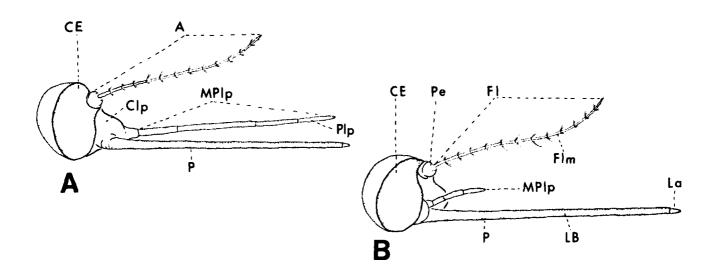
SF- scutal fossa

SFS - scutal fossal setae

Stm - scutellum

Te - tergum of abdomen

W - wing



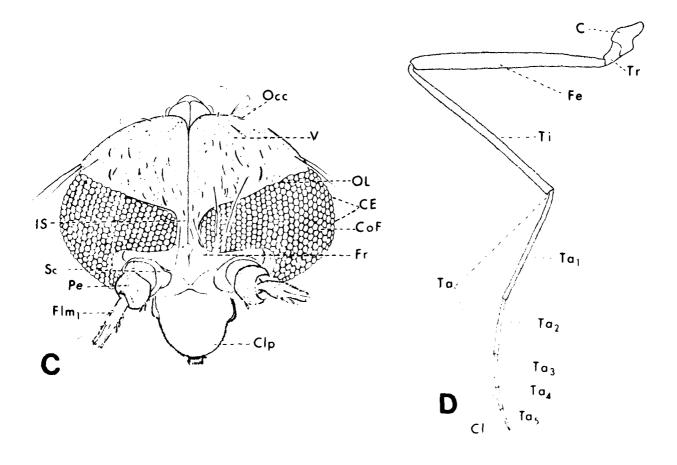
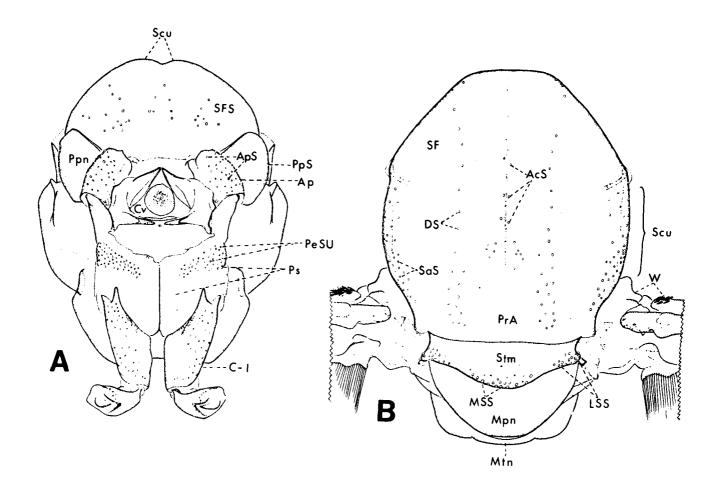


Plate 2. Head and leg of adult female mosquito. A Tateral view of an spheline head, B. Tateral view of culicine head; C. dorsal view of culicine head; D. lateral view of leg.



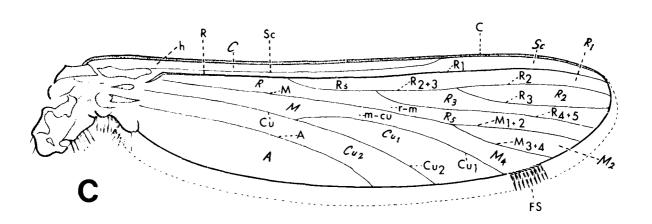
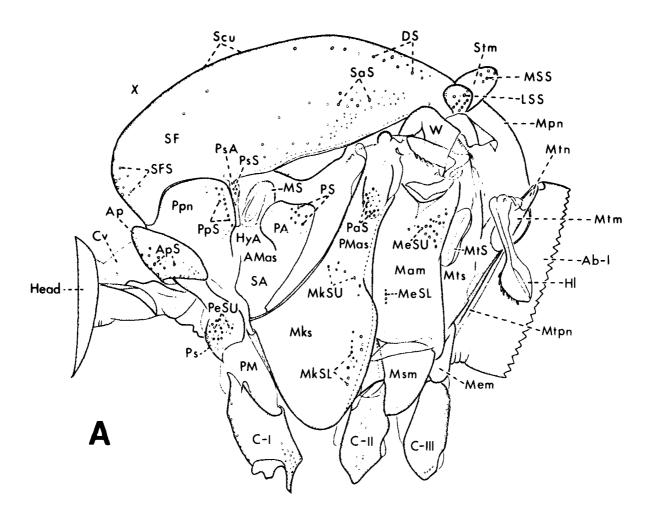


Plate 3. Thorax and wing of adult female mosquito. A. Anterior view of thorax; B. Dorsal view of thorax; C. Dorsal view of wing: longitudinal veins designated by gothic letters, cells by italics.



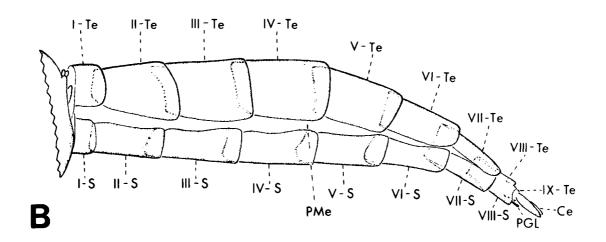


Plate 4. Thorax and abdomen of adult female mosquito. A. Lateral view of thorax; B. Lateral view of abdomen.

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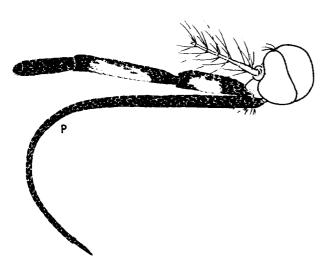
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KEY TO GENERA OF ADULT FEMALE MOSQUITOES OF NORTH AMERICA, NORTH OF MEXICO

or only slightly emarginated at apex of vein Cu₂ (Fig. 4) 2



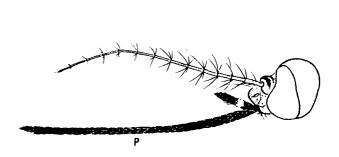


Fig. 1 — Lateral view of head - Tx. r. septentrionalis

Fig. 3 — Lateral view of head - Ae, vexans

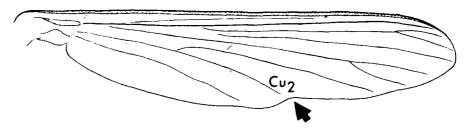


Fig. 2 — Dorsal view of wing - Tx, r, septentrionalis

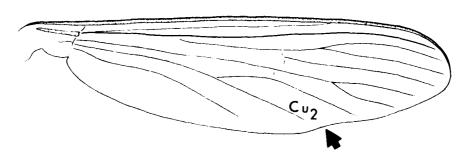


Fig. 4 - Dorsal view of wing - Ac. vexans

^{*}Refer to Plates containing maps which portray geographical distribution

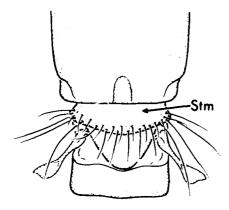


Fig. 5 — Posterior dorsal view of thorax - An, quadrimaculatus

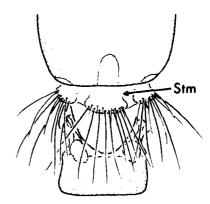


Fig. 7 — Posterior dorsal view of thorax - Ae. vexans

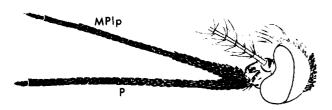


Fig. 6 — Lateral view of head - An. quadrimaculatus

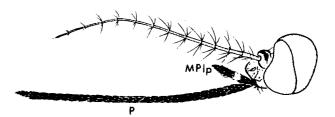


Fig. 8 — Lateral view of head - Ae, vexans

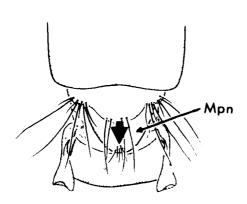


Fig. 9 — Posterior dorsal view of thorax - Wy, smithir

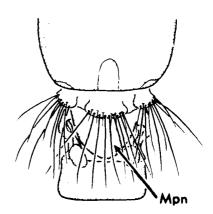
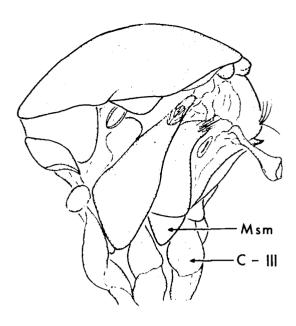


Fig. 11 — Posterior dorsal view of thorax - Ae, vexans



Msm C-III

Fig. 10 — Lateral view of thorax - Wy, smithil

Fig. 12 — Lateral view of thorax - Ae, vexans

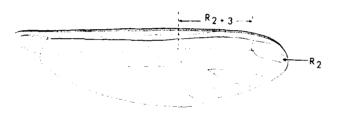


Fig. 13 — Dorsal view of wing - Ur. sapphirma

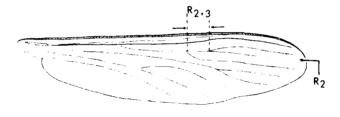


Fig. 15 — Dorsal view of wing - Cx. pipiens

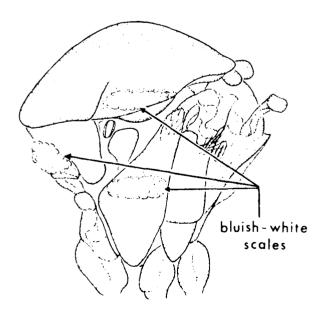
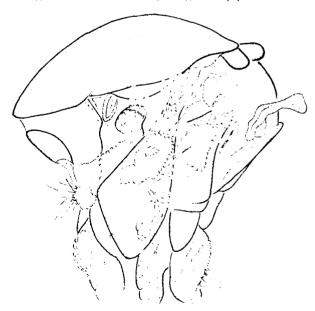
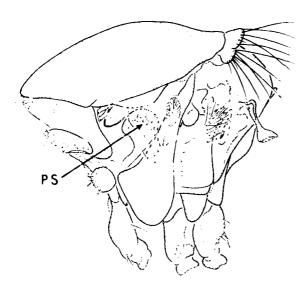


Fig. 14 Lateral view of thorax Assurphirms



Aug. 16 Lateral en a of thorax. Ar. cexans.

5(4).	Ostspiracular setae present (Fig. 17)	• • •
	Postspiracular setae absent (Fig. 18)	:



PA

Fig. 17 — Lateral view of thorax - Ps. ciliata

Fig. 18 —Lateral view of thorax - Cs. inornata

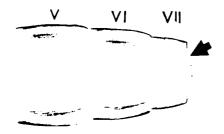


Fig. 19 — Dorsal view of abdomen - Ma, titillans

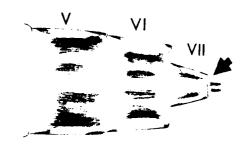


Fig. 21 — Dorsal view of abdomen - Ae. vexans



Fig. 20 - Dorsal curw of some cerns. Ma. titillans.



Fig. 22 - Dorsal view of some verns Ae, vexans

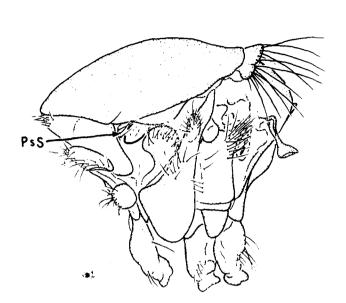


Fig. 23 - Lateral view of thorax - Ps. ciliata

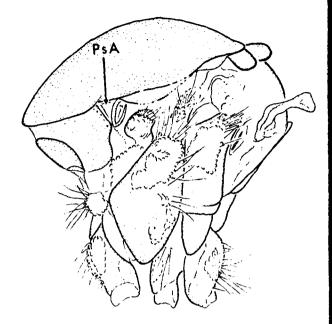


Fig. 25 — Lateral view of thorax - Ac. vexans

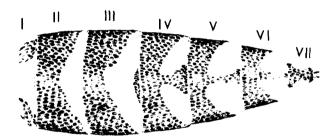


Fig. 24 — Desa' view of abdomen - Ps. cyanescens

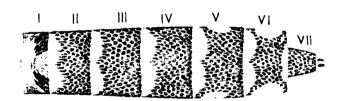


Fig. 26 — Dorsal view of abdomen - Ae. vexans

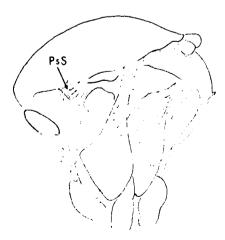


Fig. 27 — Lateral view of thorax - Cs. inornata

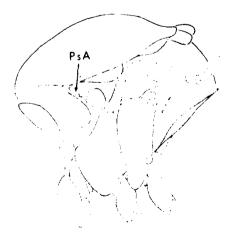
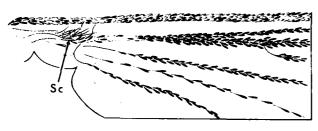


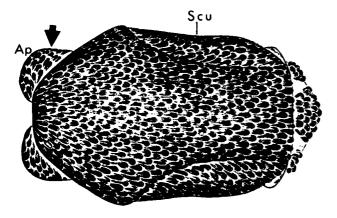
Fig. 29 -- Lateral view of thorax - Cx. pipiens



S c

Fig. 28 - Ventral view of basal half of wing - Cs. inornata

Fig. 30 - Ventral view of basal half of wing - Cx. pipiens



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Fig. 31 — Dorsal view of thorax - Hg. equinus

Fig. 32 — Dorsal view of thorax - Cx. pipiens

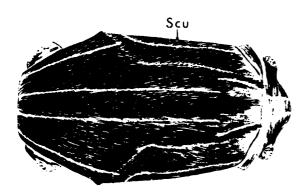


Fig. 33 — Dorsal view of thorax - Or, signifera

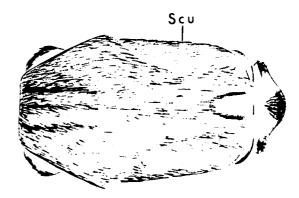


Fig. 35 — Dorsal view of thorax - Cx. pipiens

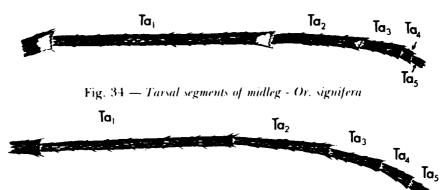


Fig. 36 — Tarsal segments of midleg - Cx. pipiens



Fig. 37 — Dorsal view of wing - Cq. perturbans



Fig. 38 — Dorsal view of wing - Cx, pipiens

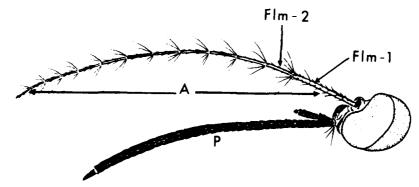


Fig. 39 - Lateral view of head - De. pseudes

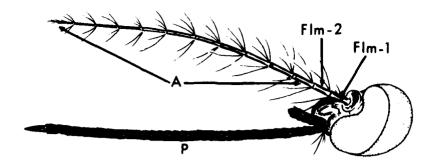


Fig. 40 — Lateral view of head - Cx. pipiens

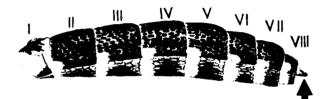


Fig. 41 — Lateral view of abdomen - Ae, purprieipes



Fig. 43 — Lateral view of abdomen - Cx, pipiens

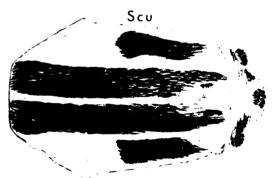


Fig. 42 - Dorsal ciew of thorax Ac. purpurcipes

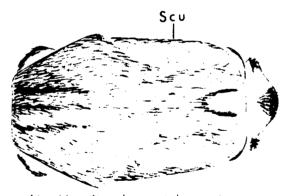


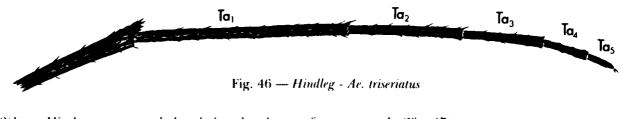
Fig. 44 - Dorsal view of thorax Cx. pipiens

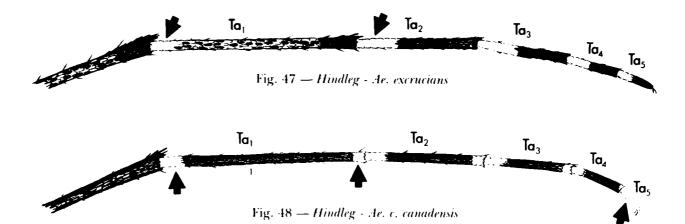
KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS AEDES

 1. Hindtarsomere with pale bands (Fig. 45)
 2

 Hindtarsomere without pale bands (Fig. 46)
 36







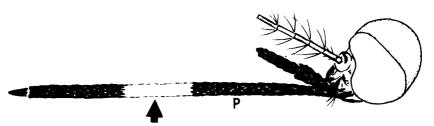


Fig. 49 — Lateral view of head - Ae. sollicitans

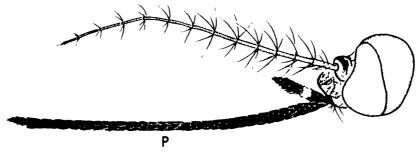
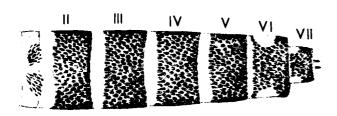


Fig. 50 - Lateral view of head Ac, vexans



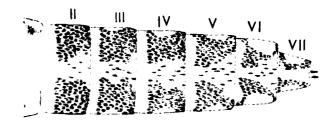


Fig. 51 — Dorsal view of abdomen - Ae, taeniorhynchus

Fig. 53 - Dorsal view of abdomen - Ae. sollicitans

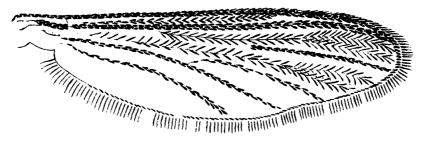


Fig. 52 — Dorsal view of wing- Ae. taeniorhynchus

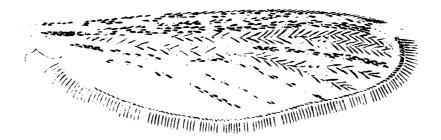


Fig. 54 — Dorsal view of wing - Ae. sollicitans



Fig. 55 - Dorsal view of wing - Ac. mitchellae

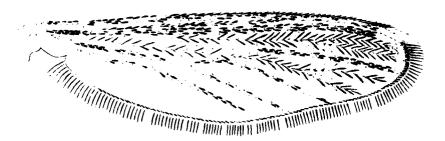
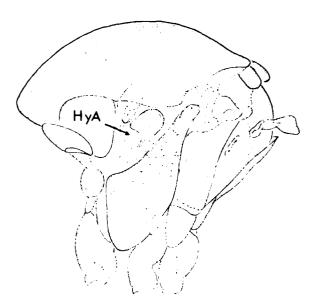


Fig. 57 — Dorsal view of wing - Ae. sollicitans



НуА

Fig. 56 - Lateral view of thorax - Ae. mitchellae

Fig. 58 - Lateral view of thorax - Ae, sollicitans

Hindtarsomere I usually without median, pale band, if present, then scales whitish (Fig. 61); basolateral patches on abdominal terga vellowish-scaled (Fig. 62) (in part) nigromaculis (Plate 25)



Fig. 59 — Hindtarus - Ac. sollicitans



Fig. 61 — Hindtarus - Ac. nigromaculis

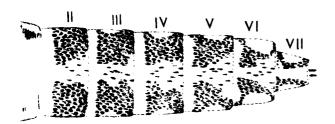


Fig. 60 - Dorsal view of abdomen - Ae. sollicitans

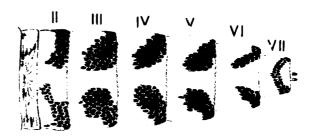


Fig. 62 - Dorsal view of abdomen - Ae. nigromaculis

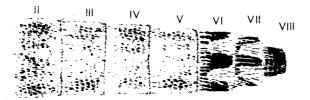


Fig. 63 — Dorsal view of abdomen - Ae. papago

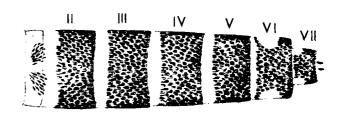


Fig. 65 - Dorsal view of abdomen - Ae. taeniorhynchus

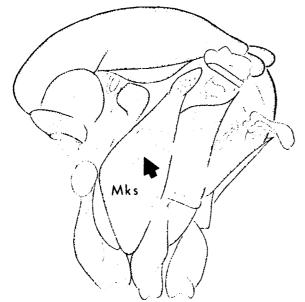


Fig. 64 — Lateral view of thorax - Ae. papago

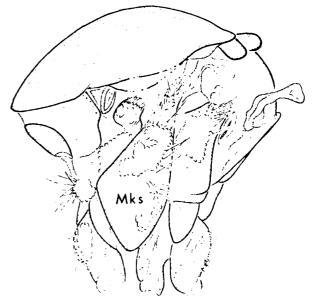
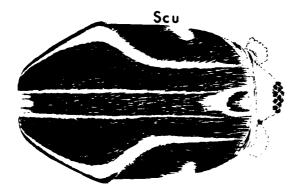


Fig. 66 — Lateral view of thorax Ac. vexans



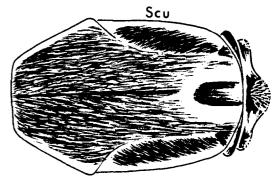


Fig. 67 — Dorsal view of thorax - Av. aegypti

Fig. 68 — Dorsal view of thorax - Ae. c. canadensis



Fig. 69 — Hindleg - Ac. zoosophus

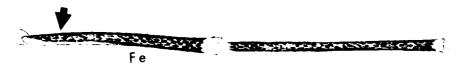
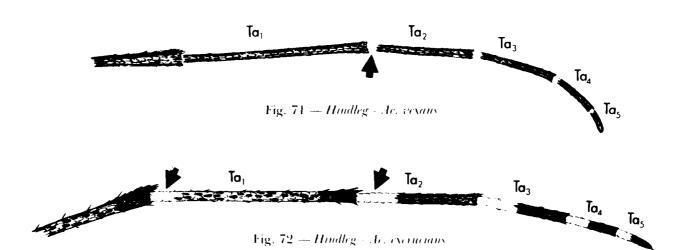
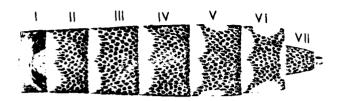


Fig. 70 — Hindleg - Ac. epactius



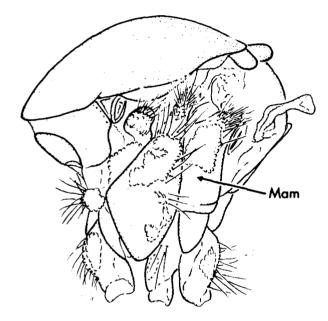
11(10).	Basal pale bands on abdominal terga II-VI with 2 posterior lobes, tergum VII mostly
	dark-scaled (Fig. 73); lower mesanepimeral setae absent (Fig. 74)vexans
	(Plate 26)
	Basal pale bands on terga II-VI not bilobed nor clearly defined, tergum VII mostly
	pale-scaled (Fig. 75); lower mesanepimeral setae present (Fig. 76)
	(Plate 19)



II IV VIII

Fig. 73 — Dorsal view of abdomen - Ae. vexans

Fig. 75 — Dorsal view of abdomen - Ae. cantator



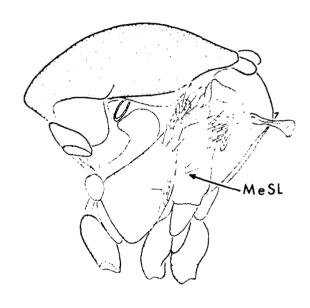


Fig. 74 — Lateral view of thorax - Ac. vexans

Fig. 76 - Lateral view of thorax - Ae, cantator

12(10)	. Wing with broad, triangular-shaped, dark and pale scales rather evenly intermixed
	dorsally (Fig. 77)
	At least some dorsal wing scales narrow, with dark and pale scales usually unevenly
	distributed (Fig. 78)

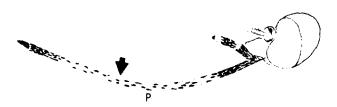


Fig. 77 — Dorsal view of wing - Ae, grossbeeki



Fig. 78 - Dorsal view of wing - Ae, stimulans

(3(12)). Proboseis with many dark and pale scales intermixed (Fig. 79); scutum with mixed brown and pale scales laterally (Fig. 80)	ger
(Plate)	
Proboscis with few scattered pale scales on basal 0.5 (Fig. 81); scutum with mostly pale	
scales laterally (Fig. 82)	ki
(Plate 2	22)



C

Fig. 79 Lateral crew of head Ar. squamger

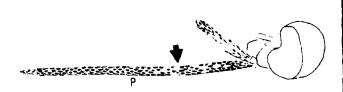


Fig. 81 --- Lateral view of head - Ac. grossbecki

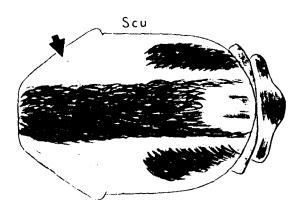


Fig. 80 Dorval chew of thorax Act squamiger

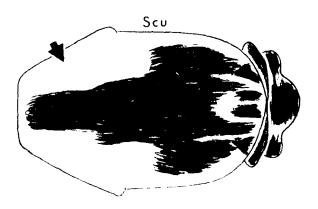


Fig. 82 Dorsal on at there's to a school.

Palpus with some pale scales (Fig. 85); pale scales of abdominal terga never forming distinct and complete, median, longitudinal stripe (Fig. 86).

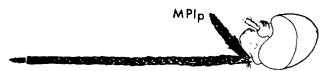
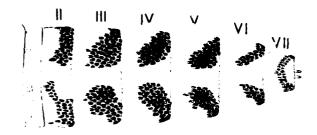




Fig. 83 — Lateral view of head - Ae, nigromaculis

Fig. 85 — Lateral view of head - Ae. increpitus



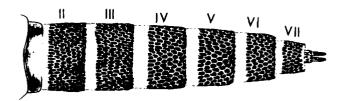


Fig. 84 — Dorsal view of abdomen - Ae. nigromaculis

Fig. 86 — Dorsal view of abdomen - Ae, increpitus

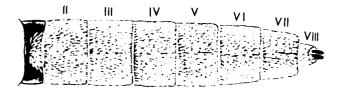
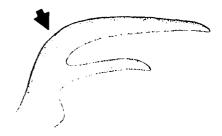




Fig. 87 — Dorsal view of abdomen - Ae. flavescens

Fig. 88 — Dorsal view of abdomen - Ac, increpitus



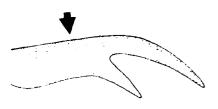


Fig. 89 Forcelase Acsexerneurs

Fig. 90 - Forcelas - 1c. increpitus

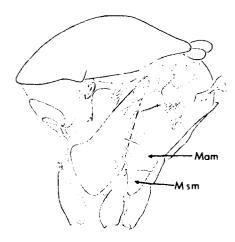




Fig. 91 — Lateral view of thorax - Ae. riparius

Fig. 92 — Lateral view of thorax - Ae. stimulans

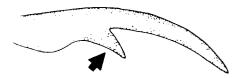


Fig. 93 - Foreclaw - Ae. riparius

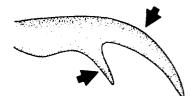


Fig. 94 — Foreclaw - Ae. fitchii

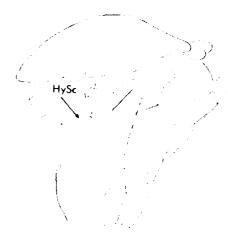


Fig. 95 - Lateral view of thorax Acriparius

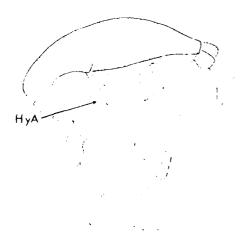


Fig. 97 — Lateral view of thorax - Ac, aloponotum

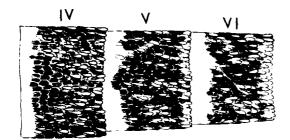


Fig. 96 — Dorsal view of abdominal segments IV-VI - Ae. riparius

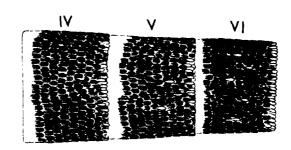


Fig. 98 — Dorsal view of abdominal segments IV-VI - Ae. aloponotum

20(18). Proboscis, cercus and tarsomere 1 of all legs with numerous pale scales (Figs. 99, 100, 101); foreclaw long, straight distal to attachment of tooth (Fig. 102) (in part) euedes (Plate 15)

Proboscis, cercus and tarsomere 1, distal to basal ring, usually dark-scaled (Figs. 103, 104, 105); foreclaw shorter and more strongly curved distal to attachment of tooth (Fig. 106) . (in part) fitchii (Plate 16)

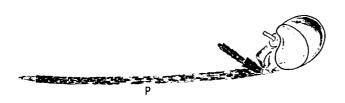


Fig. 99 — Lateral view of head - Ae. enedes

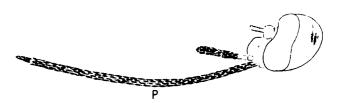


Fig. 103 — Lateral view of head - Ae, fitchii

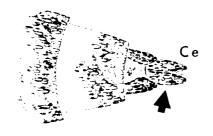


Fig. 100—Dorsal www of abdominal segments VII - X - Acenede



Fig. 104 — Dorsal view of abdominal segments VII - X - Ae.

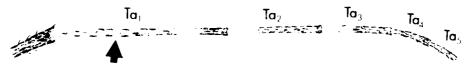


Fig. 101 - Lateral view of hindleg - Ac, eucles

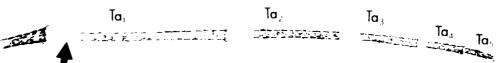


Fig. 105 -- Lateral crew of hindleg - Ac, fitchii

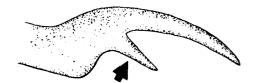


Fig. 102 — Foreclaw - Ae. euedes

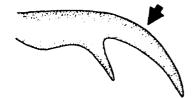


Fig. 106 — Foreclaw - Ae. fitchii

21(17). Segments 2,3 of palpus dark-scaled with apical pale-scaled rings (Fig. 107); abdominal sterna IV,V with lateral patches of dark scales (Fig. 108); proboscis dark-scaled (Fig. 107) increpitus (Plate 14)

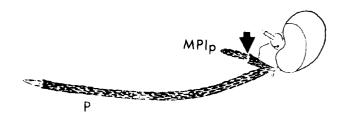


Fig. 107 — Lateral view of head - Ae. increpitus

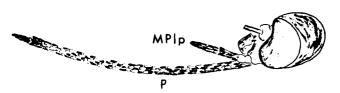


Fig. 109 - Lateral view of head - Ae. stimulans



Fig. 108 — Ventral view of abdomen - Ae, increpitus

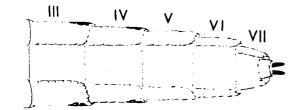


Fig. 110 - Ventral view of abdomen - Ae. stimulans

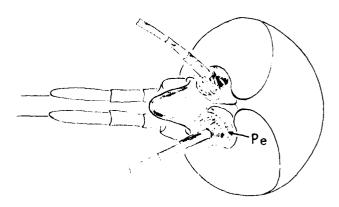


Fig. 111 - Interior crew of head Ar. fitchin

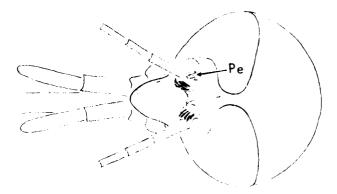
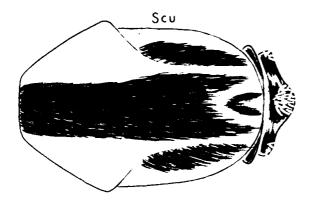


Fig. 113 - Anterior view of head A. Stimulans



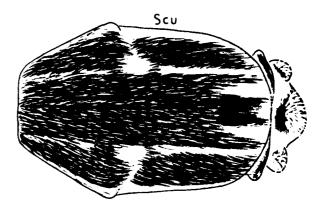


Fig. 112 — Dorsal view of thorax - Ae, mercurator

Fig. 114 — Dorsal view of thorax - Ae. euedes

23(22). Scutum with pale vellowish scales laterally (Fig. 115); dorsal, brown-scaled area of postpronotum at most 0.5 as large as ventral, pale-scaled area (Fig. 116); foretarsomere 3 (Plate 11)

Scutum with pale white, often mixed with yellow or light brown, scales laterally (Fig. 118); dorsal, brown-scaled area of postpropotum equal to or larger than pale-scaled area (Fig. 119): foretarsomere 3 with complete, basal pale ring (Fig. 120) (in part) fitchii (Plate 16)

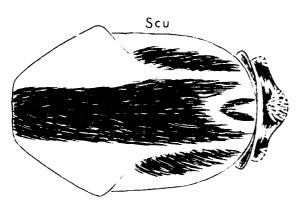
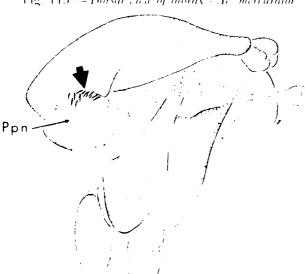


Fig. 145 - Dorsal crew of thorax - Ac. mercurator



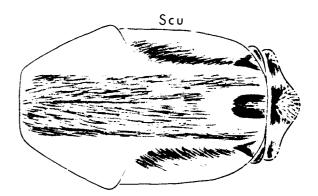


Fig. 118 — Dorsal view of thorax - Ac. fitchii

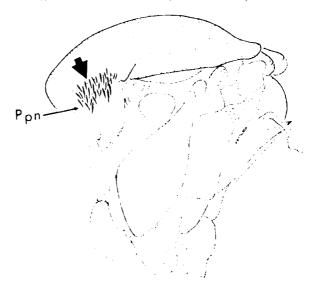


Fig. 119 I aloud case of thorax Ac, fitchic



Fig. 117 — Lateral view of foretarsus - Ae. mercurator

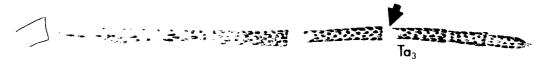


Fig. 120 — Lateral view of foretarsus - Ae. fitchii

24(22).	Foreclaw markedly bent just distad to tooth (Fig. 121); abdominal sterna VI-VIII	
(/	pale-scaled or with few dark scales only (Fig. 122)	, stimulans
	paic scarce of with ten dark sense , , , ,	(Plate 11)

Foreclaw evenly curved distad to tooth (Fig. 123); abdominal sterna VI-VIII pale-scaled with rather broad, medioapical, dark-scaled patches (Fig. 124) (in part) euedes (Plate 15)

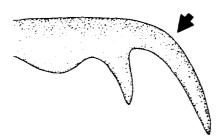


Fig. 121 - Foreclaw - Ar. stimulans

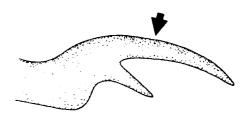


Fig. 123 - Foreclaw - Ae. enedes

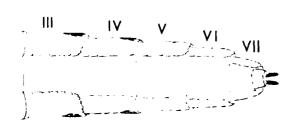


Fig. 122 - Ventral view of abdomen - Ac. stimulans



Fig. 124 — Ventral view of abdomen - Ac. ewdes

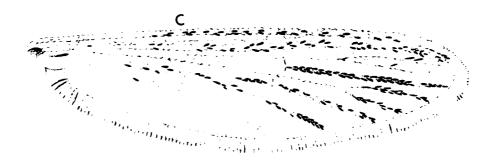


Fig. 125 - Dorsal view of wing - Ae. dorsalis

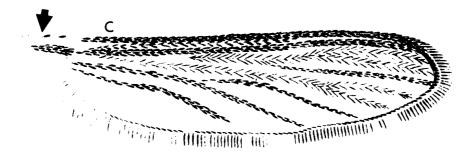
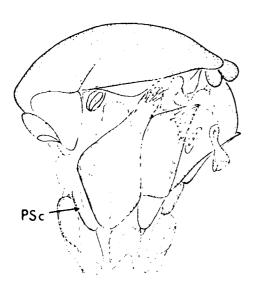


Fig. 127 — Dorsal view of wing - Ae. atropalpus



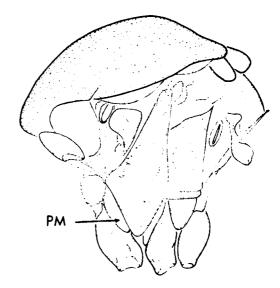


Fig. 126 — Lateral view of thorax - Ac, dorsalis Fig. 128 — Lateral view of thorax - Ac, atropalpus

). Wing yem C mostly dark-scaled (Fig. 129); abdominal tergum VII usually with more dark	
	than pale scales (Fig. 130)	
		(Plate 23
	Vein C mostly pale-scaled (Fig. 131); tergum VII with more pale than dark scales	
	(Fig. 132)	27

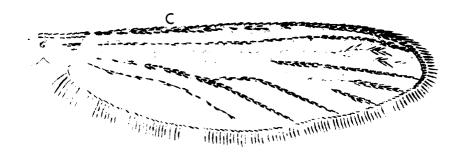


Fig. 129 - Dorsal view of wing - Ae. melanimon



Fig. 131 — Dorsal view of wing - Ae. dorsalis

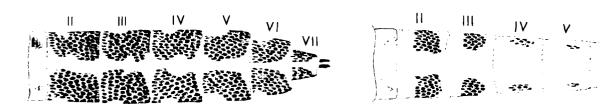


Fig. 130 — Dorsal view of abdomen - Ae, melanimon

Fig. 132 — Dorsal view of abdomen - Ac. dorsalis

27(26). Wing vein R_{4+5} with more dark scales than veins R_9 and R_8 (Fig. 133); foreclaw almost	
straight in middle (Fig. 134)	١.
(Plate 18	()
Vein R_{3+5} with as many dark scales as R_2 and R_3 (Fig. 135); foreclaw abruptly curving near	
attachment of tooth (Fig. 136)	`
(Plane 19	٠.



Tig. 133 Dorsal case of a rig. To december



Fig. 135 - Dorsal view of wing - Ac. campestris

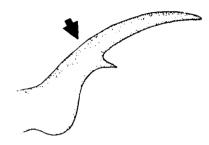


Fig. 134 — Forcelaw Ac. dorsalis

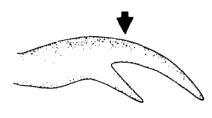


Fig. 136 — Foreclaw - Ac, campestris

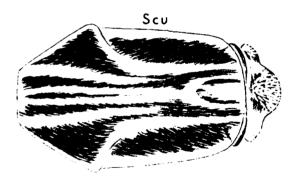


Fig. 137 — Dorsal view of thorax - Ac. togol

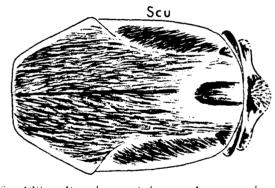


Fig. 138 — Dorsal view of thorax - Ac. e. canadensis



Fig. 139 — Dorsal view of wing Ac. e. canadensis

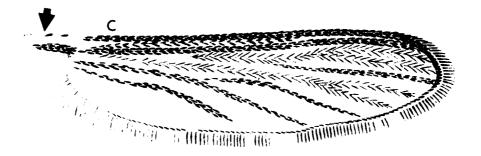
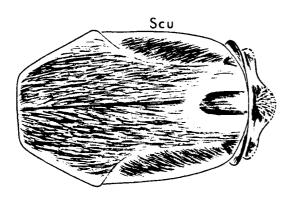


Fig. 141 — Dorsal view of wing - Ac. atropalpus



Scu

Fig. 140 — Dorsal view of thorax - Ac. c. candensis

Fig. 142 — Dorsal view of thorax - Ae. atropalpus

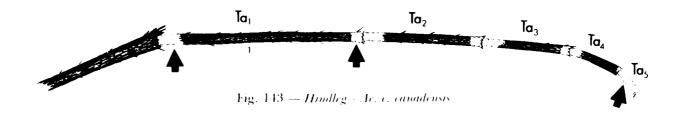
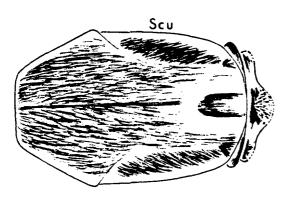




Fig. 145 - Hindleg Ac. e. mathesoni



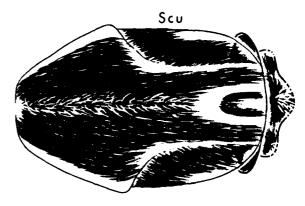


Fig. 144 — Dorsal view of thorax - Ac. c. canadensis

Fig. 146 — Dorsal view of thorax - Ae. c. mathesoni

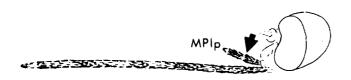




Fig. 147 — Lateral view of head - Ae, atropalpus

Fig. 150 — Lateral view of head - Ae. sierrensis

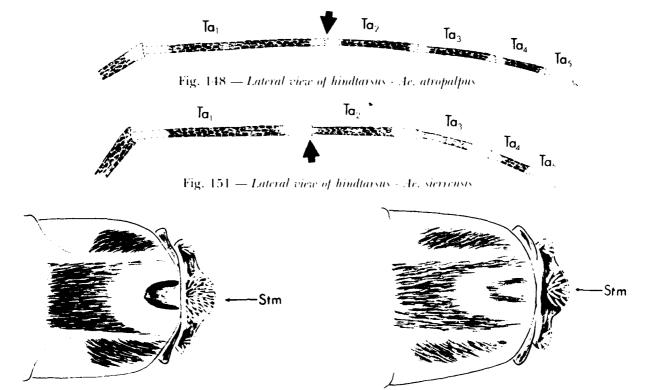


Fig. 149 - Posterior dorsal view of thorax - Ac, atropalpus

Fig. 152 — Posterior dorsal view of thorax Ae, surrensis

32(31).	Interocular space no wider than 2.0 diameter of single corneal facet (Fig. 153); hindfemur
	with dark scales to near base anteriorly (Fig. 154); scutal fossa with 1 or more strong,
	posterior setae (Fig. 155)
	(Plate 13

Interocular space at least 2.5 diameter of single corneal facet (Fig. 156); hindfemur usually entirely pale in basal 0.3-0.5 (Fig. 157); scutal fossa without posterior setae (Fig. 158) atropalpus (Plate 13)

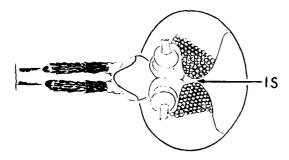


Fig. 153 — Front view of head - Ae. epactius

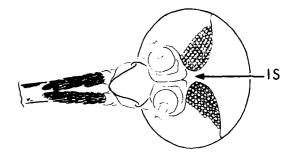


Fig. 156 — Front view of head - Ae. atropalpus



Fig. 154 — Dorsal view of hindleg - Ae. epactius



Fig. 157 — Dorsal view of hindleg - Ae. atropalpus

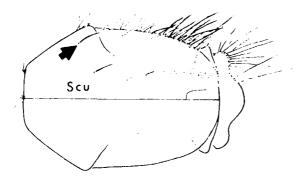


Fig. 155 — Dorsal view of thorax - Ac, epactus

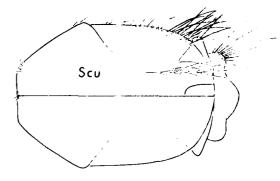


Fig. 158 — Dorsal view of thorax - Ac, atropalpus

33(31). Postprocoxal scale patch present (Fig. 159).	monticola
	(Plate 22)

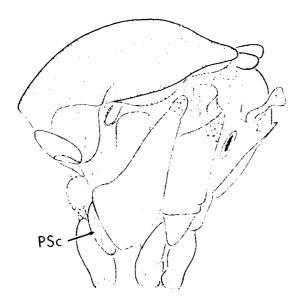


Fig. 159 — Lateral view of thorax - Ae, monticola

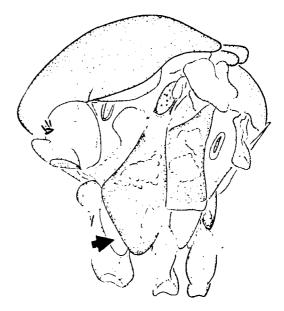


Fig. 160 - Lateral view of thorax - Ae, sierrensis

34(33). Subspiracular area with several light-colored setae arising from scale patch (Fig. 161) varipalpus (Plate 22)

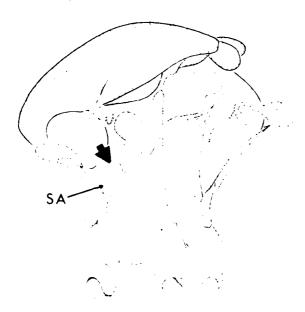


Fig. 161 -- Lateral crew of thorax Ae, varipalpus

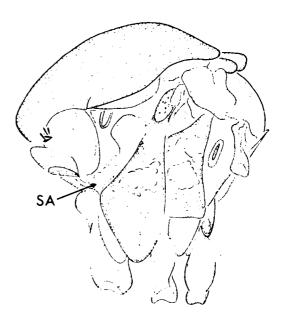


Fig. 162 - Lateral cuew of thorax Accountings

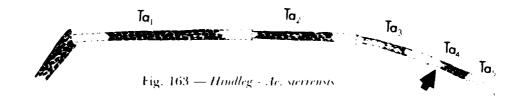




Fig. 165 - Hindleg - Ac. descritcola

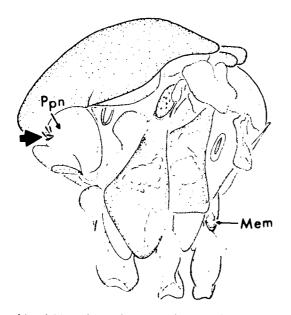


Fig. 164 — Lateral view of thorax - Ae, sierrensis

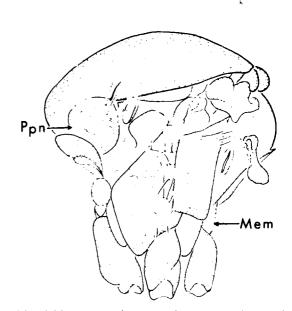


Fig. 166 — Lateral view of thorax - Ae. deserticola

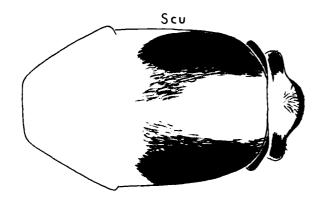


Fig. 167 — Dorsal view of thorax - Ac. fulvus pallens

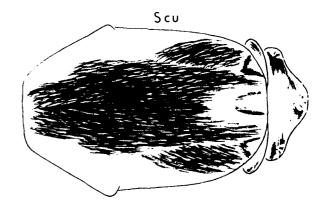


Fig. 168 — Dorsal view of thorax - Ac. triseriatus

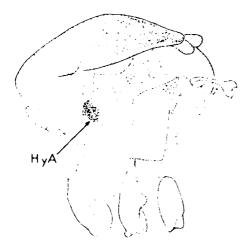


Fig. 169 — Lateral view of thorax - Ae. fulvus pallens

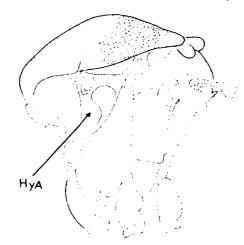


Fig. 171 — Lateral view of thorax - Ae, bimaculatus



Fig. 170 — Dorsal circu of abdomen - Ac. fulvus pallens



Fig. 172 — Dorsal view of abdomen - Ac. bimaculatus

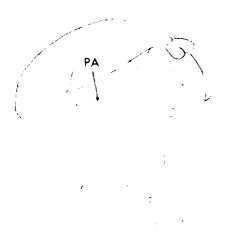


Fig. 173 Lateral case of therax. Accompanienes

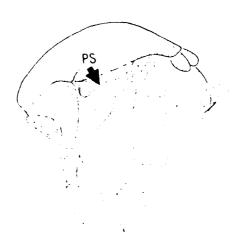
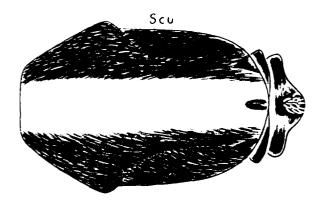


Fig. 174 Lateral in a of thorax Ac hendersoni

39(38).	Soutum with patch or 1,2 median or submedian stripes of silvery white, pale white or pale	
	vellow scales, or with silvery white scales laterally (Figs. 175, 176)	40
	Scutum without silvery white scales medially or laterally, or pale white or pale yellow scales	
	medially (Fig. 177)	49



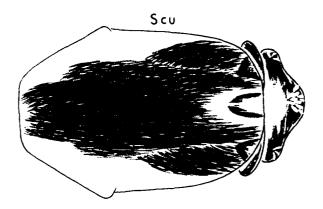


Fig. 175 — Dorsal view of thorax - Ac. atlanticus

Fig. 176 — Dorsal view of thorax - Ae, triscriatus

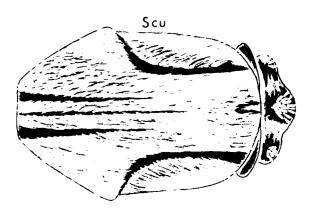
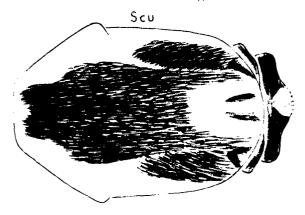


Fig. 177 -- Dorsal view of thorax - Ac. pullatus





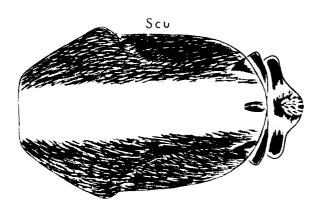


Fig. 179 Dorsal creat of thorax. Te, atlanticus

41(40).	Setae of anterior portion of scutum relatively few and weak; silver scaling of scutal fossa	
	usually restricted to lateral and posterior portions (Fig. 180); claws of fore- and midlegs	
	evenly curved, tooth less than 0.3 length of claw (Fig. 181)	trisinatus
		(Plate 16
	Setae of anterior portion of scutum numerous and well developed; silver scaling usually	
	covering entire scutal fossa (Fig. 182); claws of fore-and midlegs abruptly curving, tooth	
	0.3-0.5 length of claw (Fig. 183)	1.3

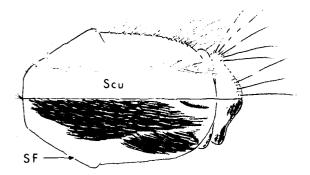


Fig. 180 - Dorsal view of thorax - Ar. triscriatus

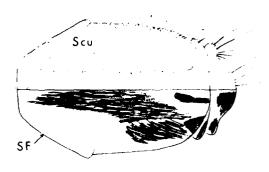


Fig. 182 - Dorsal view of thorax Ac, hendersoni



Fig. 181 — Forcelas Ac. trisermtus



Fig. 183 — Forcelaw - Ac. hendersom

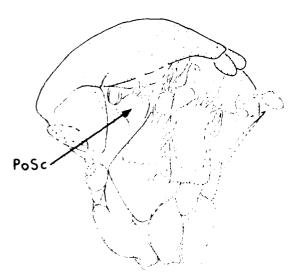


Fig. 184 - Lateral view of thorax Ac, hendersoni

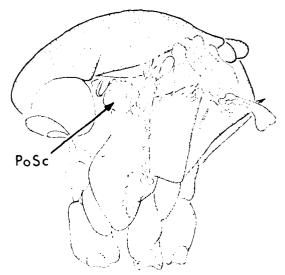


Fig. 186 Lateral view of thorax Ac. brelandi



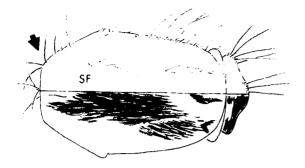
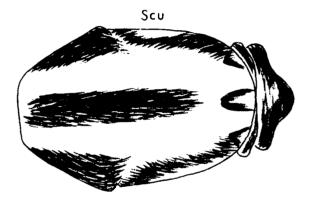


Fig. 185 - Dorsal circu of thorax - Ar. hendersoni

Fig. 187 — Dorsal view of thorax - Ac. brelandi



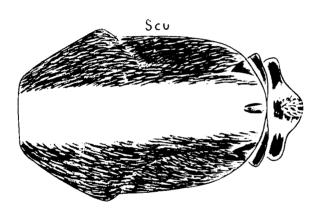


Fig. 188 — Dorsa' view of thorax Ac. trivittatus

Fig. 189 — Dorsal view of thorax - Ac, atlanticus

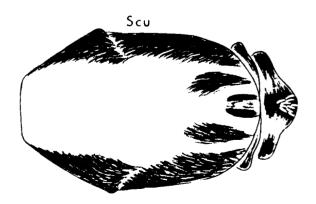


Fig. 190 — Dorsal cica of thorax — Ic. informatus

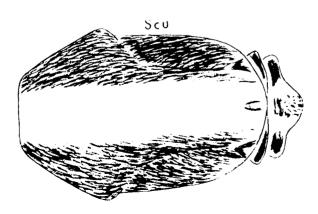


Fig. 191 Dorsal case of thorax Ac atlantions

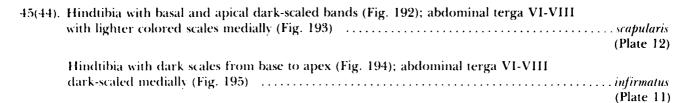




Fig. 192 — Hindleg - Ae. scapularis



Fig. 194 — Hindleg - Ac, infirmatus

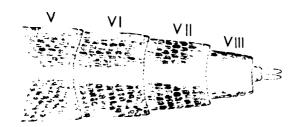


Fig. 193 - Dorsal view of abdomen - Ar. scapularis

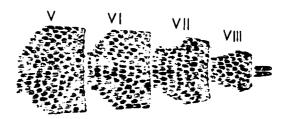


Fig. 195 — Dorsal view of abdomen—Ae, infirmatics

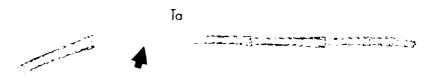


Fig. 196 Millig To burgeri



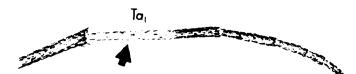
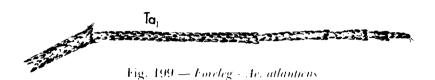


Fig. 197 — Foreleg - Ac. burgeri



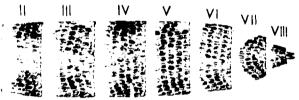
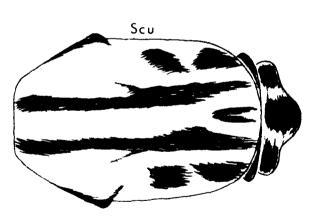


Fig. 200 - Dorsal cure of abdomen. Ac. mnelleri

Fig. 202 - Dorsal view of abdomen - Ae, atlanticus



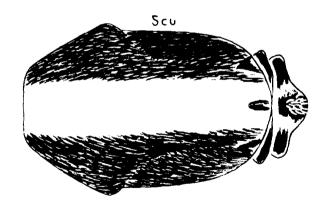


Fig. 201 Darsal crew of thorax Ac, muelleri

Fig. 203 - Dorsal view of thorax Ac, atlantious

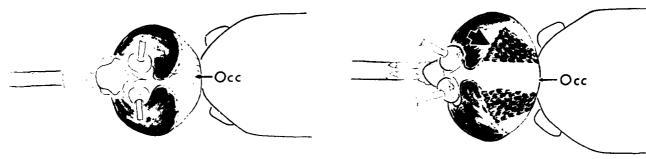


Fig. 204 — Dorsal view of head - Ae. dupreei

Fig. 205 — Dorsal view of head - Ae. atlanticus

49(39)	. Wing with many pale scales either confined to anterior veins, some on all veins, or veins	
	alternating dark and pale-scaled (Figs. 206, 207))
	Wing veins entirely dark-scaled or with pale scales at base of vein C and sometimes Sc and	
	R (Fig. 208)	,



Fig. 206 - Dorsal view of wing - Ae. niphadopsis



Fig. 207 -- Dorsal view of wing - Ac. s. idahoensis

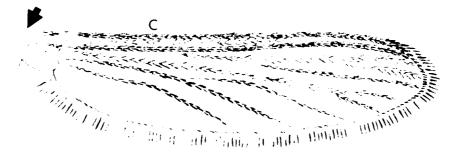


Fig. 208 - Dersal crea of army 4e. pullatus

50(49).	Wing with veins alternating dark and pale-scaled, R_1 , R_{4+5} , and Cu dark, others pale (Fig.	
	209)	i l
	Wing with pale scales scattered over all veins or confined to anterior veins (Fig. 210)	52

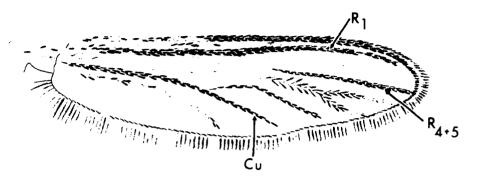


Fig. 209 - Dorsal view of wing - Ae. s. idahoensis

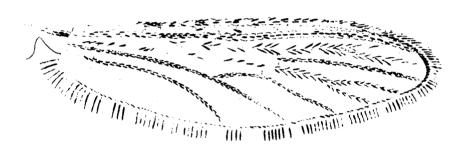


Fig. 210 - Dorsal view of wing - Ae. niphadopsis

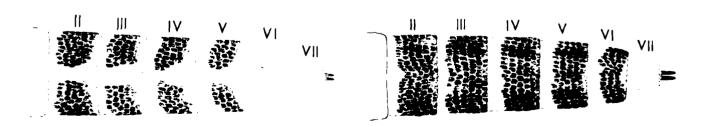
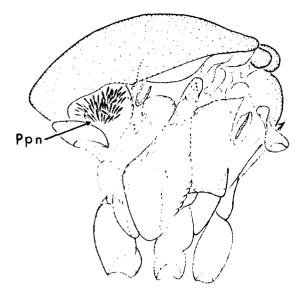
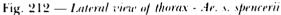


Fig. 211 — Dorsal view of abdomen Ac. s. spencern

Fig. 213 - Dorsal crew of abdomen - Ac. s. idahoensis





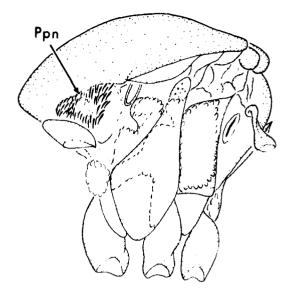


Fig. 214 — Lateral view of thorax - Ae. s. idahoensis

52(50). Palpus and proboscis dark-scaled (Fig. 215); lower mesanepimeral setae absent (Fig. 216) (in part)

ventrovittis

(Plate 19)

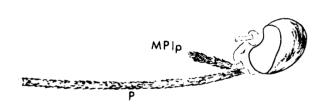


Fig. 215 — Lateral view of head - Ae, ventrovittis

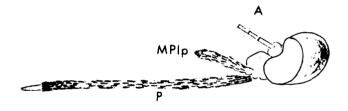


Fig. 217 — Lateral view of head - Ae. bicristatus

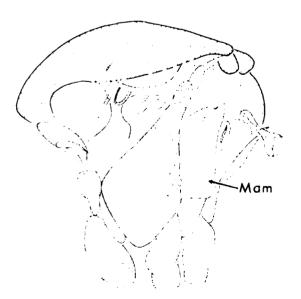


Fig. 216 — Lateral view of thorax Ac. ventrovittis

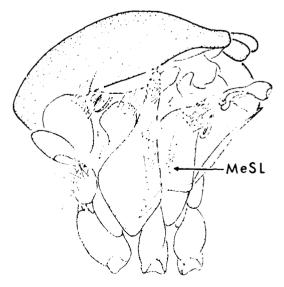


Fig. 218 — Lateral view of thorax - Ac. cataphylla

53(52).	Abdominal terga with broad, basal, pale bands and apical, pale scales, often forming
	median, longitudinal stripe (Fig. 219); pale scales numerous on wing veins anterior to Cu
	(Fig. 220)niphadopsis
	(Plate 13)

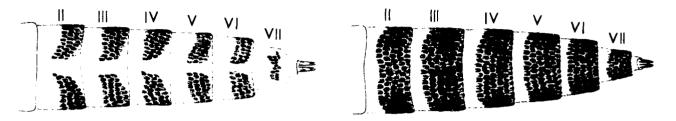


Fig. 219 — Dorsal view of abdomen - Ae, niphadopsis

Fig. 221 — Dorsal view of abdomen - Ac. cataphylla

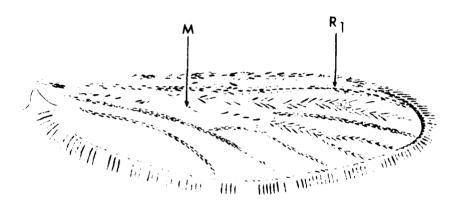


Fig. 220 -- Dorsal view of wing - Ac. inphadopsis



Fig. 222 Dorsal view of wing Accentaphylla

54(53)	 Scutum with area of broad, curved scales laterally at level of mesothoracic spiracle (Fig. 223); palpus longer than basal 3 antennal flagellomeres (Fig. 224)	ristatus
		ate 15)
	Scutium with only narrow scales laterally at level of mesothoracic spiracle (Fig. 225); palpus	
	shorter than basal 3 antennal flagellomeres (Fig. 226)	aphylla
	(P)	ate 10)

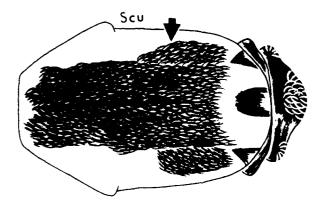


Fig. 223 - Dorsal view of thorax - Ae, bicristatus

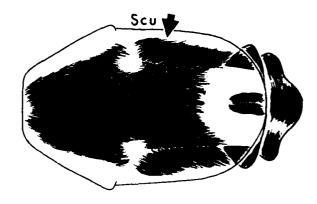


Fig. 225 — Dorsal view of thorax - Ae, cataphylla

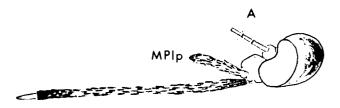


Fig. 224 — Lateral view of head - Ac. bicristatus

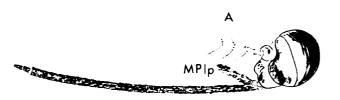


Fig. 226 — Lateral view of head - Ae, cataphylla

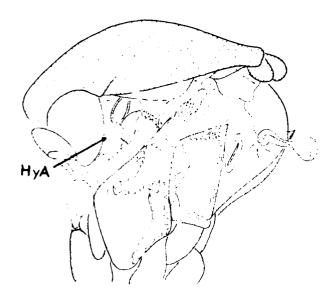


Fig. 227 — Lateral view of thorax - 4e, pullatus

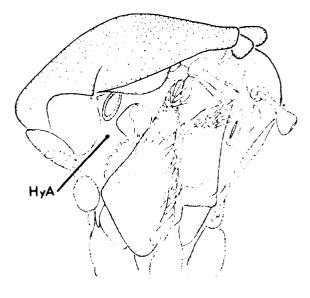


Fig. 228 = I ateral crea of thorax. Ac digitations

56(55). Postprocoxal scale patch absent (Fig. 229); palpus usually with some pale scales (Fig. 230)
 Postprocoxal scale patch present (Fig. 231); palpus entirely dark-scaled (Fig. 232)
 58

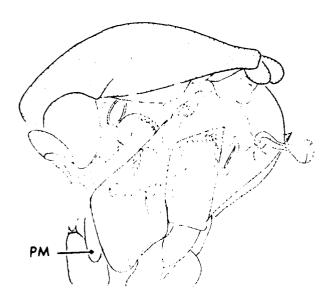


Fig. 229 Lateral circu of thorax. Ac. pullatus

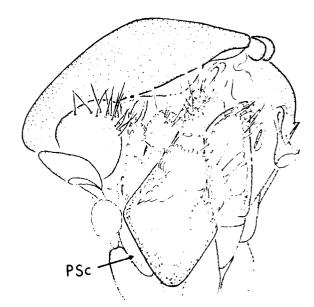


Fig. 231 — Lateral ciew of thorax - Ac. implicatus



Fig. 230 - Lateral view of head Ac. fullatus

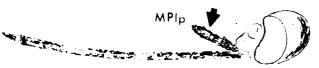


Fig. 232 -- Lateral view of head Ac. implicative

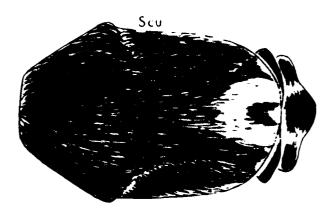


Fig. 233 Dorsal case of thorax Ac introdens

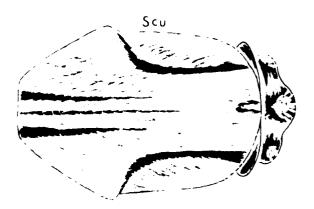
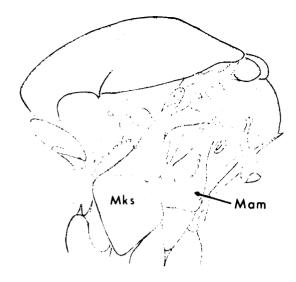


Fig. 235 Dorsal and of thorax of pullatus



Mks Mam

Fig. 234 — Lateral view of thorax - Ac. intrudens

Fig. 236 — Lateral view of thorax - Ae. pullatus

Mesokatepisternum with scales extending to anterior angle, not separated from posterior mesanepisternal scale patch (Fig. 239); femora without apical, pale ring (Fig. 240) provocans (Plate 26)



Fig. 237 - Lateral crew of thorax Ac, implicatus



Fig. 239 - Lateral cuew of thorax - 4c, proceeding





Fig. 240 - Hindleg - Ae. provocans

59(55)					oands, or, if							60
					isal bands o ripe of pale							63
			V V	VI	M	~			IV	V	۷I	
					=						3	VII
/_SI	. 241 — <i>D</i>	orsal view	ot abdom	en Ac. dia	ntacus	ـــــــــــــــــــــــــــــــــــــ	. 942 —	Dorsal vi	ew of abdo	men - Ae	. intrue	lens
). Abdomi	nal sterna	centirely	pale-scaled	(Fig. 243);	forecoxa	i with at l	east some	scales bro	own	0	
		some abe g. 246)			dark scales							61
I	!! 	III .	IV	V . VI	VII `æ	. 1			IV	>	VI	VII अंजे⇒
					,							43

Fig. 243 - Lentral crew of abdomen Ac, anrifer

Fig. 245 - Ventral ciew of abdomen - Ac, thibaulti



Fig. 244 Interior clear of thorax Ac. aurifor

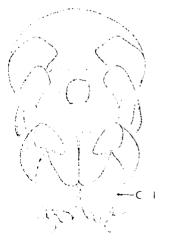


Fig. 246 Interior case of thorax Ac. thibaidh.

stripe of dark brown scales, broadening abruptly	Scutum with broad, median longitudinal	61(60).
· · · · · · · · · · · · · · · · · · ·	just posterior to scutal angle (Fig. 247).	
(Plate 23		

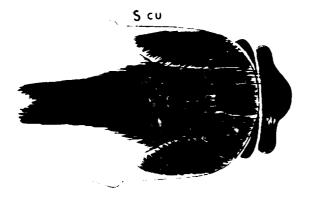


Fig. 247 - Dorsal crew of thorax Ac. thibaulti

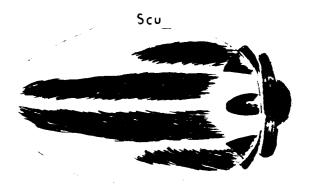


Fig. 248 - Dorsal view of Porax - 4c. dections

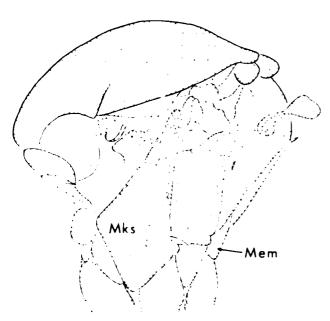


Fig. 249 — Lateral view of thorax Ac. dections

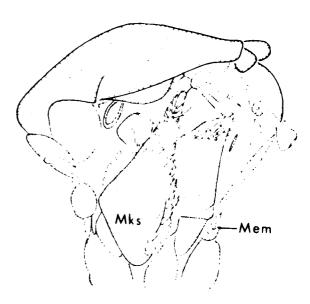


Fig. 251 - Lateral view of thorax. Ac diantaeus

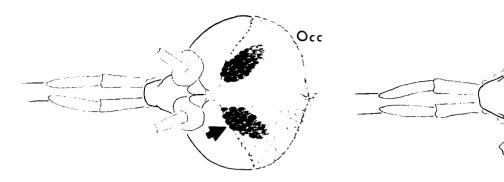


Fig. 250 - Dorsal view of head - Ae, decticus

Fig. 252 — Dorsal view of head - Ae, diantaeus

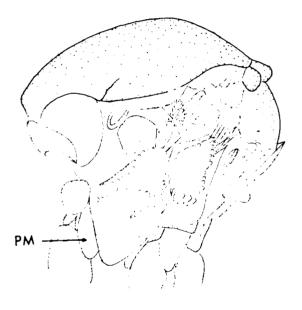


Fig. 253 — Lateral view of thorax - Av. sticticus

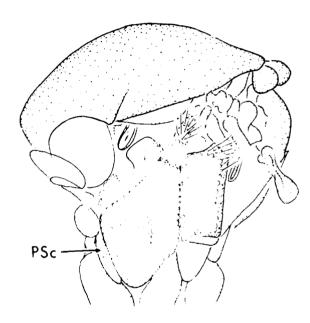


Fig. 254 — Lateral view of thorax - Ac. punctor

64(63). Abdominal terga II-VI with median, basal, triangular patches of pale scales (Fig. 255) theleter (Plate 23)

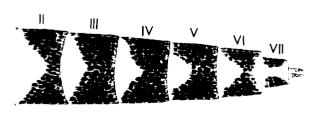


Fig. 255 - Dorsal cuse of abdomen Act theleter

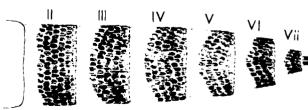


Fig. 256 - Dorsal view of abdomen - Ac. intrudens

anterior angle (Fig. 258)	. 66
Scutum with dark, median, longitudinal stripe (Fig. 259); mesokatepisternum with scales extending to near anterior angle (Fig. 260)	

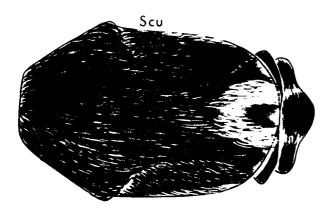


Fig. 257 — Dorsal view of thorax - Ac. intrudens

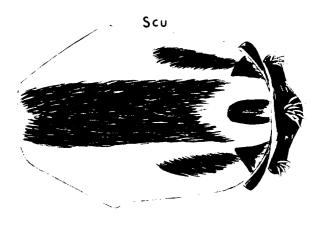


Fig. 259 — Dorsal view of thorax - Ac. sticticus



155 I storal on a of thorax Accombinations

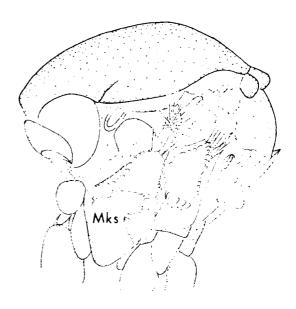


Fig. 260 - Interal error of thorax Accordings

- expole, or with few dark scales only (Fig. 263); subspiracular area with

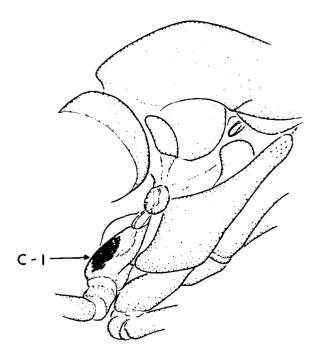


Fig. 261 — Anterior view of thorax - Ae. cinereus

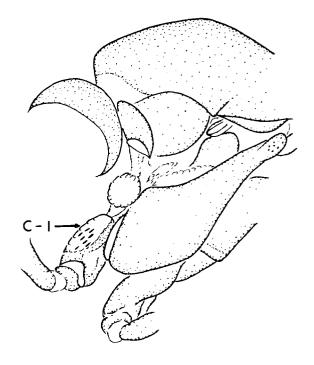


Fig. 263 — Anterior view of thorax - Ae. intrudens

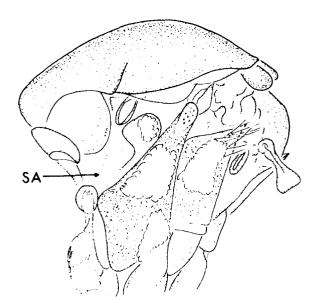


Fig. 262 — Lateral view of thorax Ae, emercus

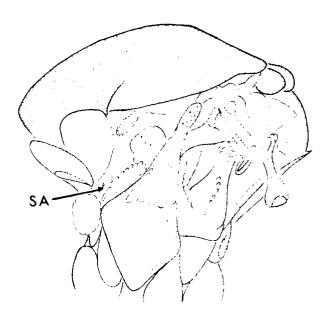
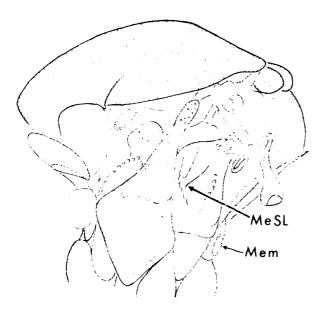


Fig. 264 - Lateral view of thorax - Ac. intrudens

67(66). Lower mesanepimeral setae present; metameron with scales (Fig. 265) (in part) introdens (Plate 9)



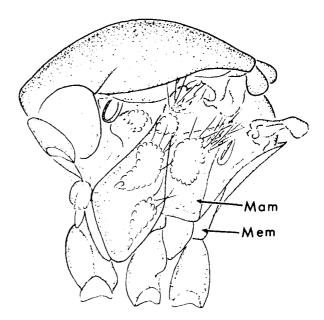


Fig. 265 - Lateral view of thorax - Ac. intrudens

Fig. 266 — Lateral view of thorax - Ac, tortilis

68(65). Scutum with submedian, dark-scaled, longitudinal band wide and varying in width, especially with dark scales in scutal fossa (Fig. 267); foreclaw elongated (Fig. 268) rempeli (Plate 3)

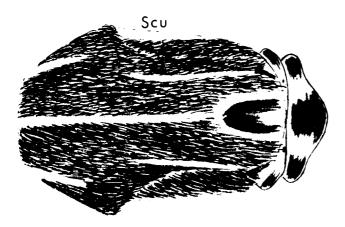


Fig. 267 — Dorsal view of i' max. Ac. rempeli.

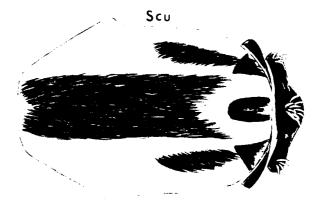


Fig. 269 - Dorsal view of thorax Ac, stictions

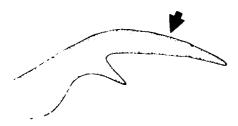


Fig. 268 - Foreclase Ar. rempeli



Fig. 270 - Foreday Ac, stieneus

69(68).	Scutellar and supraalar setae vellowish (Fig. 271); mesanepimeron usually without lower	
	setae, ventral 0.25 devoid of scales (Fig. 272)	sticticus
	···	(Plate 25)
	Scutellar and supraalar setae brown or black (Fig. 273); mesanepimeron with lower setae,	
	control 0.95 scaled (Fig. 974)	70

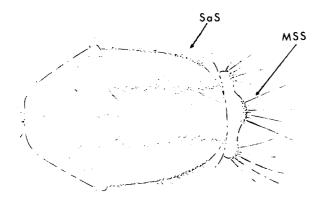


Fig. 271 — Dorsal view of thorax - Ae. sticticu

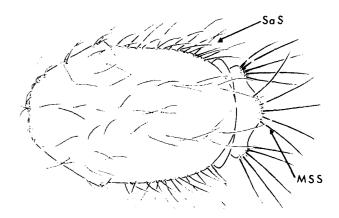


Fig. 273 — Dorsal view of thorax - Ae, communis

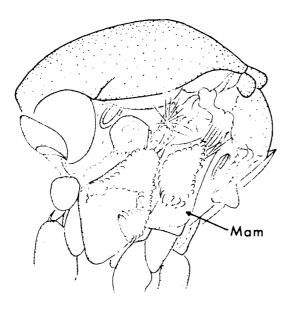


Fig. 272 Lateral view of thorax Ac, stations

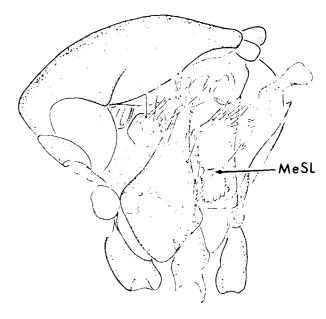


Fig. 274 - Lateral enex of thorax Accommunis

70(69). Looth of hindelaw long, thin, claw usually curving abruptly distal to tooth (Fig. 275) commons (Plate 17)

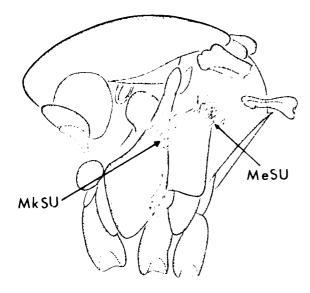


Fig. 275 Hindelaw Accommunis



Fig. 276 -- Hindelaw . Ac, nevadensis

71(70).	Usually 17-27 upper mesanepimeral setae, range 14-33; upper mesokatepisternal setae
	5-8 (Fig. 277)nevadensis
	(Plate 10)
	Usually 12-19 upper mesanepimeral setae, range 10-22; usually 4,5 upper
	mesokatepisternal setae (Fig. 278)
	(Plate 20)



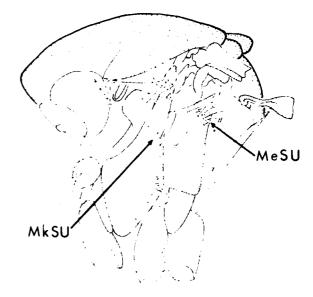
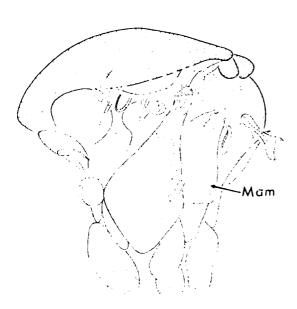
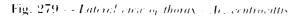


Fig. 277 — Lateral view of thorax - Ae. nevadensis

Fig. 278 — Lateral view of thorax - Ae, churchillensis

72(63). Lower mesanepimeral setae absent (Fig. 279); pale basal band on abdominal tergum II narrowed, or completely interrupted, medially (Fig. 280) (in part) ventrovittis (Plate 19)





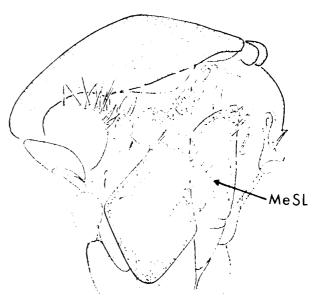


Fig. 281 - Lateral view of thorax - Ac, implicatus

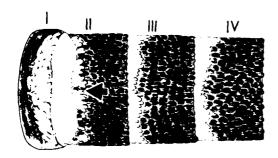


Fig. 280 - Dorsal view of abdomen Ac. centrocittis

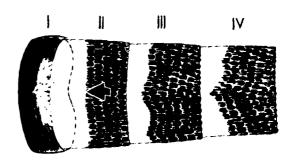


Fig. 282 - Darsal ciew of abdomen - Ac. punctor

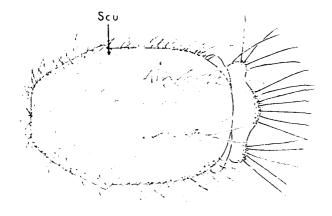


Fig. 283 Dorsal crew of thorax Ac. impiger

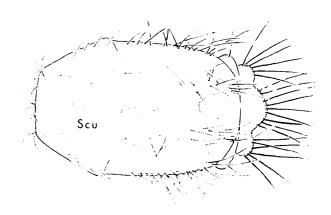
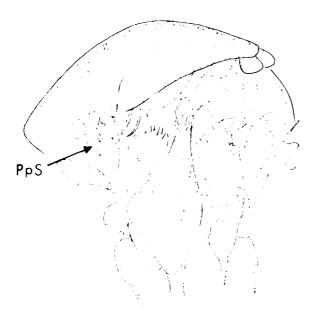


Fig. 285 - Dorsal crew of thorax A. Ir. promps



Tig. 284 Lateral crew of thorax Tel impiger



Fig. 286 Lateral crew of thorax Accomplicatus

74(73).	Foreclaw sharply bent apical to long tooth (Fig. 287); postspiracular setae numbering 10 or
	fewer (Fig. 288)impiger
	(Plate 22)
	Foreclaw elongate, very gradually curving distal to short tooth (Fig. 289); postspiracular
	setae numbering 14 or more (Fig. 290)
	(Plate 25)

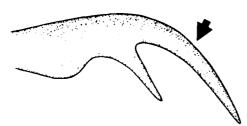


Fig. 287 — Foreclaw - Ac. impiger

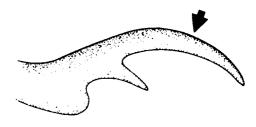


Fig. 289 - Foreclaw - Ac. nigripes

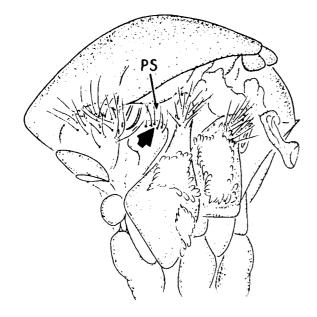


Fig. 288 - Lateral view of thorax - Ac, impiger

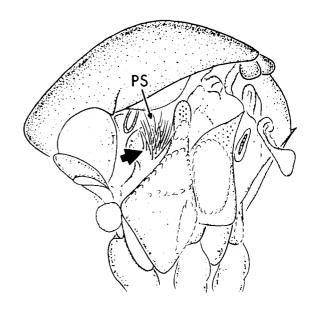


Fig. 290 — Lateral view of thorax (Ac. nigripes

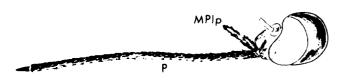


Fig. 291 — Lateral view of head and probosers (Ac. schizopinax)

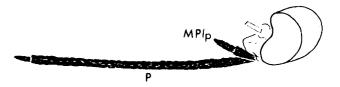


Fig. 293 -- Lateral view of head and probosers. Ac, punctor

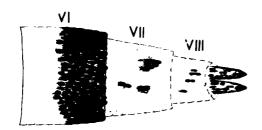


Fig. 292 — Dorsal view of abdomen - Ac. schizopinax

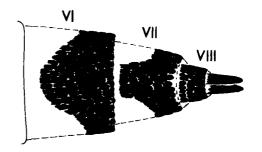


Fig. 294 — Dorsal view of abdomen - Ac. punctor

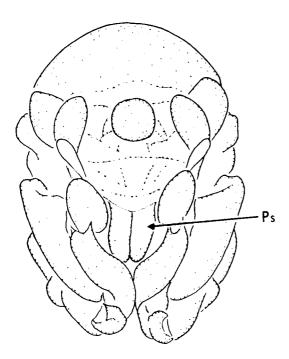


Fig. 295 — Anterior view of thorax Ae, implicatus

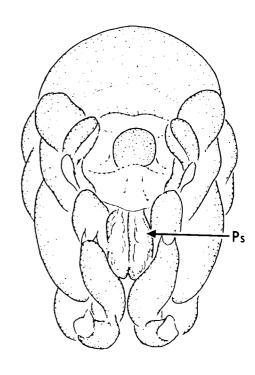


Fig. 296 — Anterior view of thorax - Ac. hexodontus

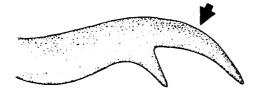


Fig. 297 — Foreclaw - Ae. implicatus

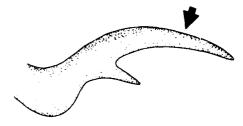


Fig. 300 — Foreclaw - Ae. punctor



Fig. 298 — Lateral view of thorax - Ae. implicatus

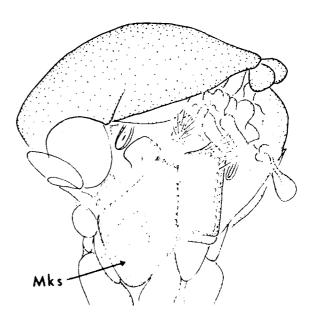


Fig. 301 — Lateral view of thorax - Ae. punctor

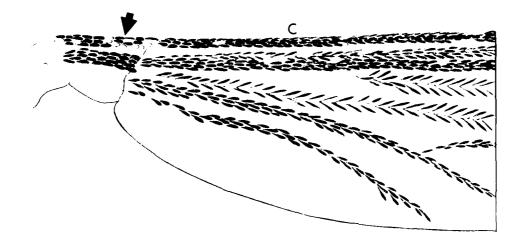


Fig. 299 - Dorsal view of wing - Ac. implicatus

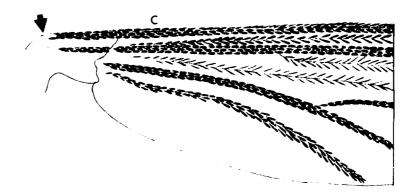


Fig. 302 — Dorsal view of wing - Ac. punctor

78(76).	Supraalar and scutellar setae dark brown or black (Fig. 303); with 15 or more
	postmetasternal scales present (Fig. 304)
	(Plate 18)
	Supraalar and scutellar setae vellow to vellow-brown (Fig. 305); postmetasternal scales
	absent or with 2,3 scales only (Fig. 306)

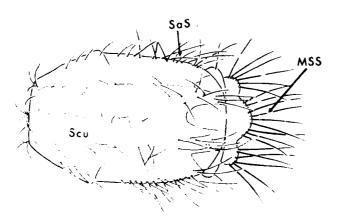


Fig. 303 - Dorsal cure of thoras - Ar. pionips

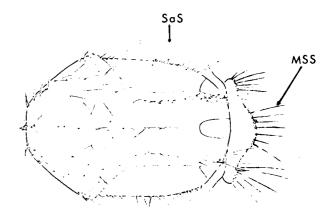


Fig. 305 — Dorsal view of thorax - Ac. hexodontus

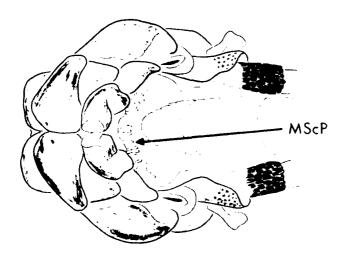


Fig. 304 — Ventral view of abdomen and part of venter of metathorax - Ac. pionips

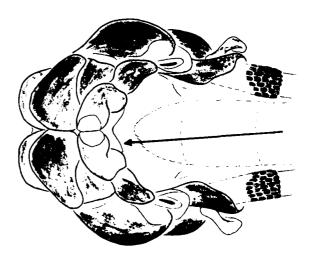


Fig. 306 — Ventral view of abdomen and part of venter of metathorax - Ac. hexodontus

78). Large patch of pale scales at base of wing vein C (Fig. 307); abdominal sterna III-VI pale-scaled apically, or rarely with few dark scales (Fig. 308)	79(78).
Wing dark-scaled or with fewer than 8 pale scales at base of vein C (Fig. 309); abdominal sterna 111-V1 with many dark scales apically (Fig. 310) (in part) punctor punctodes abserratus (Plates 13, 25, 10)	
a. Found in eastern North America (Fig. 311)	
aa. Found in Alaska only (Fig. 311)	
aaa. Widely distributed in northern North America (Fig. 311)	

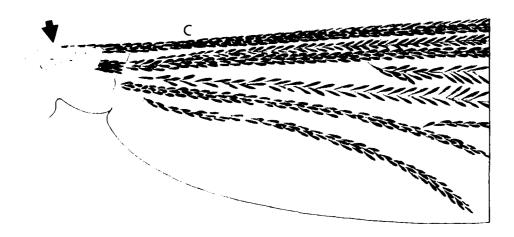


Fig. 307 — Dorsal view of wing - Ar. hexodontus

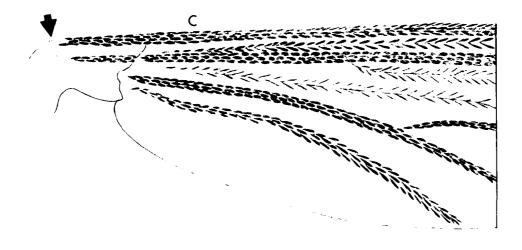


Fig. $309 \sim Dorsal\ crew of\ wing \sim Ae,\ punctor$

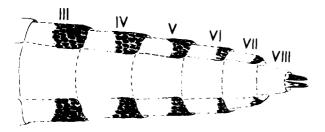


Fig. 308 — Ventral view of abdomen - Ae, hexodontus



Fig. 310 — Ventral view of abdomen - Ae, punctor

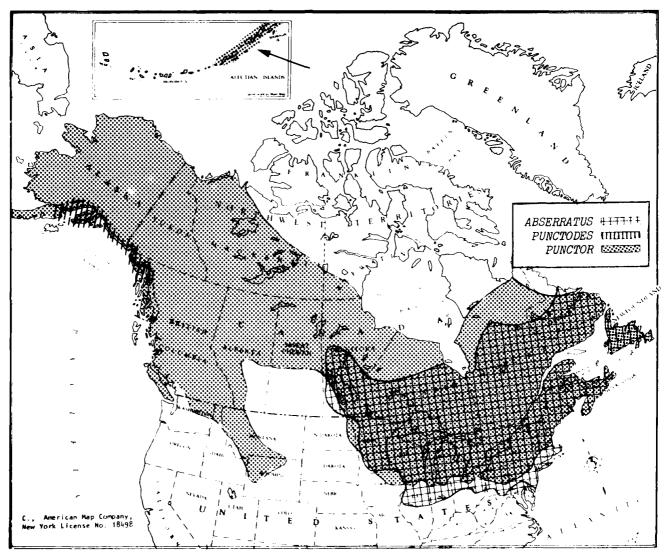


Fig. 311 — Distributional map

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS ANOPHELES

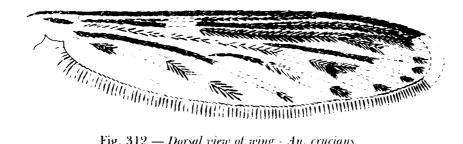


Fig. 312 — Dorsal view of wing - An. crucians

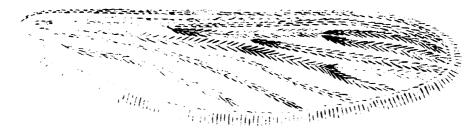


Fig. 313 — Dorsal view of wing - An, quadrimaculatus



Fig. 314 — Dorsal view of wing - An. earlei

2(1). Hindtarsomeres with apical 0.5 of 2, all of 3, 4, and 5 pale-scale cept for basal ring of (Plate 30)



Fig. 315 -- Hindleg - An. albimanus



Fig. 316 - Hindley An. punctipennis

Wing with apical pale spot, otherwise vein C dark-scaled; vem A with 3 dark spots (Fig. 3(2).Inadlesi georgianus

(Plates 31, 29, 30)

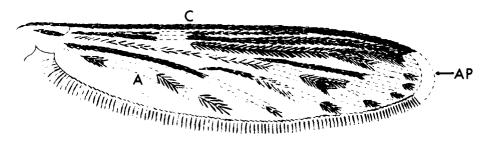


Fig. 317 — Dorsal view of wing - An. crucians

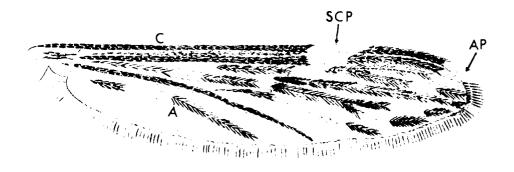


Fig. 318 — Dorsal view of wing - An. punctipennis



Fig. 319 — Lateral view of head - An, punctipennis

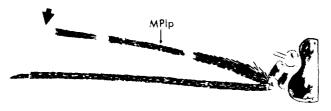
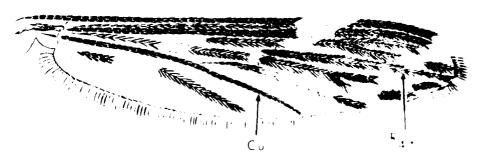
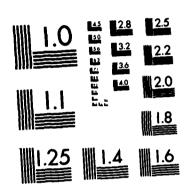


Fig. 321 — Lateral view of head - An, pseudopunctificials



132 320 - 20

IDENTIFICATION AND GEOGRAPHICAL DISTRIBUTION OF THE MOSQUITOES OF NORTH AMERICA NORTH OF MEXICO(U) WALTER REED ARMY INST OF RESEARCH WASHINGTON DC F F DARSIE ET AL. 81 AUG 81 F/G 6/3 2/4 AD-A125 975 UNCLASSIFIED NL



MICROCOPY RESOLUTION TEST CHART NATIONAL BUREAU OF STANDARDS-1963-A

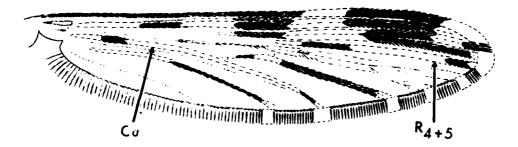


Fig. 322 — Dorsal view of wing - An. pseudopunctipennis

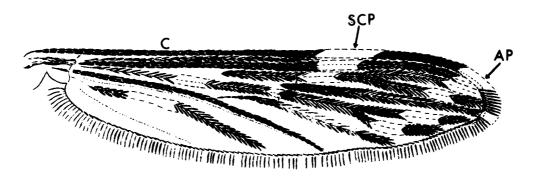


Fig. 323 - Dorsal view of wing - An. punctipennis

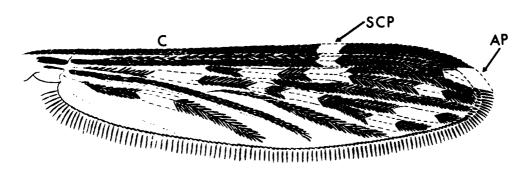


Fig. 324 - Dorsal view of wing - An. perplexens

Vein M mostly dark-scaled (Fig. 327); apical segment of palpus with dark scales (Fig. 328) franciscanus (Plate 28)

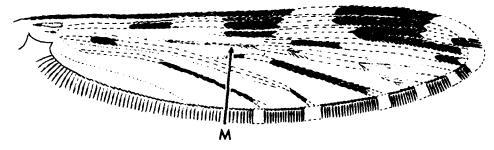


Fig. 325 - Dorsal view of wing - An. pseudopunctipennis

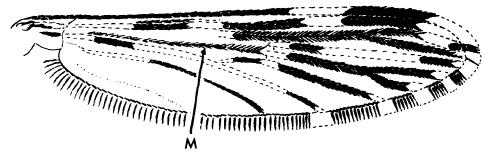


Fig. 327 - Dorsal view of wing - An. franciscanus



MPIp

Fig. 326 — Lateral view of head - An. pseudopunctipennis

Fig. 328 - Lateral view of head - An. franciscanus



Fig. 329 — Dorsal view of wing - An. earlei

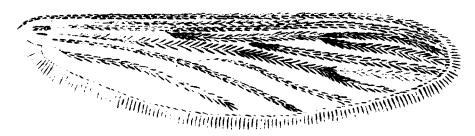


Fig. 330 - Dorsal view of wing - An. quadrimaculatus

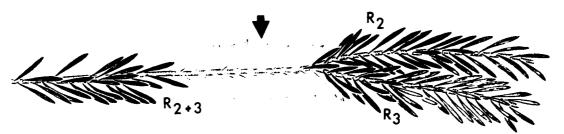


Fig. 331 - Dorsal view of wing - An. earlei

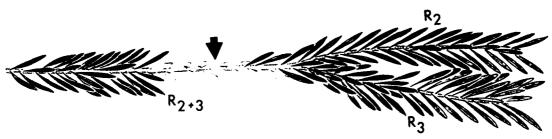


Fig. 332 - Dorsal view of wing - An. occidentalis



Fig. 333 - Dorsal view of wing - An. barberi

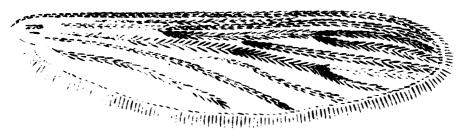


Fig. 335 — Dorsal view of wing - An. quadrimaculatus

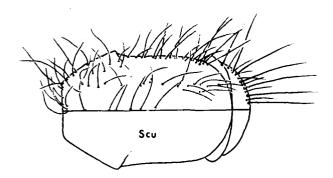


Fig. 334 — Dorsal view of thorax - An. barberi

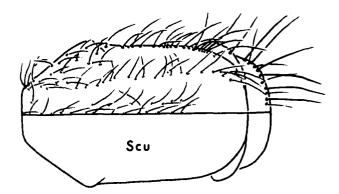


Fig. 336 - Dorsal view of thorax - An. freeborni

(Plate 30)

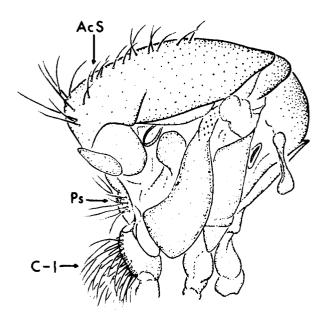


Fig. 337 — Lateral view of thorax and mesoscutum - An. barberi

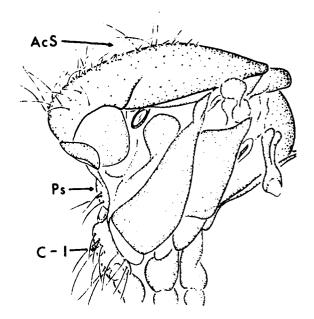


Fig. 338 — Lateral view of thorax and mesoscutum - An. judithae



Fig. 339 - Lateral view of head - An. freeborni

Fig. 341 — Lateral view of head - An. walkeri

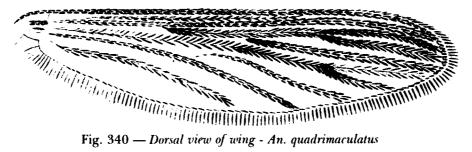


Fig. 340 — Dorsal view of wing - An. quadrimaculatus



Fig. 342 - Dorsal view of wing - An. atropos

12(11). Scales on basal part of wing vein Cu elongate with apices truncate (Fig. 343); in western	
USA and Canadafreeb	orni
(Plate	
Scales on base of vein Cu obovate with apices rounded (Fig. 344); in eastern USA and	
Canadaquadrimacul	atus
. (Plate	

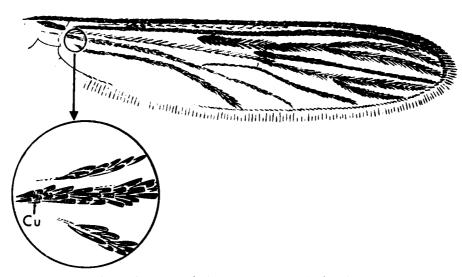


Fig. 343 — Dorsal view of wing - An. freeborni

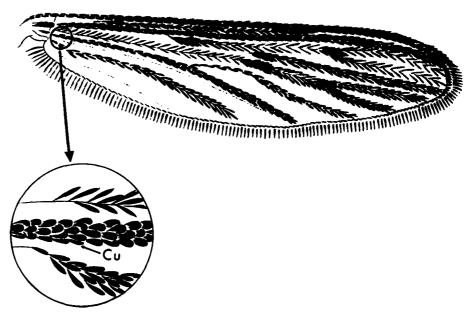


Fig. 344 — Dorsal view of wing - 4n. quadrimaculatus



Fig. 345 — Halter enlarged - An. walkeri



(Plate 28)

Fig. 348 — Halter enlarged - An. atropos

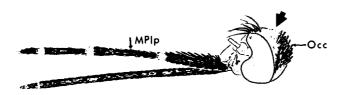


Fig. 346 — Lateral view of head - An. walkeri

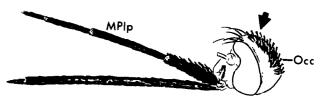
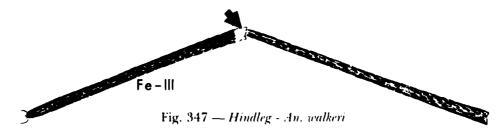
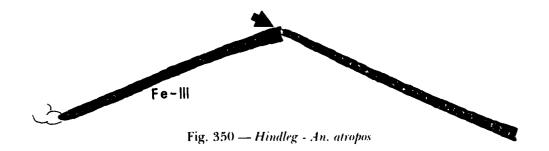


Fig. 349 — Lateral view of head - An. atropos





KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS CULEX

1.	Scutum with middorsal, acrosticnal setae (Fig. 351); occiput with narrow scales dorsally			
	(Fig. 352)	. 2		
	Scutum without middorsal, acrostichal setae (Fig. 353); occiput with broad, appressed			
	scales dorsally, sometimes limited to borders of eyes (Fig. 354)	19		

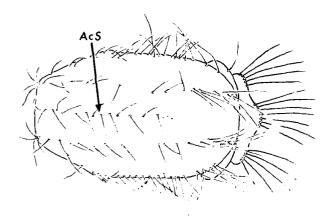


Fig. 351 - Dorsal view of thorax - Cx. pipiens

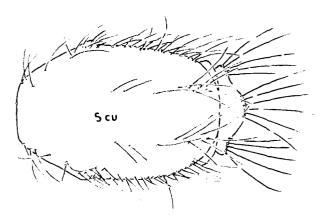


Fig. 353 — Dorsal view of thorax - Cx, erraticus

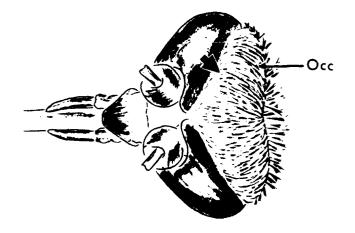


Fig. 352 - Dorsal view of head - Cx. pipiens

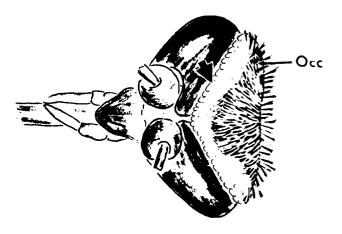


Fig. 354 — Dorsal view of head - Cx, erraticus

2(1).	Abdominal terga with bands or lateral spots of pale scales along basal border (Fig. 355) (subgenus Culex)						
Fig	. 355 — Dorsal view of abdomen - Cx. restuan		II IV	V VI VII domen - Cx. territar	B		
3(2).	Hindtarsomeres with rather distinct, base	Ü	-				
J(Z).	Hindtarsomeres dark-scaled, or if with pa	lle scales, then as very narr	ow, basal rings	(Fig.			
	Ta ₁	Ta ₂	Ta ₃	Ta ₄ Ta ₅			
	Fig. 357	— Hindleg - Cx. tarsalis					
	Ta ₁	Ta ₂	To ₃	Ta ₄ Ta			
	Fig. 358 -	— Hindleg - Cx. restuans		Ta ₅			
4(3).	Proboscis with complete, distinct ring of Proboscis without complete, distinct ring						
~	♦ P		P				
F	ig. 359 — Lateral view of head - Cx. tarsalis	Fig. 360 —	- Lateral view o _l	Chead - Cx. pipiens			
5(4).	Hindtarsomeres with basal and apical rin	ngs of pale scales narrow	(Fig. 361)		uensis e 25)		
	Hindtarsomeres with basal and apical rit	ngs of pale scales rather b	road (Fig. 362)	6		

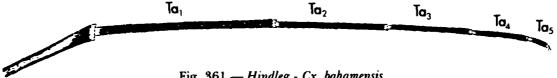


Fig. 361 — Hindleg - Cx. bahamensis

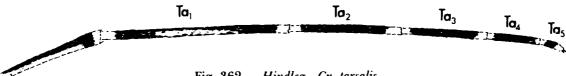


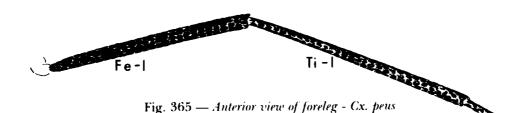
Fig. 362 — Hindleg - Cx. tarsalis

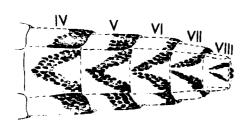
Anterior surface of forefemur and tibia with pale stripe or line of pale-scaled spots (Fig. 6(5).363); V-shaped, dark-scaled marks on abdominal sterna (Fig. 364) tarsalis (Plate 34)

Forefemur and tibia without pale stripe or line of spots (Fig. 365); sternal dark marks on abdomen oval in shape (Fig. 366)peus (Plate 37)



Fig. 363 - Anterior view of foreleg - Cx. tarsalis





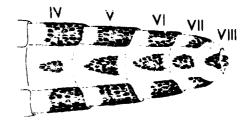
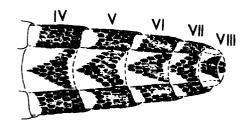


Fig. 364 — Ventral view of abdomen - Cx. tarsalis

Fig. 366 - Ventral view of abdomen - Cx. peus

Abdominal sterna with median triangular areas of dark scales (Fig. 367)thriambus 7(4). (Plate 39)



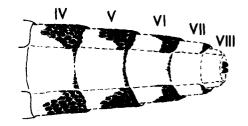
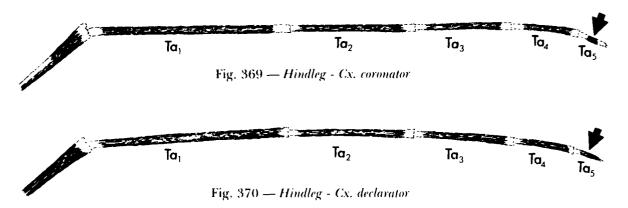


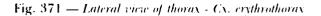
Fig. 367 — Ventral view of abdomen - Cx. thriambus

Fig. 368 — Ventral view of abdomen - Cx. coronator

8(7).	Hindtarsomere 5 with rings of pale scales basally and apically, with dark scales medially	
	(Fig. 369)	. coronator
		(Plate 35)
	Hindtarsomere 5 with narrow ring of pale scales basally, otherwise dark-scaled (Fig. 370)	declarator
		(Plate 36)







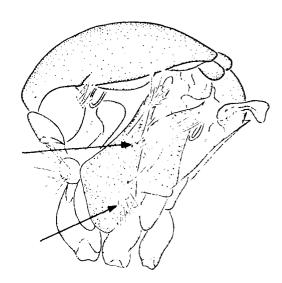


Fig. 373 — Lateral view of thorax - Cx. nigripalpus

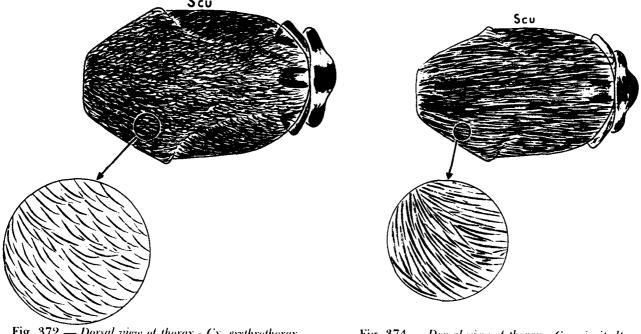


Fig. 372 — Dorsal view of thorax - Cx. erythrothorax

Fig. 374 — Dorsal view of thorax - Cx. nigripalpus

Abdominal terga with conspicuous basal bands of pale scales (Fig. 376) $\dots 13$



I۷ VII VIII

Fig. 375 — Dorsal view of abdomen - Cx. nigripalpus

Fig. 376 — Dorsal view of abdomen - Cx. restuans

11(10). Scale patches on thoracic pleura absent, or if present, in groups of fewer than 6 scales (Fig. (Plate 37) Thoracic pleura with several groups of pale scales with 6 or more scales each (Fig. 379);

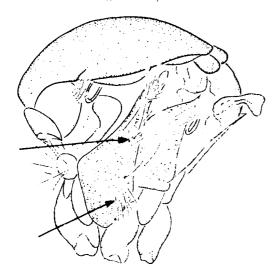


Fig. 377 — Lateral view of thorax - Cx. nigripalpus

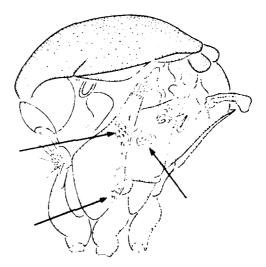
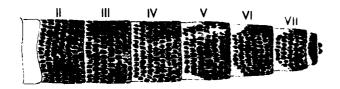


Fig. 379 — Lateral view of thorax - Cx. salmarius



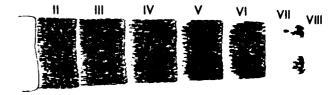


Fig. 378 — Dorsal view of abdomen - Cx. nigripalpus

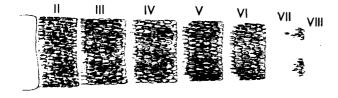
Fig. 380 — Dorsal view of abdomen - Cx. salinarius

12(11). Abdominal tergum VII mostly with dingy yellow scales; terga II-VI with only basolateral patches or with narrow, basal bands of dingy yellow scales, sometimes blended with similar scales on apex of previous segment (Fig. 381)salinarius

(Plate 35)

Abdominal tergum VII mostly with dark scales; terga II-VI with only basolateral patches

(Plate 34)



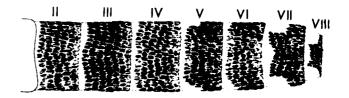


Fig. 381 — Dorsal view of abdomen - Cx. salinarius

Fig. 382 — Dorsal view of abdomen - Cx. chidesteri

13(10). Basal pale bands of abdominal terga rounded posteriorly, with marked sublateral constrictions, narrowly joined to lateral pale patches (Fig. 383); scutum without pale-scaled spots (Fig. 384) pipiens

quinquefasciatus (Plate 36)

Basal pale bands of abdominal terga not rounded posteriorly, broadly joined to lateral pale patches with only slight sublateral constrictions, most evident on tergum IV (Fig.

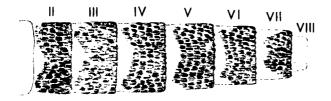


Fig. 383 — Dorsal view of abdomen - Cx. pipiens

Fig. 385 — Dorsal view of abdomen - Cx. restuans

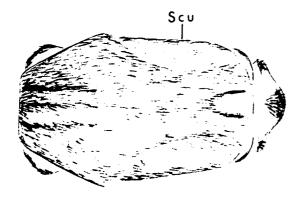


Fig. 384 — Dorsal view of thorax - Cx. pipiens

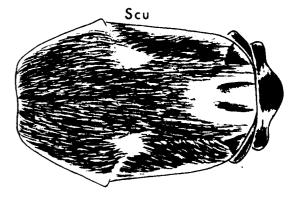


Fig. 386 — Dorsal view of thorax - Cx. restuans

14(13).	Wing cell R_2 4.5 or more length of vein R_{2+3} (Fig. 387); scutum usually with pair of pale,	
	submedian spots (Fig. 388); medium-sized species, wing length 4.0 mm or greaterrestuans	S
	(Plate 38)
	Wing cell R_9 about 3.0-4.0 length of vein R_{9+3} (Fig. 389); scutum without pale spots (Fig.	
	390); small species, wing length 2.8 mm or less	٠.,
	(Plate 32	

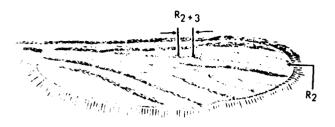


Fig. 387 — Dorsal view of wing - Cx. restuans



Fig. 389 — Dorsal view of wing - Cx. interrogator

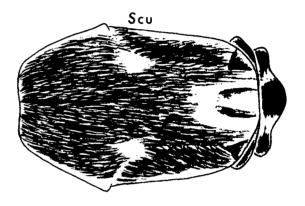


Fig. 388 — Dorsal view of thorax - Cx. restuans

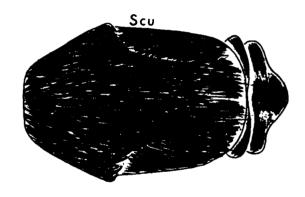


Fig. 390 — Dorsal view of thorax - Cx. interrogator



Fig. 391 — Dorsal view of abdomen - Cx. reevesi

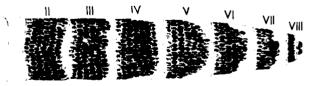


Fig. 392 — Porsal view of abdomen - Cx. territans



Fig. 393 — Dorsal view of abdomen - Cx. territans



Fig. 395 — Dorsal view of abdomen - Cx. arizonensis

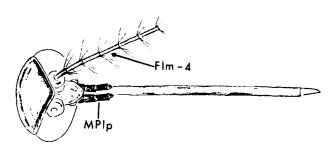


Fig. 394 — Dorsal view of head - Cx. territans

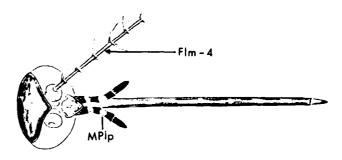


Fig. 396 — Dorsal view of head - Cx. apicalis



Fig. 397 — Dorsal view of wing - Cx. territans

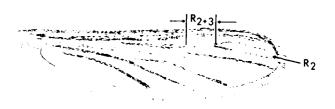


Fig. 399 — Dorsal view of wing - Cx, boharti



Fig. 398 — Dorsal view of abdomen - Cx, territans

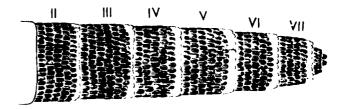
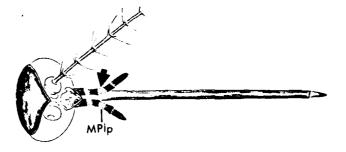


Fig. 400 — Dorsal view of abdomen - Cx, boharti



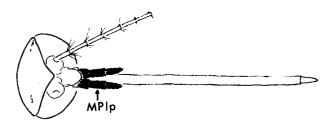
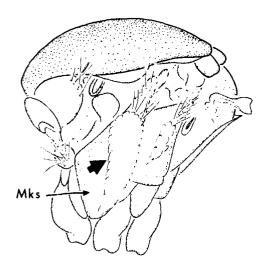


Fig. 401 — Dorsal view of head - Cx. apicalis

Fig. 402 — Lateral view of head - Cx. arizonensis



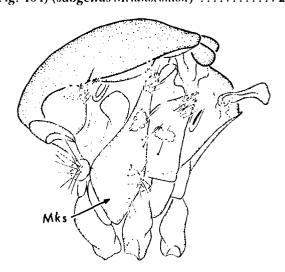


Fig. 403 — Lateral view of thorax - Cx. latisquama

Fig. 404 — Lateral view of thorax - Cx. pipiens

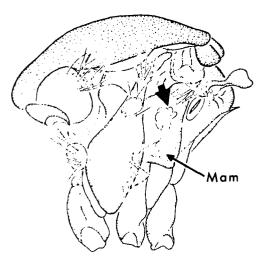


Fig. 405 - Lateral view of thorax - Cx. erraticus

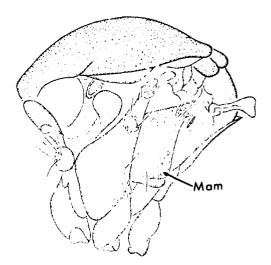
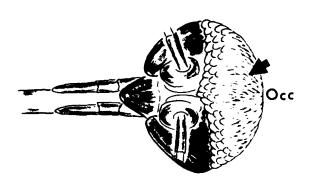


Fig. 406 — Lateral view of thorax - Cx. peccator

	sternum with patch of more l area (Fig. 407)				22
	sternum without scales or w gumental area (Fig. 408)				24
Mks	Mam	Mks —			Mam
Fig. 407 — Lateral view	of thorax - Cx. peccator	Fig. 408 —	Lateral view	of thorax -	Cx. atratus
base to apex (Fig.	ands on joints of tarsomeres 409) ntirely dark-scaled (Fig. 410			• • • • • • • • • • • • • • • • • • • •	(Plate 36)
		7			
A SECTION AND ADDRESS OF THE PARTY OF THE PA	Ta ₁	Ta ₂	Ta₃	Ta₄	Ta ₅
	Fig. 409 — <i>Hin</i>	dleg - Cx. opisthopus			
WEST SEE STATE	Ta ₁	Ta ₂	Ta ₃	To ₄	īa ₅
	Fig. 410 — <i>Hi</i>	ndleg - Cx. peccator			•
23(22). Ecciput with broad	I, dingy white scales antero	medially (Fig. 411)			abominator (Plate 32)
Occiput with broad	l, dark brown scales antero	medially (Fig. 412)			peccator anips (Plates 38, 32)



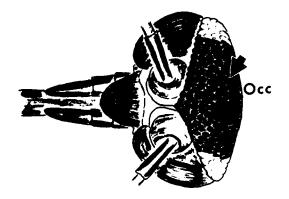
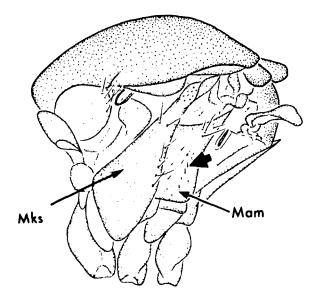


Fig. 411 — Dorsal view of head - Cx. abominator

Fig. 412 - Dorsal view of head - Cx. peccator

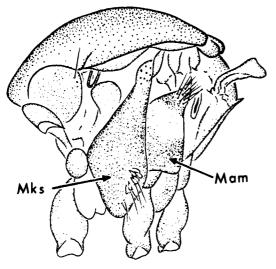


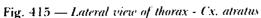
Mks Mam

Fig. 413 - Lateral view of thorax - Cx. iolambdis

Fig. 414 — Lateral view of thorax - Cx. atratus

	Mesanepimeron with distinct pale spot connected with anterior border, with dark area		
	ventrally continuous with dark, central area of mesokatepisternum (Fig. 415)atratu		
	(Plate 34		
	Mesanepimeron without distinct pale spot but with part of integument light in color (Fig.		
	416)		





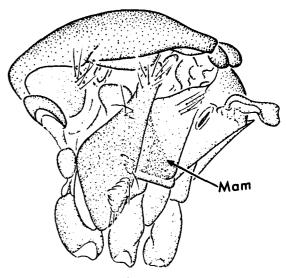


Fig. 416 - Lateral view of thorax - Cx. pilosus

26(25). Mesanepimeron with light integumental area covering upper 0.66 of sclerite; part of mesokatepisternum below ventral border of mesanepimeron with width/length ratio of

Mesanepimeron with light integumental area confined to narrow, pale border; part of mesokatepisternum below ventral border of mesanepimeron with width/length ratio of 1 to 1 (Fig. 418)mulrennani

(Plate 38)

(Plate 32)

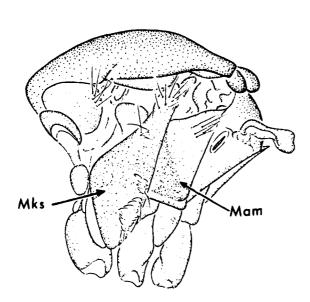


Fig. 417 — Lateral view of thorax - Cx. pilosus

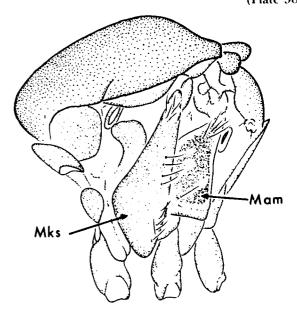


Fig. 418 — Lateral view of thorax - Cx. mulrennani

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS CULISETA

1. (Plate 42)

	Dorsum of abdomen with distinct basa	al, pale bands (Fig. 420)	
	I III IV V VI V	/II VIII	IV V VI VIII VIII
Fig.	419 — Dorsal view of abdomen - Cs. melan	nura Fig. 420 — Dorsal a	view of abdomen - Cs. morsitans
2(1).	Hindtarsomeres with pale-scaled band Hindtarsomeres unbanded (Fig. 422)		
	Ta ₁	Ta ₂	Ta₃ _
A	Fig. 421	1 — Hindleg - Cs. morsitans	Ta ₄ Ta ₅
1	Ta ₁	Ta₂	Ta ₃ To ₄
	Fig. 422	2 — Hindleg - Cs. impatiens	Ta ₅
3(2).	Hindleg with broad pale bands, covering wing with scales (Fig. 424)	g 0.25-0.33 of tarsomere 2 (Fig. 425	3); crossveins of
	Hindleg with pale bands narrow, covering without scales (Fig. 426)	ng 0.1 or less of tarsomere 2 (Fig. 42	25); crossveins
	Ta ₁	Ta ₂	Ta ₃ Ta ₄
44	Fig. 425	3 — Hindleg - Cs. particeps	la ₅
	Taı	Tα ₂	To ₃
	Fig. 495	— Hindleg - Cs. morsitans	Ta ₄ Ta ₅

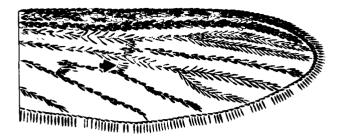


Fig. 424 - Dorsal view of wing - Cs. particeps

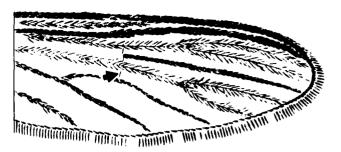
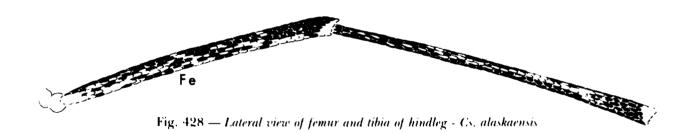


Fig. 426 - Dorsal view of wing - Cs. impatiens



Fig. 427 — Lateral view of femur and tibia of hindleg - Cs. particeps



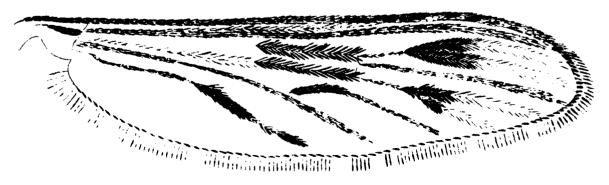


Fig. 429 — Dorsal view of wing - Cs. incidens

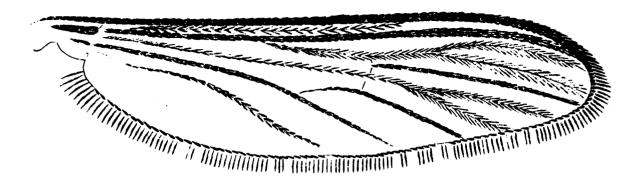
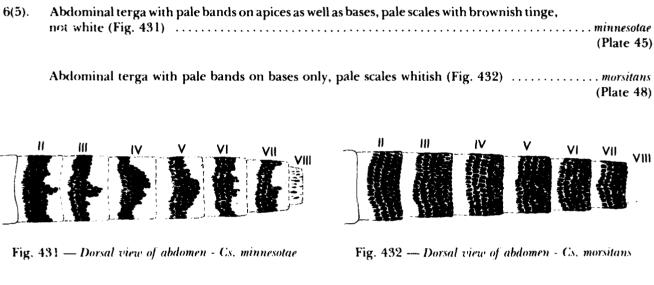


Fig. 430 - Dorsal view of wing - Cs. impatiens



7(2). Wing with dark and pale scales intermixed on anterior veins (Fig. 433); hindtarsomeres

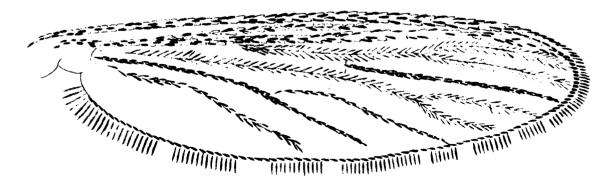


Fig. 433 - Dorsal view of wing - Cs. mornata

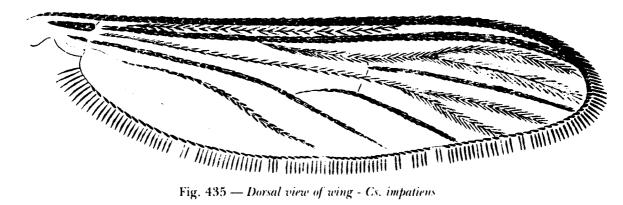


Fig. 435 - Dorsal view of wing - Cs. impatiens



Tα Ta_2

Fig. 436 — Hindleg - Cs. impatiens

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS DEINOCERITES

1. (Plate 45)

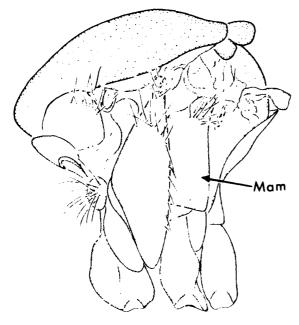


Fig. 137 - Lateral view of thorax De pseudes

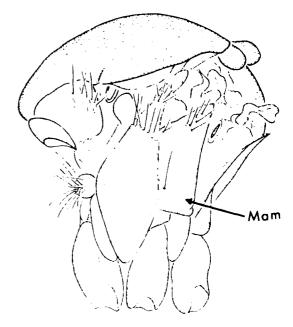


Fig. 438 = Lateral view of thorax Deceancer

2(1).	Cercus with 2 long, spatulate, apical or subapical setae (Fig. 439); medium-sized species,
	wing length about 2.9 mm
	(Plate 27)
	Cercus without specialized setae (Fig. 440); small species, wing length about 2.5 mm mathesoni

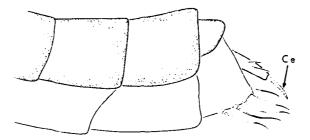


Fig. 439 — Lateral view of abdominal segments VII-X - De. cancer

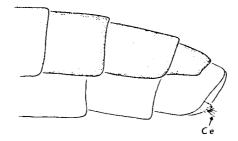


Fig. 440 — Lateral view of abdominal segments VII-X - De. mathesoni

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS MANSONIA

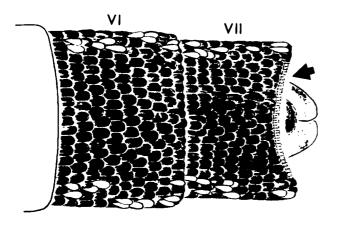
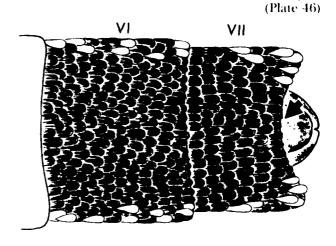


Fig. 441 — Dorsal view of tergum VII - Ma, titillans



(Plate 42)

Fig. 443 — Dorsal view of tergum VII - Ma. dyari



Fig. 442 — Ventral lateral view of head and proboscis - Ma. titillans

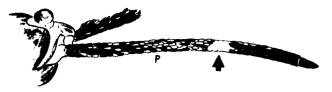


Fig. 444 — Ventral lateral view of head and proboscis - Ma. dyari

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS ORTHOPODOMYIA

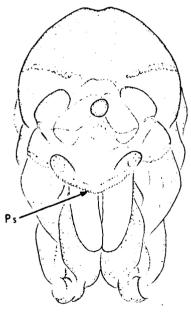


Fig. 445 — Anterior view of thorax - Or, kummi

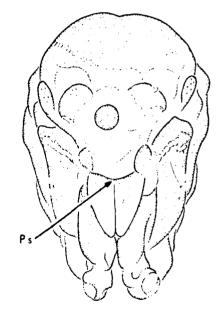


Fig. 448 — Anterior view of thorax - Or. alba



Fig. 446 — Dorsal view of wing - Or. kummi



Fig. 449 — Dorsal view of wing - Or, signifera



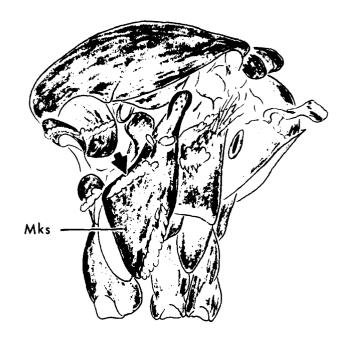


Fig. 447 — Lateral view of thorax - Or. kummi

Fig. 450 - Lateral view of thorax - Or. alba

Lower mesokatepisternal setae 0-2 (Fig. 453); base of vein R_{4+5} usually dark-scaled (Fig. 454) alba (Plate 41)

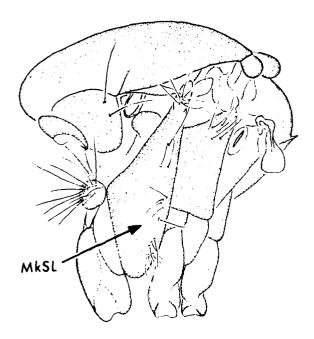


Fig. 451 — Lateral view of thorax - Or, signifera

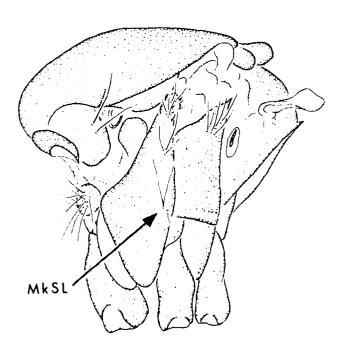


Fig. 453 — Lateral view of thorax - Or, alba

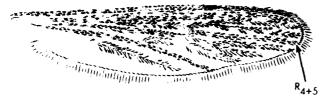


Fig. 452 - Dorsal view of wing - Or, signifera



Fig. 454 — Dorsal view of wing - Or, alba

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS PSOROPHORA



Fig. 455 — Dorsal view of wing - Ps. columbiae

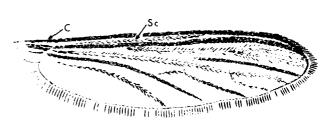


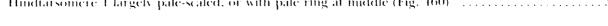
Fig. 457 — Dorsal view of wing - Ps. ciliata



Fig. 456 — Hindleg - Ps. columbiae



Fig. 458 — Hindleg - Ps. cyanescens



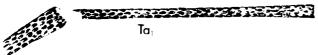






Fig. 459 -- Hindleg Ps. psgmace

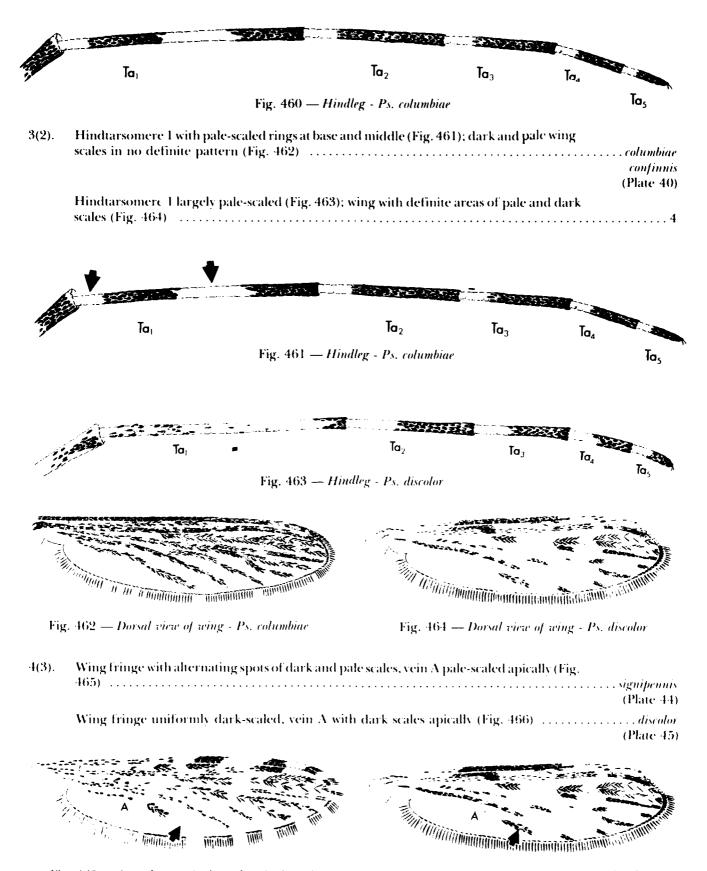
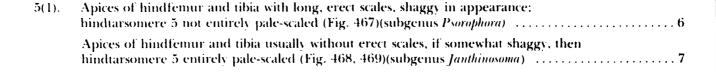


Fig. 465 — Dorsal view of wing - Ps. significants

Fig. 466 — Dorsal view of wing Ps. discolor



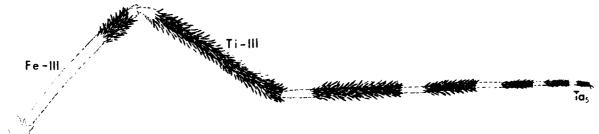
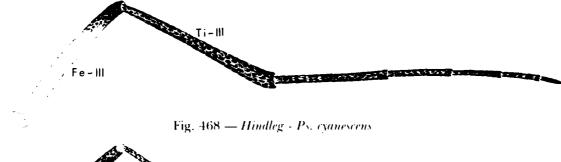


Fig. 467 — Hindleg - Ps. cilata



Ta.

Fig. 469 — Hindleg - Ps. ferox

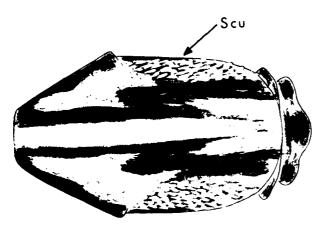


Fig. 470 — Dorsal view of thorax - Ps. ciliata

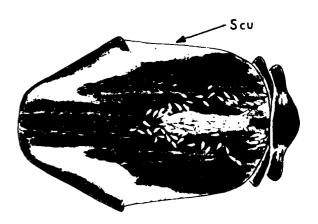


Fig. 472 — Dorsal view of thorax - Ps. howardii





Fig. 471 — Lateral view of head and proboscis - Ps. ciliata

Fig. 473 — Lateral view of head and proboscis - Ps. howardii

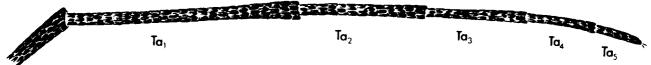


Fig. 474 — Hindleg - Ps. cyanescens

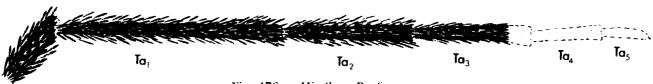


Fig. 476 — Hindleg - Ps. ferox

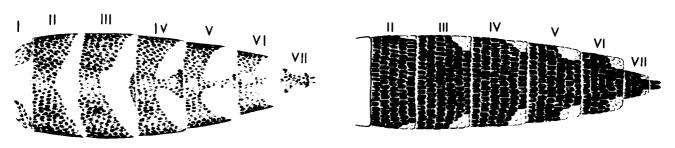


Fig. 475 — Dorsal view of abdomen - Ps. cyanescens

Fig. 477 — Dorsal view of abdomen - Ps. ferox



Fig. 478 — Hindleg - Ps. mathesoni

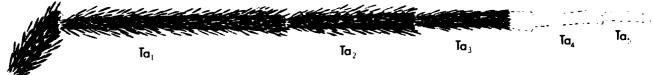


Fig. 479 — Hindleg - Ps. ferox

9(8).	Scutum entirely covered with yellowish-white scales (Fig. 480)	johnstonii
		(Plate 33)
	Scutum with broad, longitudinal, median stripe of dark scales, vellowish-white scales	
	laterally (Fig. 481)	10

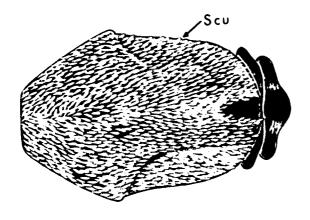


Fig. 480 - Dorsal view of thorax - Ps. johnstonii

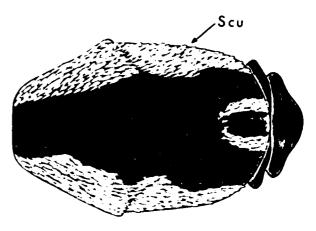


Fig. 481 — Dorsal view of thorax - Ps. mathesoni

10(9).	Subspiracular area with few or no scales (Fig. 4	82)
	Subspiracular area with many scales (Fig. 483)	varipes

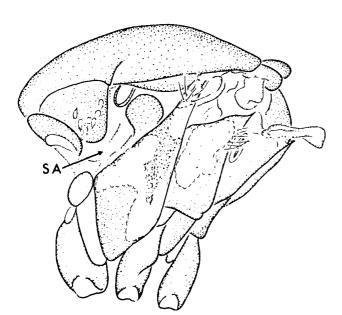


Fig. 482 — Lateral view of thorax - Ps. mathesoni

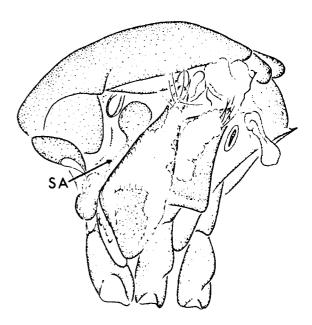


Fig. 483 — Lateral view of thorax - Ps. varipes

11(8).	Hindtarsomere 5 pale-sca	ded, others dark-scaled (Fig.	484)		mexicana (Plate 39)
	Hindleg with tarsomeres	4.5 and often part of 3 pale-	scaled (Fig. 485)		12
	Ta	the state of the s	THE PERSON NAMED IN	Ta ₄	■
Á	la,	iu ₂	Ja ₃	To₄	Ta _s
	•	Fig. 484 — Hindleg - P	s, mexicana		
	di -				

 $\mathbf{T}_{\mathbf{G}_1}$ $\mathbf{T}_{\mathbf{G}_2}$ $\mathbf{T}_{\mathbf{G}_3}$ $\mathbf{T}_{\mathbf{G}_4}$ $\mathbf{T}_{\mathbf{G}_5}$

Fig. 485 — Hindleg - Ps. ferox

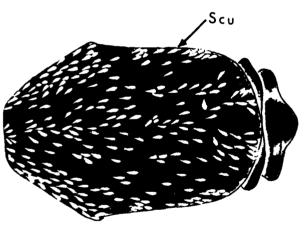


Fig. 486 - Dorsal view of thorax - Ps. ferox

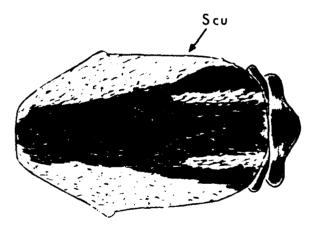


Fig. 488 — Dorsal view of thorax - Ps. horrida

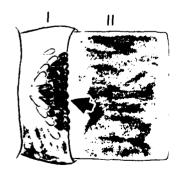


Fig. 487 — Dorsal view of abdominal segments I-II - Ps.

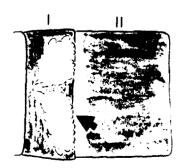


Fig. 489 — Dorsal view of abdominal segments I-II - Ps. horrida

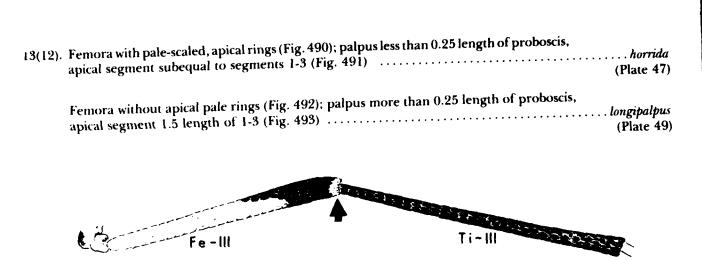


Fig. 490 - Hindleg - Ps. horrida

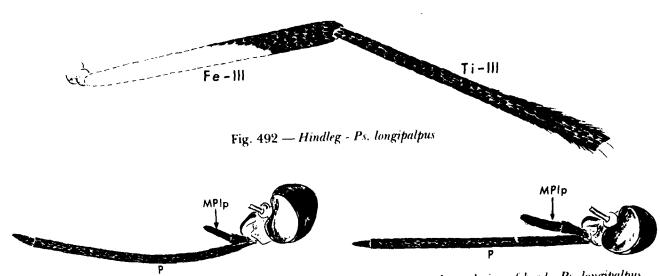


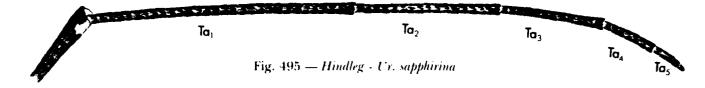
Fig. 491 - Lateral view of head - Ps. horrida

Fig. 493 - Lateral view of head - Ps. longipalpus

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS URANOTAENIA

1.





2(1).	Narrow, median, longitudinal stripe of scutum and midlobe of scutellum with iridescent
	blue scales (Fig. 496)
	(Plate 49)

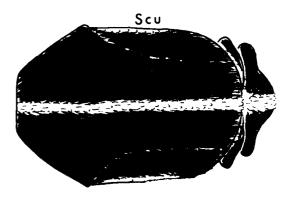


Fig. 496 — Dorsal view of thorax - Ur. sapphirma

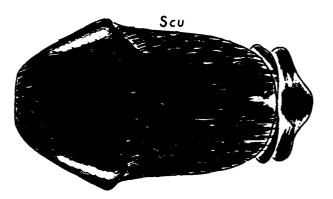


Fig. 497 — Dorsal view of thorax - Ur. a. anhydor

3(2).	Scutum with lateral line of iridescent blue scales incomplete, broken above mesothoracic spiracle (Fig. 498)
	(Plate 42)
	Scutum with continuous lateral line of iridescent blue scales from anterior promontory to
	wing base (Fig. 499)
	(Plate 42)



Fig. 498 - Dorsolateral view of thorax - Ur. a. anhydor

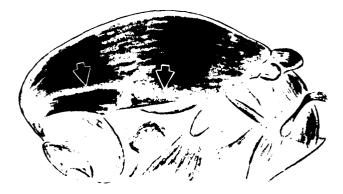


Fig. 499 — Dorsolateral view of thorax - Ur. a. syntheta

KEY TO ADULT FEMALE MOSQUITOES OF THE GENUS WYEOMYLA

1.	pale scales posteriorly (Fig. 501)vanduzeei
	(Plate 46)
	Antepronotum with mostly bluish to purplish scales (Fig. 502); hindtarsomeres with or
	without basal patches of pale scales (Fig. 503)

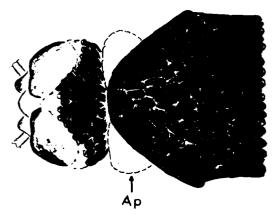


Fig. 500 - Dorsal view of thorax - Wy, vanduzeei

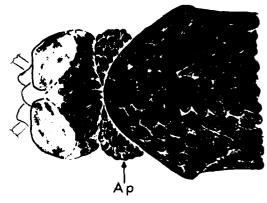


Fig. 502 - Dorsal view of thorax - Wy. smithii

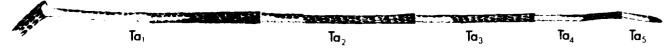
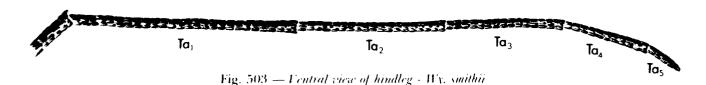


Fig. 501 - Ventral view of hindleg - Wy. vanduzeei



Occiput with dark scales along ocular line (Fig. 506); postpronotum with overlapping, dark scales (Fig. 507)

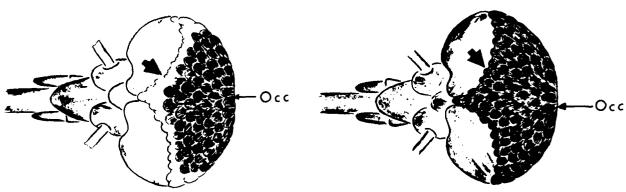
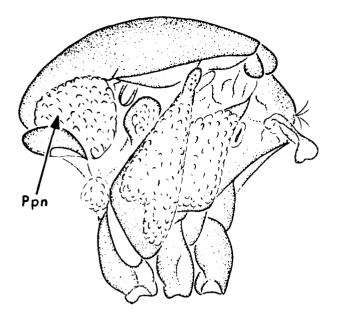


Fig. 504 — Dorsal view of head - Wy, mitchellii

Fig. 506 - Dorsal view of head - Wy, smithir



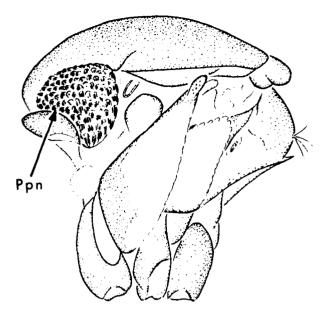


Fig. 505 — Lateral view of thorax - Wy. mitchellii

Fig. 507 — Lateral view of thorax - Wy, smithii

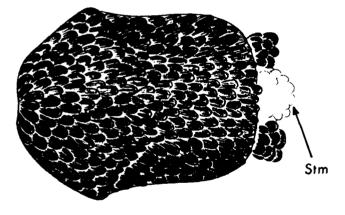


Fig. 508 - Dorsal view of thorax - Wy. haynei

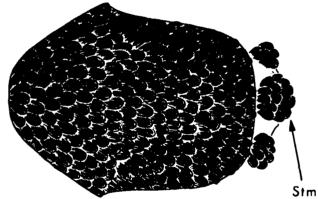


Fig. 509 - Dorsal view of thorax - Wy, smithii

MORPHOLOGY OF FOURTH STAGE MOSQUITO LARVA^{1,2}

The fourth stage mosquito larval body, contrary to the adult, is largely composed of soft, membranous tissue, but with some parts consisting of hardened, sclerotized plates. This allows for the characteristic swimming movements and doubling of the body when cleaning the lateral palatal brushes. The body is divided into the head, thorax and abdomen. The head capsule is completely sclerotized, while the thorax and abdomen are largely membranous. The larval body is adorned with some 190 pairs of setae (Plates 5, 6), a study of the arrangement of which is called chaetotaxy. These, along with various kinds of spicules are known collectively as the vestiture, i.e., protrusions from the cuticle of the integument, the covering of the body (Harbach and Knight 1978C), and are thus defined as cuticular projections. The organization and nomenclature of these structures is very important to know in larval identification. A complete treatment of the vestiture in general and the chaetotaxy in particular may be found in articles by Knight and Laffoon (1971B), and Harbach and Knight (1978A, 1978C). However, only those structures used in the present keys will be defined herein.

VESTITURE - The two main components of the larval vestiture are spicules and setae (synonyms: hairs, hair tufts, bristles). In larvae whose thorax and/or abdomen are sparsely or densely covered with a pubescence, the spicules are called aculeae, and the cuticle is aculeate (Fig. 672). Without this pile the surface would be smooth or glabrous. Where parts of a structure bear thornlike spicules, varying from tiny to very coarse, they are termed aciculae, and the condition is known as aciculate (Fig. 903). The lateral aspect of abdominal segment VIII, and also the siphon, in many kinds of mosquito larvae, bear specialized projections (Plates 6, 8). On abdominal segment VIII the structures are known as **comb scales** (CS) and they usually bear along their free posterior border a fringe of subequal spinules, or a median, large spine and lateral, smaller spinules. The **pecten** (Pt) is a comb-like row of spines, borne on a **pecten plate** (PP) in anophelines and posterolaterally on the **siphon** (S) in most culicine species. Each unit may bear one to many lateral denticles on one, or less frequently, both margins (Fig. 856). In subgenus *Psorophora*, the **pecten spines** (PS) are extended apically into long filaments (Fig. 931).

Setae may be distinguished from spicules by the presence of a basal alveolus from which the seta arises, (see Plate 7A - L). The **alveolus** is a membranous 800ket located in the integument, allowing the seta movement. Setae may be found attached to the selecotized structures, such as the head, siphon and saddle, or directly to the membranous integument of the larval body. At times, the membrane may bear a special sclerite, to which one or more setae are attached, called a setal support plate. Setae can be simple, unbranched or variously branched. Unbranched setae (A) are usually cylindrical and attenuated apically. They can also be very thick and spinelike, in which case they are called **spiniform setae** (B). **Branched setae** are composed of a main stem and ramifying members (F). In some, the branches arise directly from the base and therefore have no stem or an extremely short one. Those which have only a few branches arising beyond the basal third of the main stem are termed forked (E), while those with a very stout stem and many branches are called fanlike setae (L). Setae with numerous, regularly arranged branches arising on either side of the stem are plumose (G). When setae have various stems with branches that are divided and subdivided so that they resemble the branches on a tree, they are known as dendritic (H). Specialized seta, characteristic of the genus Anopheles, have flattened moveable branches usually radiating horizontally from a short, stout stem and are named **palmate** (1, 1). The branches are known as leaflers which can have smooth or serrate margins. The flat surface of the leafler is the blade and it may have a terminal **filament.** They may be fully developed (I) or partially formed (I). The lateral palatal brushes (mouth brushes) are composed of unique specialized spicules, termed **comb-tipped filaments** (K), bearing a row of rigid processes apically on one side, like the teeth of a comb. Special mention needs to be made of seta 4 of abdominal segment X, a group of setal tufts. known as the ventral brush, see Figs. 670, 671. In most mosquito larvae, it is composed of a row of fanlike setae, some or all of which are usually attached to a heavily selectorized network of bars called the grid (G); see Plate 8A. It is composed of a number of transverse grid bars (TGB) connected to lateral grid bars (LGB). In some cases, the setae are joined to a setal support plate,

The use of stage instead of instar follows Jones (1978)

Tot references cited herein, see Selected Bibliography of Mosquito Morphology at the end of section on Morphology of Adult Lemale.

known as a boss. Those setal tufts attached to the grid or boss are called cratal setae and those which are attached to the segment anterior to the cratal setae and grid are the precratal setae.

Single, or the components of branched, setae may be smooth, or spiculate. Their parts may be beset with, thin needlelike processes, which may vary in thickness. This condition is known as **aciculate** (C), whereas if the processes are small and spinelike, it is **spinulate** (D). If the setal parts have no processes, they are called **smooth.**

The following abbreviations will be used in a discussion of the morphology of the larval body regions (Plates 5, 6), and also in the larval keys.

A = antenna

C = head

IV = abdominal segment IV

V = prothorax

V = abdominal segment V

V = abdominal segment V

VI = abdominal segment VI

VII = abdominal segment VIII

Abdominal segment VIII

VIII = Abdominal segment VIII

Abdominal segment VIII

S = abdominal segment VIII

S = siphon

HEAD

The head is composed of a sclerotized capsule, bearing the mouthparts and antennae anteriorly and the occipital foramen, the opening of the cranium to which the cervix is attached, posteriorly. The shape of the head is distinctive in some mosquito larvae. Most have an ovate head, wider than long, with the greatest width at the level of the eyes. In the genus *Deinocerites*, the head is rather triangular, with the greatest width anteriorly at the level of the bases of the antennae. In the genus *Uranotaenia*, larval heads are thin, longer than wide, while in the predatory larvae of the genera *Toxorhynchites* and *Psorophora* heads are quadrate-shaped.

For a few species the integument of the **dorsal apotome** (frontoclypeus), the large sclerite forming the dorsal aspect of the head, contains patterns of pigment which may be diagnostic. To evaluate this character correctly, the larval head must be examined under low magnification.

The mouthparts will not be discussed here. For their descriptions and understanding, consult Gardner et al. (1973), Harbach (1977, 1978), Harbach and Knight (1977A, 1977B, 1977C), Knight (1971B), Knight and Harbach (1977), Pao and Knight (1970A, 1970B), Pucat (1965), and Shalaby (1956, 1957A, 1957B). Dorsolateral to the mouthparts, of which the mandibles and maxillae are most obvious externally, is a lobe which bears a large brush formed of specialized spicules. The lobe is composed of the **lateral tormal process** and the **lateral palatal plate** and the brush, the **lateral palatal brush** (=mouth brush). Usually the brush is made up of many comb-tipped filaments; but in the predatory larvae, they consist of a few, stout, prehensile spicules, see Fig. 520.

Setae of Head - On the head are found 16 pairs of setae, of which setae 2-C to 9-C are used in identification. The letter "C" is used to indicate that it is a seta located on the head. Formerly some of those setae were called by descriptive names, such as inner, outer and posterior clypeal hairs in anopheline larvae for setae 2-C, 3-C and 4-C and upper and lower head hairs in culicine larvae for 5-C and 6-C. In the keys that follow only numbers and letters or Roman numerals will be used in naming the larval setae.

The position of the setae in relation to one another is often used in identification. In anophelines, the two 2-C setae may be so close together that they have not the diameter of one of their alveoli separating them (Fig. 789); or they may be widely separated, closer to 3-C than to each other (Fig. 815). Two culicine species, *Ae. cinereus* and *Ae. hemiteleus* (Fig. 548), are distinguished in having setae 5-C, 6-C and 7-C in a straight line; while others of that genus have 6-C anteriorly out-of-line.

In several species, e.g. Ae. abservatus (Fig. 575), the setae of the head are very coarse, the diameters are about equal, extending almost to the apex; while in most larvae the setae are attenuated, gradually tapering apically. Usually 4-C is a weak, small seta, but in some species of the subgenus *Protomacleaya*, e.g. Ae. triseriatus (Fig. 692), it is a well developed, many branched seta.

In many instances, the size of the seta or the relative size of one in comparison to another, the number of branches, the manner of branching, the presence or absence of aciculae are all used as diagnostic characters. In some cases, the individual branches may be unequal, some shorter than others, e.g., *Ps. longipalpus* (Fig. 965). The setae 5-C and 6-C of larvae in the subgenus *Uranotaenia* are unique. They are very stout spiniform setae with spinulate surfaces.

Antennae - The antenna is a cylindrical, sensory appendage attached anterolaterally to the head. It bears six setae, 1-A to 6-A. In the genera Coquillettidia and Mansonia, the antenna is modified. It has an additional segment distal to the point of attachment of setae 2-A and 3-A, called here the flagellum, after Wharton (1962), see Figs. 514, 516. Another unique variation of the antenna is its sinuate, inflated shape in Ps. discolor (Fig. 932). In most species of the genus Culex (Fig. 833), the antenna is markedly constricted in the distal 0.33, beyond the attachment of seta 1-A. The antennal length is significant; in most species it is shorter than the head, but in some it is as long as or much longer than the head (Fig. 938). In the subgenus Psorophora the antenna is very small, hardly reaching the anterior margin of the head (Fig. 930). The surface of the antenna is usually beset with spinules, but may vary from none, as in Ac. triscriatus (Fig. 666), to a few small spinules, as in Ac. muelleri (Fig. 679), to many coarse spinules, as in Ac. fitchii (Fig. 667). Some of the 6 setae offer assistance in identification. The attachment of seta 1-A is diagnostic for some larvae. It may be near the middle of the antenna but may also occur in the basal 0.33 or distal 0.33 depending on the species. The number and size of the branches of 1-A also are used. In several cases, the size of setae 2-A and 3-A, as between species, or as compared with the size of 4-A is helpful, see Cq. perturbans, Figs. 514, 516.

THORAX

The thorax is an ovate unit of the body, somewhat wider than the head in well nourished, fourth stage larvae. As in the adult, it consists of the 3 segments, the pro-, meso- and metathorax. They are distinguished by the 3 distinct sets of setae, 0-P to 14-P on the prothorax, 1-M to 14-M on the mesothorax and 1-1 to 13-T on the metathorax. The integument of the thorax is sometimes aculeate. This is most easily detected by checking under the compound microscope the edges of the thorax on a vertical surface where debris, often found covering the body of mature larvae, does not seem to accumulate. The non-aculeate surface is called smooth or glabrous and is the more usual condition.

Of the 42 pairs of setae available on the thorax, only 10 are used in the larval keys. Setae 1-P, 3-P and 7-P have diagnostic size and or number of branches useful in separating species of several genera. In culicines setae 1-, 2-, 3-P are in a line, very close to one another; so it is hard to distinguish them. Likewise, often they are borne on a setal support plate; see Figs. 742, 803, 968. Seta 1-M is particularly useful in separating a number of Acdes larvae. In most it is a short seta, but in several it is long and stout. It is compared in the keys to the length of the antenna or to 2-M or 3-M. In the other thoracic setae, their number of branches or size are used.

ABDOMEN

The larval abdomen consists of 10 segments, each designated by the appropriate Roman numeral. The first 7 segments are very similar, segment I bearing 13 setae and II through VII, 15. Segments VIII-X are functionally specialized and morphologically different from the others. Segment IX does not exist as a distinct morphological unit, but is incorporated into VIII and X and will not be used in the keys.

In anophelines, abdominal segments I-VII possess a **tergal plate** anteriorly and may also have 1 or more **accessory tergal plates**, as in Figs. 818, 820. They do not ordinarily occur in culicine larvae, but some species of *Orthopodomyia* have well developed tergal plates on VII and VIII. *Uranotarina* and some *Psorophora* larvae have lateral selecties on VIII known as **comb plates**, to which the comb scales are attached; and the *Toxorhymhites* larvae have numerous small **setal support plates** on their thoracic and abdominal segments, a larger one laterally on VIII (Fig. 521).

Segments I-VII - Although there are 97 pairs of setae on abdominal segments I-VII, only 24 are used as key characters. Seta 1 is developed as a palmate type in some or all of abdominal segments I-VII of anophelines. The fully developed palmate setae usually have 10 or more large leaflets; and when one is in its normal position, it is spread to at least 180 degrees. The number of segments with fully formed palmate setae varies with the species. Segments I-III and VII sometimes have palmate setae not fully developed, which is expressed as 0.5 or 0.7 as large (Figs. 806, 807). Seta 6 (= lateral abdominal hair of authors) is used in a number of instances. It is usually a very prominent seta on each abdominal segment, especially on I-II. It is plumose on those 2 segments in anophelines and aciculate, commonly double or triple, in culicines. In 2 species of Anopheles, barberi and judithue, 6-1-VI are plumose. When seta 6 is more than single on segments III-VI, it is usually diagnostic for the species on which it occurs, e.g., Ae. taeniorhynchus, Fig. 598; Cx. peus, Fig. 845. Its size may also be characteristic, as in Ps. horrida (Fig. 958). Seta 0 is usually a tiny, single seta in anophelines, but in An. crucians (Fig. 800) it is well developed, with 4 or more branches. The other setae found on I-VII, employed in the keys are 2, 3, 7, and 13. Their size and number of branches are traits of certain species. In Ae. monticola, setae 1 and 13-IV-V are similar in size and number of branches; while in Ae. varipalpus, they are not. These 2 setae are located dorsoventrally opposite each other on the segment.

Segment VIII - Mosquito larvae are metapneustic, that is, the only functional external orifices of the respiratory system the spiracular openings (SOp), are located posteriorly on abdominal segment VIII; (see Plate 8). These openings are surrounded by the **spiracular apparatus** (SAp). In anophelines this structure is sessile; while in culicines it is borne on the end of a sclerotized tube, the siphon. There are only 5 setac on the segment, 1-VIII to 5-VIII. Laterally, in all larvae, except those of the genus Toxorhynchites, there occur the **comb scales** (CS). They may be arranged in a single row, double row, or in an irregular patch. There may be as few as 4, as in Ae. papago, Fig. 670, or as many as 70, as in Ae, pionips, Fig. 741. The total number, within ranges, is diagnostic and used throughout the keys. Among those larvae of the subgenus Melanoconion, Cx. abdominator has a short comb scale without a narrow elongation in the middle, while the others have a rather slippershaped scale, elongated and narrow in the middle; as in Cx. iolambdis, Fig. 883. The character of the median spine and the comparison of its size to that of the subapical spinules are extensively utilized. The size of the median spine ranges from only slightly larger than the subapical spinules, as in Ae, melanimon, Fig. 769, to very long, with tiny subapical and lateral spinules, as in Ae, riparius, Fig. 657. Extreme development of the median spine occurs in some larvae. In the subgenera Protomacleaya and Ochlerotatus, varitalpus group, the whole posterior projection of the comb scale is a rather blunt spine, fringed all along the edges with tiny spinules, Fig. 669. In Ae. nevadensis, larvac sometimes have 3 large, median spines (Gjullin et al., 1968, p. 135, Fig. 2C). In 4 species of Aedes, the subapical spinules are almost as stout as the median spine, e.g., Ae. thibaulti, Fig. 750; Ae. aegypti, Fig. 676.

Spiracular Apparatus - The spiracular apparatus (SAp) is a 5-lobed valve which closes the spiracular openings during submersion of the larva and protects them. The 5 lobes are: the **anterior spiracular lobe** (ASL), the two **anterolateral spiracular lobes** (L.3L) and two **posterolateral spiracular lobes** (PSL). They are moveable, flaplike projections and bear a total of 11 pairs of setae, 3-S to 13-S. Seta 6-S is unusually long in one species of *Psorophora* (Fig. 939). The posterolateral spiracular lobes are prolonged into taillike processes in one anopheline, see *An. pseudoponetipennis*. Fig. 794. In North America, the genera *Coquillettidia* and *Mansonia* have the spiracular apparatus highly modified for piercing the roots of certain aquatic plants, in which the larvae find a source of air. It is in the form of an attenuated tube, bearing hooklike teeth at the apex, the **inner and outer spiracular teeth** (IST, OST) and a row of teeth on the anterior surface, known as the **saw** (SAW) (Plate 8B). Such modified apparatuses possess 4 visible pairs of setae, 1-, 2-, 6-, and 8-S, according to Belkin (1962, Vol. 2, Figs. 198-204).

Siphon-The **siphon** (S) in culicines is one of the most useful structures in identification. Its size and shape vary considerably. The length/width dimensions are expressed by the **siphon index**. Harbach and Knight (1978A) have defined it as the ratio of the length of the siphon to the median width, but since so many descriptions of North American mosquito larvae have used the index as the ratio of the length to the basal width, it is being followed here. Actually, in most instances it makes very little difference; but for larvae of subgenera *Janthinosoma* and *Grabhamia*, where the siphon is swollen medially, measurements would be dissimilar. In the species treated here the index varies from 1.4 (*Ae. togoi*, Fig. 748) to 10.0 (*Cx. opisthopus*, Fig. 876). At the base of the siphon

is attached a small, lateral sclerite, the **siphon acus** (SA). In some species it is absent (*Ae. papago*, Fig. 670), while in others it is detached from the siphon "floating" in the basal membrane (*Ae. hendersoni*, Fig. 699).

Pecten- Five North American genera, Coquillettidia, Mansonia, Orthopodomyia, Toxorhynchites and Wycomyia have no pecten spines on the siphon; see Fig. 518. The **pecten spines** (PS) in the larvae of those genera bearing them are so variable as to offer good characters that are used extensively in the keys. A common variant is to have 1-4 of the distalmost spines more widely spaced than the others. In the keys they are termed "detached apically," e.g., Ae, excrucians, Fig. 606. The **pecten** (Pt) may be very short, with few spines, as in Ae, descritcola, Fig. 684, and Ps. columbiae, Fig. 933, or extend almost to the apex of the siphon, as in Ae, cataphylla, Fig. 608. The number of spines and the proportion of the siphon to which it extends from the base are used in the keys. In some species several apical spines are quite large; and their length is compared to the apical diameter of the siphon, as in Ae, fitchii, Fig. 704, or to the length of seta 2-S, as in Ae, campestris, Fig. 760. The pecten spine usually has 1-4 lateral denticles on its ventral edge, or less frequently on the dorsal edge, too; but their number varies from none in Cx. latisquama, Fig. 854, to about 20 in Cx. anips, Fig. 886.

The siphon may be adorned with other types of spicules. It may bear a large patch of aciculae apically, as in *Cx. bahamensis*, Fig. 824, or a set of spines near the apex, as in *Cx. coronator*, Fig. 834.

Siphonal Setae - The siphon ordinarily has 2 pairs of setae, 1-S and 2-S; however, when there are several setae present, the basalmost one is named 1a-S, then in sequence 1b-, 1c-, 1d-S, etc., proceeding distally (Belkin, 1950). Seta 2-S is small, preapical, and located anteriorly. It is called by Carpenter and LaCasse (1955) the dorsal preapical spine. Its length, curvature and presence or absence of a secondary branch are all useful characters; see Figs. 858, 859, 886, and 887. The position of 1-S with respect to the pecten is beneficial in separating groups of species in Aedes. Normally 1-S is attached distal to the apicalmost pecten spine. At times it is attached basal to the distalmost pecten spine, and it is described as being "attached within the pecten"; see Ae. tormentor, Fig. 566. The number of setae and their positions on the siphon are diagnostic in many species. Several species of Aedes, e.g., provocans, Fig. 539, have at least 1a-S to 1c-S. A trait of Culex larvae is the presence of 3 or more pairs of setae on the siphon. The total number is often characteristic, and in many instances the penultimate seta is dorsally out-of-line with the others (Fig. 826). They are also frequently in a straight line and in the subgenus Melanoconion have an additional one or more subdorsal, small setae, Fig. 856. The genus *Culiscia* has as its principal recognizing feature a pair of basal, ventrolateral setae, 1-8 see Plate 8A. Furthermore, species of the subgenus Culiseta have a row of short setae just distal to the pecten, Fig. 891. In some larvae, the siphonal setae are irregularly placed, e.g., Cx. restrians Fig. 830, Wy. smithii, Fig. 978. The length of seta 1-S is compared to many other structural dimensions, e.g., basal or apical diameter (Fig. 646), total length (Fig. 937), and distance from its alveolus to the apex of the siphon (Fig. 926). Likewise, its location at or distal to the middle of the siphon is peculiar to some larvae; see Ar. melanimon. Fig. 607. Of course, the numbers of branches of 1-S vary and are employed in the keys.

Segment X - This highly modified abdominal segment commonly called the anal segment, is the most posterior. It possesses a large sclerite, the **saddle** (Sa) which partially or entirely encircles the segment, usually 2 pairs of anal papillae, the homeostatic, almost transparent, cylindrical organs attached terminally to the segment, and 4 pairs of setae, I-X to 4-X.

Saddle - In most larvae there is a single saddle sclerite, but those of the genus *Deinocerites* bear small ones dorsally and ventrally. Of the remaining culicine genera, larvae of *Haemagogus*, *Wyeomyia*, some *Aedes* and species *bahamensis* of the genus *Culex* possess saddles which do not completely encircle segment X. It is often necessary to determine the extent to which the saddle encircles the segment. Some are small and do not extend even 0.5 the distance to the midventral line, e.g., *Ae. atropalpus*, Fig. 613, in which case seta 1-X is attached ventrally to the saddle. On the other hand, some species have very long, though incomplete, saddles, almost reaching to the midventral line, e.g., *Ae. princtodes*, Fig. 708. At times it is extremely difficult to determine the exact size of the saddle sclerite of larvae which have been mounted in Canada balsam for some years, because of clearing by the mountant. Very fine focusing by a compound microscope with 200-400X magnification will help to locate its ventral edge. Some saddles are deeply incised along the ventral margin, as in *Ae. enedes*, Fig. 647; and in a number of larvae of the genera *Aedes*, *Haemagogus*, and *Culiseta*, the saddles have prominent aciculae along the posterior border, which vary in size with the species; see Figs. 776, 779, 903.

Anal papillae - Of those species treated here, 2 have larvae with only one pair of anal papillae (APP), i.e., Cx. bahamensis, Fig. 825, and Wy. smithii, Fig. 980. Ae. dupreei larvae are unique for having very long anal papillae, about 8.0 the length of the saddle and darkly pigmented (Fig. 578). At the other extreme, those species with larvae which breed in brackish water have very small anal papillae; see Ae. taeniorhynchus, Fig. 599. It is customary to express the length of the anal papillae as a ratio with the length of the saddle. It is known as the anal papilla-saddle index and is computed by dividing the length of the papilla by the length of the saddle, i.e., its anteroposterior me surement along the middorsal line.

Setae - Setae of segment X provide differentiating characters. The length of seta 1-X, the saddle seta, is frequently used in the Aedes key, e.g., Fig. 754, 755. It is commonly compared with the saddle length. Setae 2-X and 3-X are known collectively as the dorsal brush; 2 is ordinarily multibranched and 3 long and single. Ae. abserratus larvae are unusual in that both these setae are long and single, Fig. 574. Seta 4 is composed of a variable number of paired and unpaired setae. The most posterior seta is designated as 4a; then proceeding anteriorly, they are 4b-, 4c-, 4d-X, etc. This group of setae acts as a rudder during swimming. It is particularly well developed in the larvae of the genus Psorophora, in which the numerous precratal fanlike setae usually extend anteriorly more than 0.5 the length of the segment (Fig. 528). Contrarily, it is poorly developed in those tree hole-inhabiting larvae belonging to subgenera Abraedes, Kompia, Protomacleaya and the varipulpus group of Ochlerotatus, as well as in those larvae of the genera Coquillettidia and Mansonia, which attach themselves to roots of plants. They have no more than 3 to 7 pairs of setae in the brush: see Figs. 700, 701, 922. In some of these larvae a boss is present for attachment of the setae instead of a grid; see Fig. 670. The number of branches in the 2 caudalmost setae (Ae. sierrensis, Fig. 680), or the 2 anteriormost setae (Ae. brelandi, Fig. 701) is diagnostic. The position of the ventral brush is important in distinguishing those Aedes larvae possessing a completely circular saddle. In them the setae are confined to that part of the segment posterior to the saddle. The total number of fanlike setae is distinctive for a number of species, e.g., Ar. zoosophus, Fig. 694, and Cs. minnesotar, Fig. 897. In Wycomyia larvae no regular, rudderlike ventral brush is present. Seta 4 is nothing more than a pair of long or short setae ventrolateroposteriorly on the segment; see Figs. 972, 974.

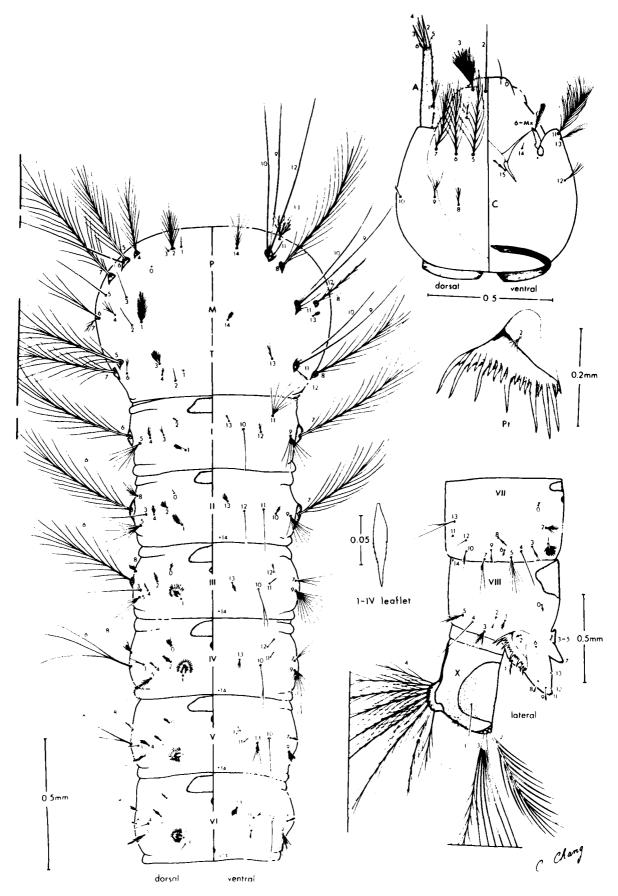


Plate 5. Fourth stage anopheline larva; dorsal left, ventral right.

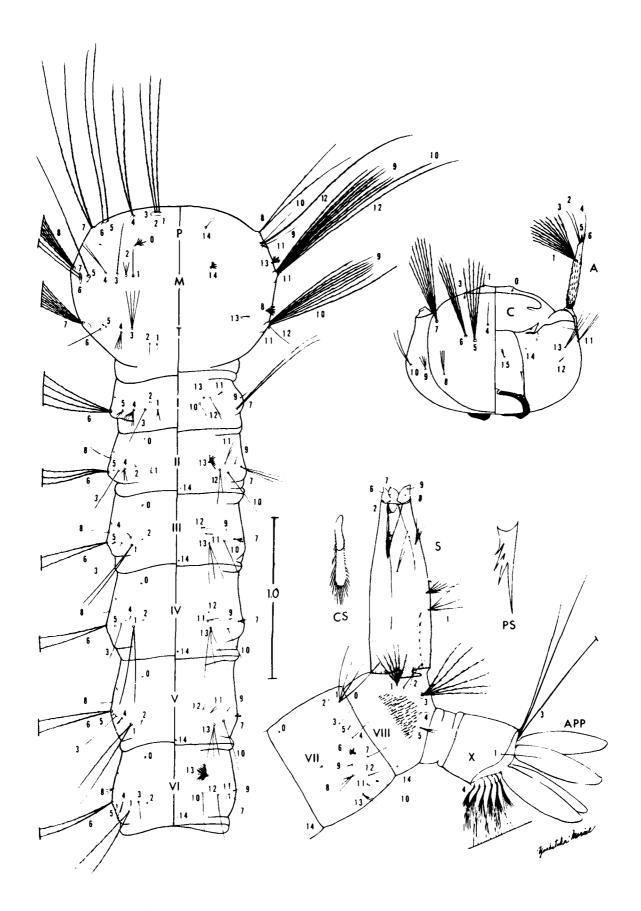


Plate 6. Fourth stage culicine larva; dorsal left, ventral right,

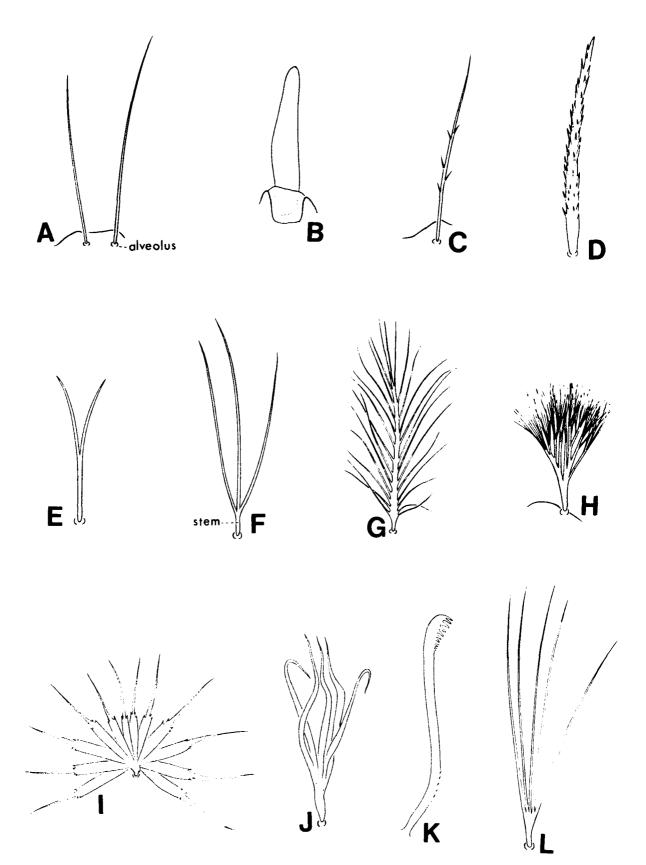


Plate 7. Examples of kinds of setae found in mosquito larvae. A. Unbranched smooth setae; B. Spiniform seta; C. Unbranched aciculate seta; D. Spinulate spiniform seta; E. Forked seta; F. Branched seta; G. Plumose seta; H. Dendritic seta; I. Palmate seta, fully developed; J. Palmate seta, 0.5 developed; K. Comb-tipped filament; L. Fanlike seta of ventral brush.

ABBREVIATIONS IN PLATE 8

APP - anal papilla

ASL - anterior spiracular lobe

ASLP - anterior spiracular lobe plate

C - comb

CS - comb scales

G - grid

IST - inner spiracular teeth

LGB - lateral grid bar

LSL - anterolateral spiracular lobe

LSLP - anterolateral spiracular lobe plate

MdP - median plate

OST - outer spiracular teeth

PP - pecten plate PS - pecten spines

PSL - posterolateral spiracular lobe

PSLP - posterolateral spiracular lobe plate

PSP - posterior spiracular plate

Pt - pecten

S - siphon

Sa - saddle

SA - siphon acus

SaA - saddle acus

SAd-spiracular apodeme

SAp- spiracular apparatus

SAW - saw

SOp - spiracular opening

TGB - transverse grid bar

VII - abdominal segment VII

VIII - abdominal segment VIII

X - abdominal segment X (anal

segment)

2-S - seta 2 of siphon

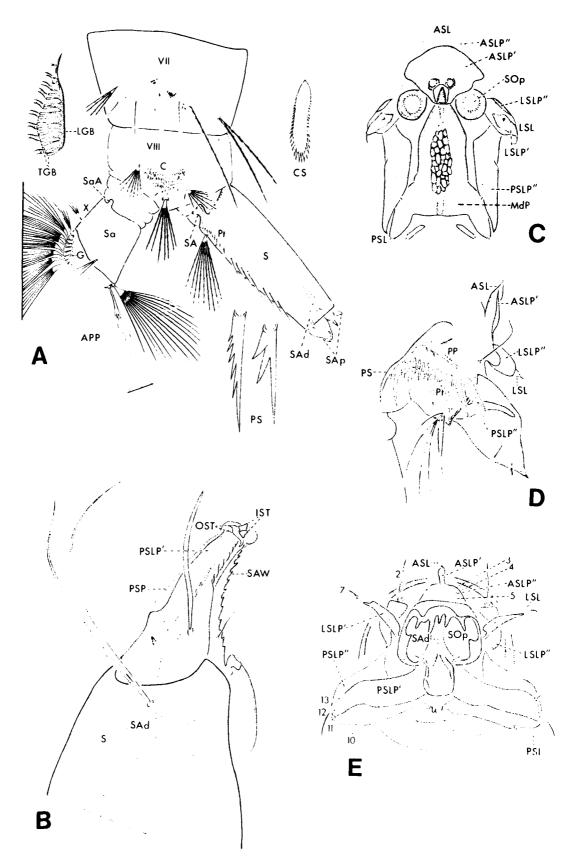
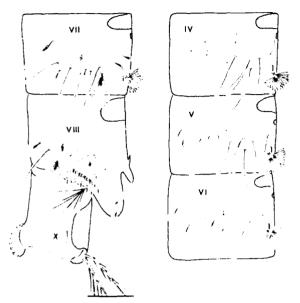


Plate 8. Morphology of terminal abdominal segments of mosquito larvae. A. segments VII-X of *Culiseta*; B. Siphon and spiracular apparatus of *Mansonia*; C.D. Spiracular apparatus of *Anopheles*; C. dorsal view, D. lateral view; E. Dorsal view of spiracular apparatus of *Culex*.

KEY TO GENERA OF FOURTH STAGE MOSQUITO LARVAE OF NORTH AMERICA, NORTH OF MEXICO



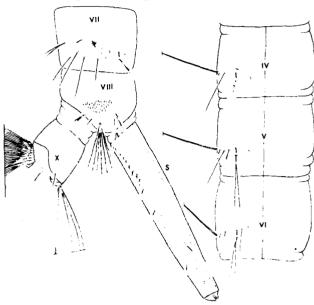


Fig. 510 — Lateral view of abdominal segments IV-X - An. quadrimaculatus

Fig. 511 — Dorsal and lateral view of abdominal segments IV-X - Cx. pipiens

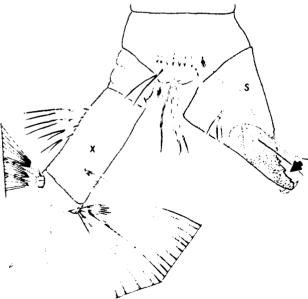


Fig. 512 — Lateral view of abdominal segments VIII-X - Ma. dyari

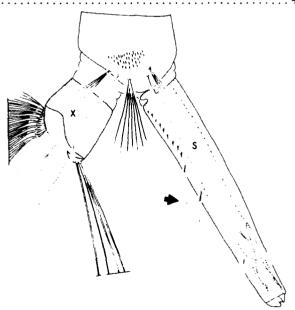


Fig. 513—Lateral view of abdominal segments VIII-X - Cx. pipiens

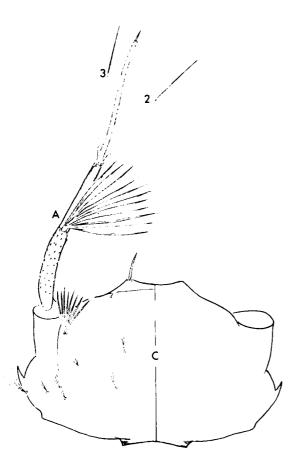


Fig. 514 - Dorsal view of head and antenna - Ma. dyari

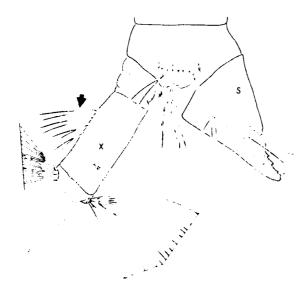


Fig. 515 — Lateral view of abdominal segments VIII-X - Ma. dyari

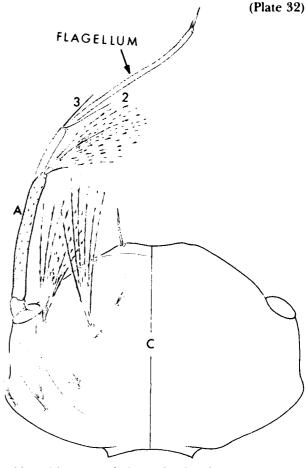
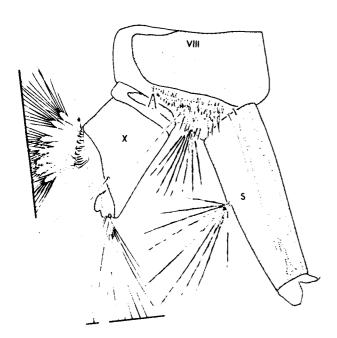


Fig. 516 — Dorsal view of head and antenna - Cq. perturbans



Fig. 517 — Lateral view of abdominal segments VIII-X - Cq. perturbans





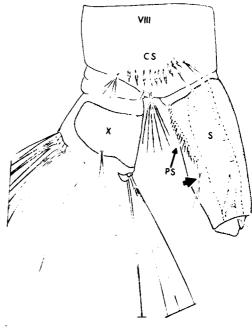


Fig. 518—Lateral view of abdominal segments VIII-X - Or. signifera

Fig. 519—Lateral view of abdominal segments VIII-X - Ae. aegypti

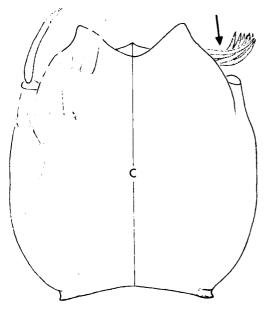


Fig. 520 — Dorsal view of head - Tx. r. septentrionalis

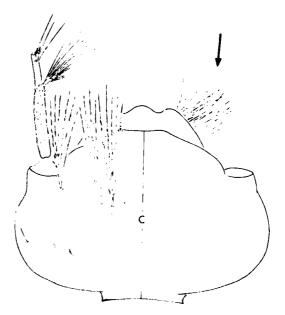
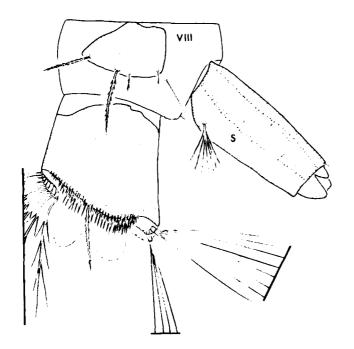


Fig. 522 — Dorsal view of head - Cx. pipiens



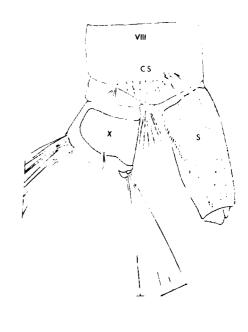


Fig. 521 — Lateral view of abdominal segments VIII-X - Tx. r. rutilus

Fig. 523—Lateral view of abdominal segments VIII-X - Ae. aegypti



Fig. 524 — Lateral view of abdominal segments VIII-X -Wy, smithii

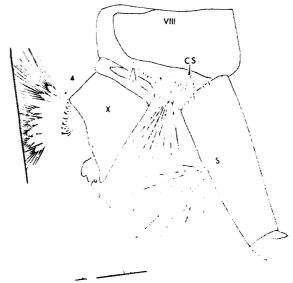


Fig. 525—Lateral view of abdominal segments VIII-X - Or. signifera

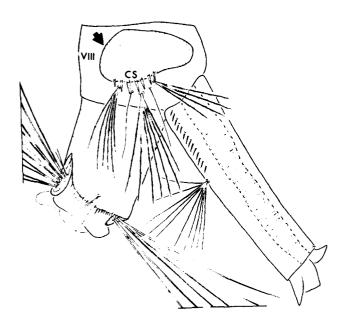


Fig. 526—Lateral view of abdominal segments VIII-X - Ur. sapphirina

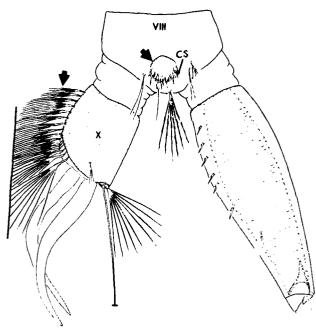


Fig. 528—Lateral view of abdominal segments VIII-X - Ps. columbiae

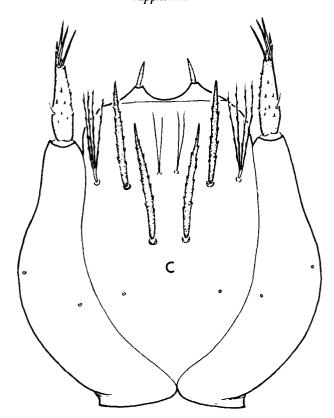


Fig. 527 — Dorsal view of head - Ur. sapphirina

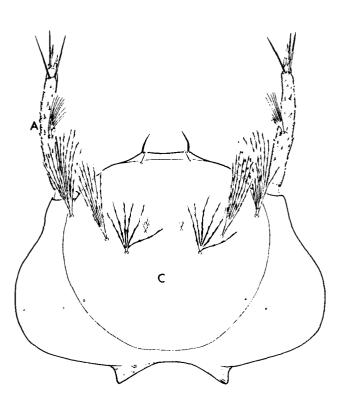


Fig. 529 — Dorsal view of head - Ps. columbiae

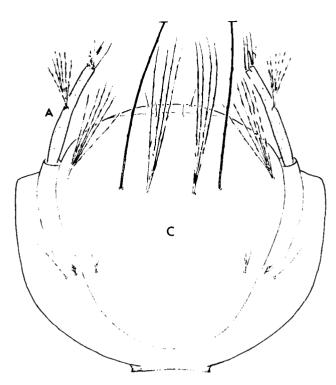


Fig. 530 - Dorsal view of head - De. pseudes

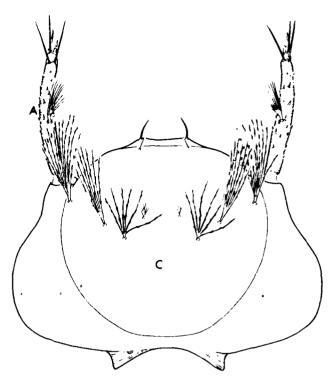


Fig. 532 — Dorsal view of head - Ps. columbiae

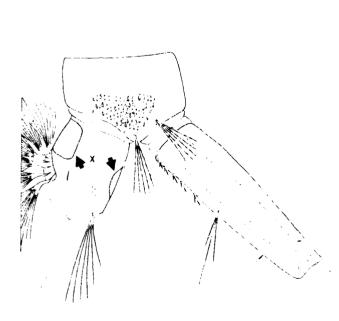


Fig. 531 — Lateral view of abdominal segments VIII-X - De. pseudes

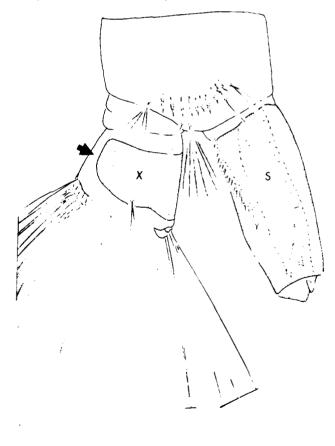


Fig. 533 — Lateral view of abdominal segments VIII-X - Acaegypti



VIII

CS

PS

Fig. 534 — Lateral wiew of abdominal segments VIII-X - Cs. inornata

Fig. 535—Lateral view of abdominal segments VIII-X - Ae. aegypti

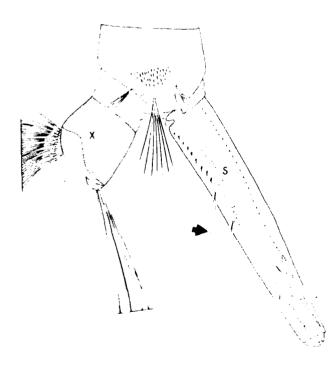
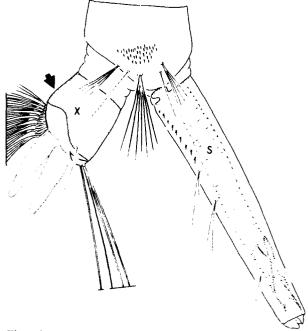


Fig. 536—Lateral view of abdominal segments VIII-X - Cx. pipiens



Fig. 537 — Lateral view of abdominal segments VIII-X · Ac. aegypti



VIII CS

Fig. 538—Lateral view of abdominal segments VIII-X - Cx. pipiens

Fig. 539—Lateral view of abdominal segments VIII-X - Ae. provocans

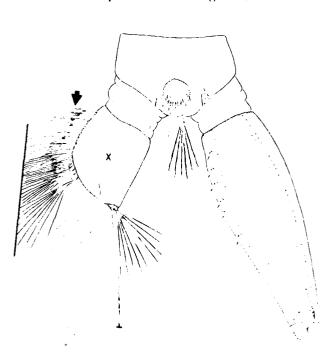


Fig. 540 — Lateral view of abdominal segments VIII-X Ps. columbiae

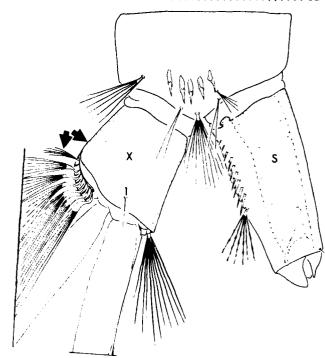
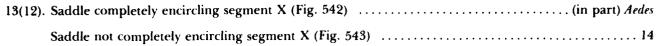


Fig. 541—Lateral view of abdominal segments VIII-X - Ac. atlanticus



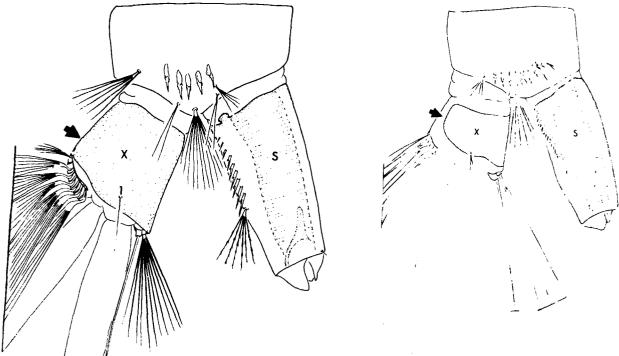


Fig. 542 — Lateral view of abdominal segments VIII-X - Ae. Fig. 543 — Lateral view of abdominal segments VIII-X - Ae. atlanticus aegypti

atlanticus

aegypti

14(13). Saddle bearing prominent aciculae on posterior border; seta 3 well developed, longer than

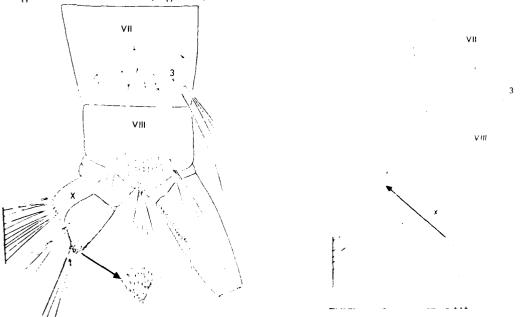


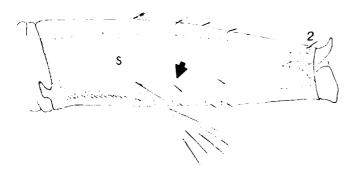
Fig. 544—Lateral view of abdominal segments VII/X/Hg. equinus

Fig. 545 — Lateral circle of abdominal segments VH Δ -to arguetts

(Plate 40)

KEY TO FOURTH STAGE LARVAE OF THE GENUS AEDES

1.	Siphon with more than 1 pair of setae, excluding seta 2-S (Fig. 546)	2
	Siphon with but 1 pair of setae, excluding seta 2-S (Fig. 547)	4



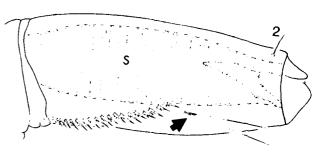
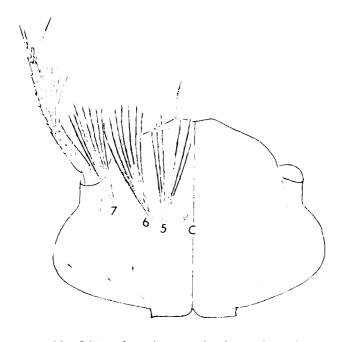


Fig. 546 — Lateral view of siphon - Ae, provocans

Fig. 547 — Lateral view of siphon - Ac. aegypti



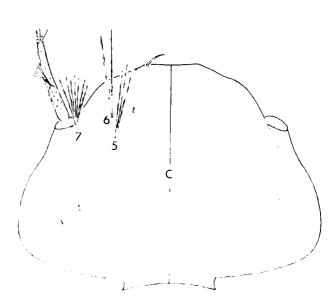
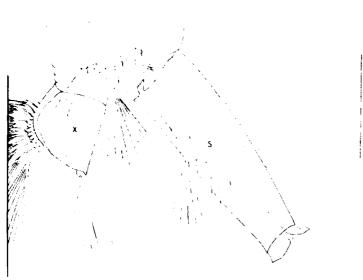


Fig. 548 - Dorsal crew of head - 4e hemiteleus

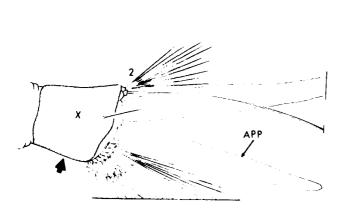
Fig. 549 - Dorsal crea of head. Ac, prococurs



CS S

Fig. 550—Lateral view of abdominal segments VIII-X-Av. provocans

Fig. 551—Lateral view of abdominal segments VIII-X - Ae. bicristatus



X

Fig. 552 — Lateral view of abdominal segment X - Ae, atlanticus

Fig. 553 — Lateral view of abdominal segment X - Ac.

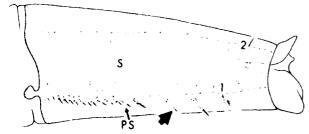


Fig. 554 — Lateral view of siphon - Ac. nigromaculis

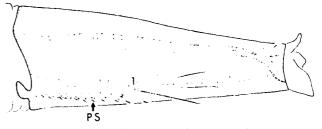


Fig. 555 — Lateral view of siphon - Ac. abservatus

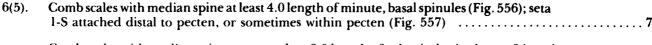




Fig. 556 — Comb scale - Ae. nigromaculis



Fig. 558 — Comb scale - Ae. f. pallens

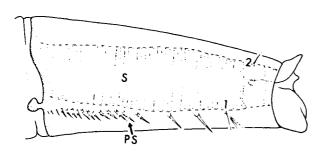


Fig. 557 — Lateral view of siphon - Ac. nigromaculis

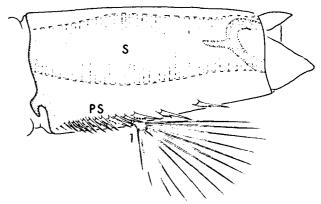


Fig. 559 - Lateral view of siphon - Ae. f. pallens

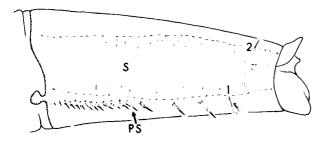


Fig. 560 - Lateral view of siphon - Ac. nigromaculis

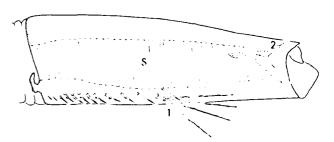


Fig. 561 — Lateral view of siphon - Ac. nigripes

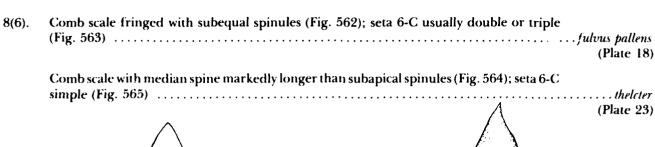




Fig. 562 — Comb scale - Ae. f. pallens



Fig. 564 — Comb scale - Ae, thelcter

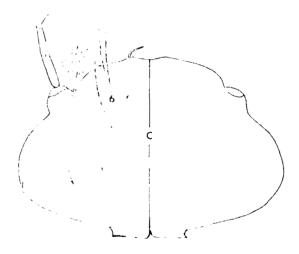


Fig. 563 — Dorsal view of head - Ae. f. pallens 9(5).

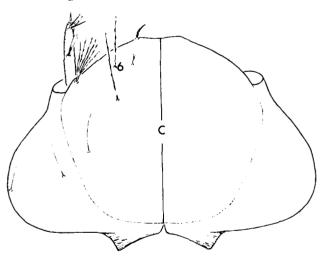


Fig. 565 — Dorsal view of head - Ae. theleter

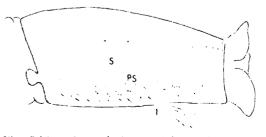


Fig. 566 — Lateral view of siphon - Ae. tormentor

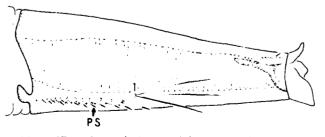
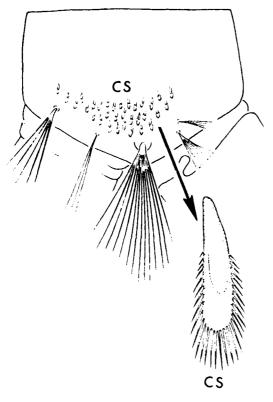


Fig. 567 — Lateral view of siphon - Ac. abservatus

10(9). Comb scales 30-40, evenly fringed with subequal spinules (Fig. 568)bimaculatus (Plate 16)

Comb scales 9-12, with large median spine and minute basal spinules (Fig. 569)tormentor (Plate 24)



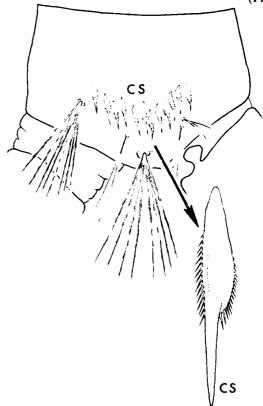


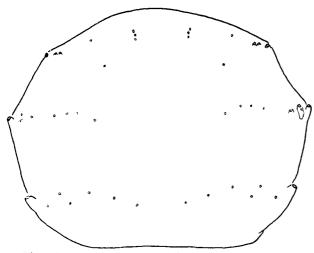
Fig. 568 — Lateral view of abdominal segment VIII - Ac. Fig. 569 — Lateral view of abdominal segment VIII - Ac. bimaculatus tormentor

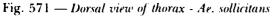


Fig. 570 Comb scale Accatlantions



CS Fig. 572 — Comb scale - Ac, tacniorhynchus





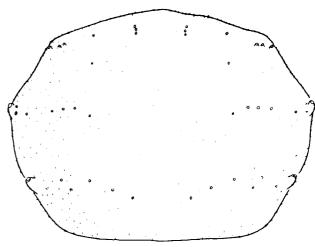


Fig. 573 - Dorsal view of thorax - Ae. taeniorhynchus

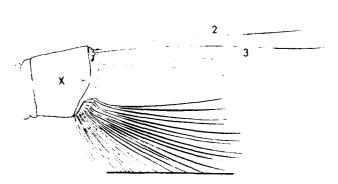


Fig. 574 — Lateral view of abdominal segment X - Ae. abservatus

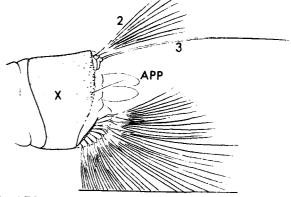


Fig. 576 — Lateral view of abdominal segment X - Ac. taeniorhynchus

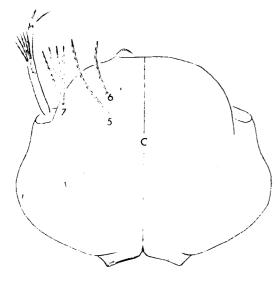


Fig. 575 — Dorsal view of head Ac. abserratus

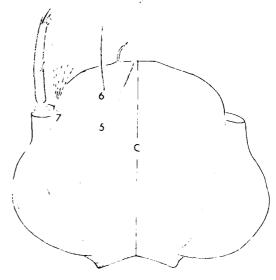


Fig. 577 — Dorsal view of head - Ac. taemorhynchus

13(12).	. Anal papilla-saddle index at least 8.0, papilla with darkly pigmented tracheae; seta 2-X with 2,3 branches (Fig. 578)	dupreei (Plate 17)
	Anal papilla-saddle index at most 5.0, usually much less, papilla lacking dark tracheae; seta 2-X with 4 or more branches (Fig. 579)	14



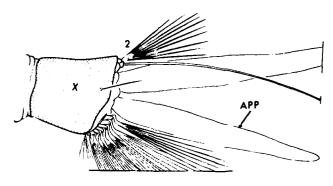
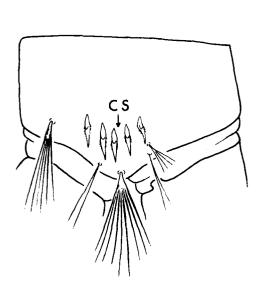


Fig. 578 — Lateral view of abdominal segment X - Ae. dupreei

Fig. 579 — Lateral view of abdominal segment X - Ae. atlanticus

14(13). Comb scales 4-9, large (Fig. 580)		15
Comb scales usually 10-30, small ((Fig. 581)	16



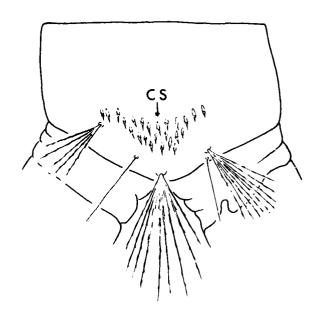
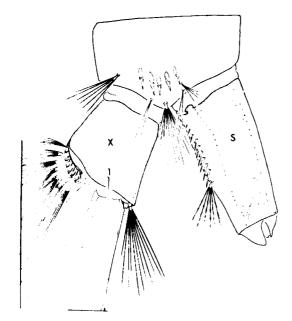


Fig. 580 — Lateral view of abdominal segment VIII - Ae. atlanticus

Fig. 581 — Lateral view of abdominal segment VIII - Ae. sollicitans

15(14). Siphon index about 2.0; seta 1-X shorter than saddle (Fig. 582)	
Siphon index about 3.0; seta 1-X equal to length of saddle or longer (Fig. 583)	



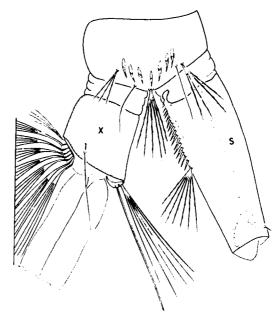
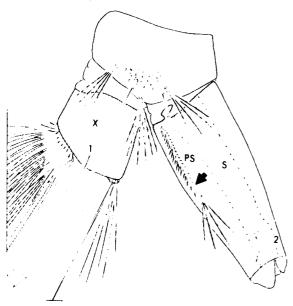


Fig. 582 — Lateral view of abdominal segments VIII-X - Ae. Fig. 583 — Lateral view of abdominal segments VIII-X - Ae. hexodontus

16(14). Seta 2-S much shorter than apical pecten spine; seta 1-X subequal to saddle (Fig. 584) punctor (Plate 13)



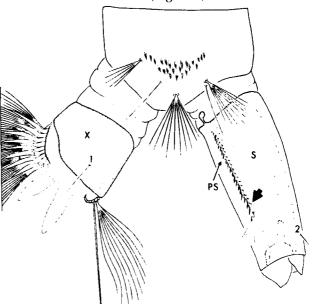


Fig. 584 — Lateral view of abdominal segments VIII-X - Ae. Fig. 585 — Lateral view of abdominal segments VIII-X - Ae. punctor sollicitans

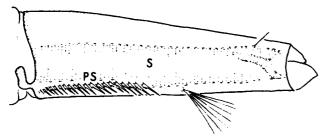


Fig. 586 — Lateral view of siphon - Ae. mitchellae

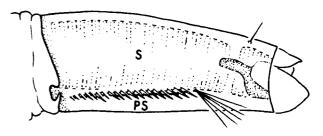


Fig. 588 - Lateral view of siphon - Ae. sollicitans

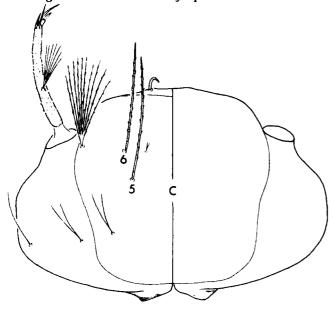


Fig. 587 — Dorsal view of head - Ae. mitchellae

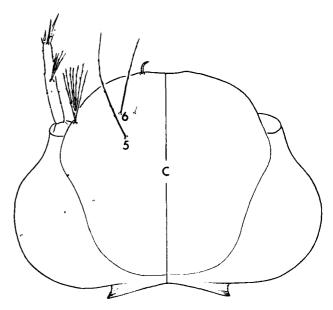


Fig. 589 — Dorsal view of head - Ae. sollicitans



Fig. 590 — Comb scale - Ae. infirmatus



Fig. 591 — Comb scale - Ae. taeniorhynchus

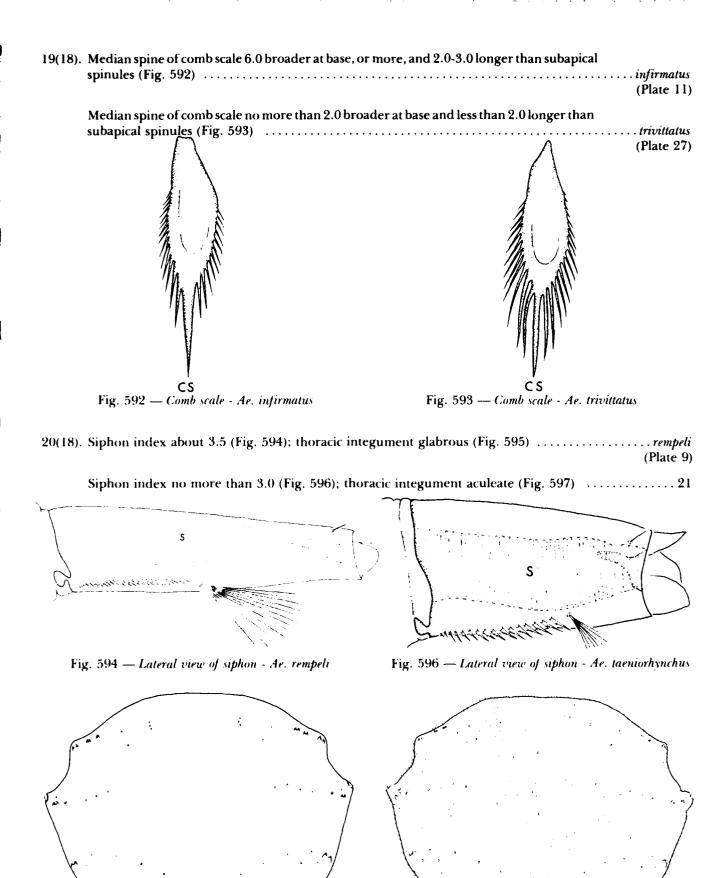


Fig. 595 — Dorsal view of thorax - Ae. rempeli

Fig. 597 — Dorsal view of thorax - Ae, scapularis

21(20). Seta 6-III-V with 2-5 branches (Fig. 598); anal papilla-saddle index 0.5 or less (Fig. 599) ... taeniorhynchus (Plate 9)

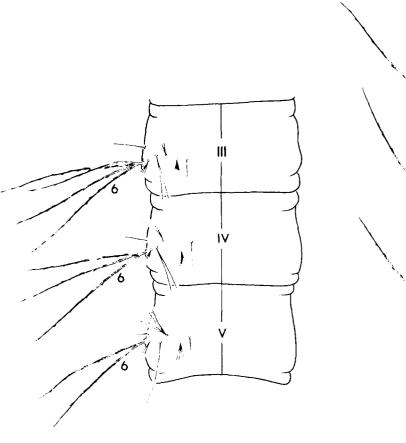


Fig. 598 — Dorsal view of abdominal segments III-V- Ae. taeniorhynchus

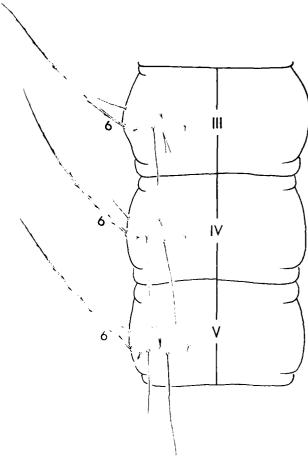


Fig. 600 — Dorsal view of abdominal segments III-V - Ae. scapularis

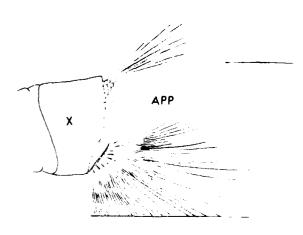


Fig. 599 — Lateral view of abdominal segment X. Ar. tacnio hynchus

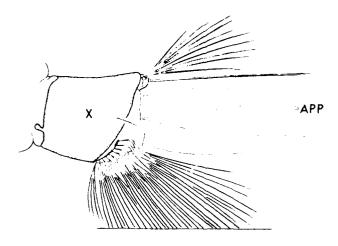


Fig. 601 — Lateral view of abdominal segment X - Ac. scapularis

22(21). Seta 13-III long, single (Fig. 602); thoracic integument densely aculeate (Fig. 603)scapularis (Plate 12)

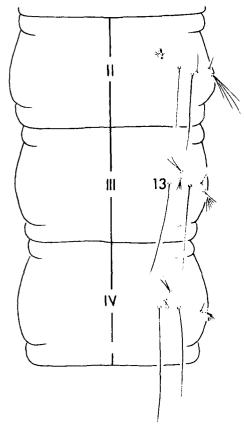


Fig. 602 — Ventral view of abdominal sterna II-IV - Ae. scapularis

Fig. 604 — Ventral view of abdominal sterna II-IV - Ae. tortilis

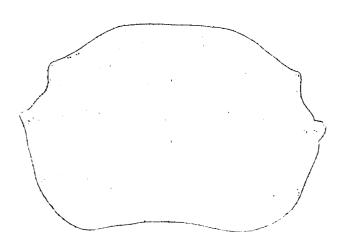


Fig. 603 — Dorsal view of thorax Ac. scapulars

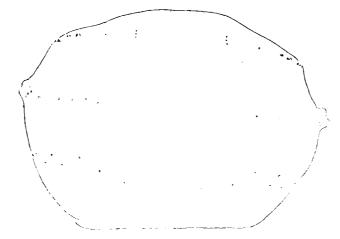
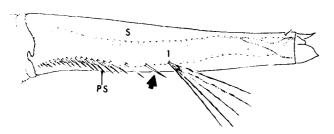


Fig. 605 — Dorsal view of thorax - Ac. tortilis

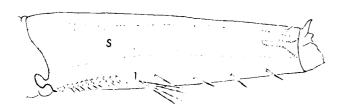


Smilletting

Fig. 606 - Lateral view of siphon - Ae. excrucians

Fig. 607 — Lateral view of siphon - Ae. melanimon

24(23). Seta 1-S attached within pecten (Fig. 608)	
Seta 1-S attached distal to pecten (Fig. 609)	27



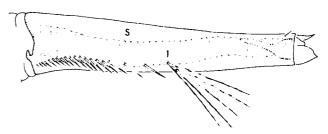


Fig. 608 - Lateral view of siphon - Ae. cataphylla

Fig. 609 - Lateral view of siphon - Ae. excrucians

al spine and short, lateral spinules (Fig. 610); seta 1-X attached	***
(Plate 10)	to saddle (Fig. 011)
bequal spinules (Fig. 612); seta 1-X attached ventral to saddle	



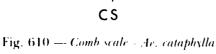




Fig. 612 — Comb scale - Ac. atropalpus

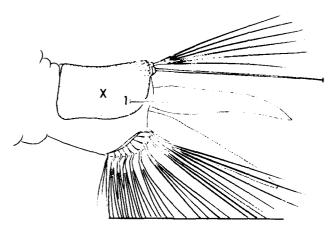


Fig. 611 — Lateral view of abdominal segment X - Ae. cataphylla

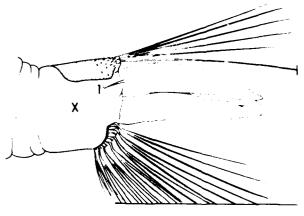


Fig. 613 — Lateral view of abdominal segment X Ae. atropalpus

26(25). Seta 1-M long, reaching near to level of seta 1-P (Fig. 614); with 34 or more comb scales (Plete 13)

Seta 1-M short, only reaching near to level of seta 0-P (Fig. 616); with fewer than 34 comb scales (Fig. 617) . . epactius (Plate 13)

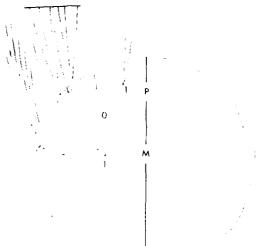


Fig. 614 — Dorsal view of thorax - Ae, atropalpus

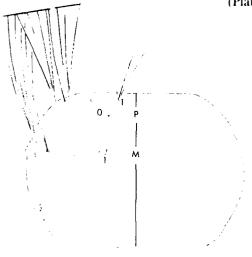
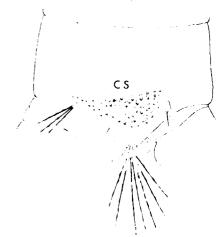


Fig. 616 - Dorsal view of thorax - Ae. epactus



atropalpus

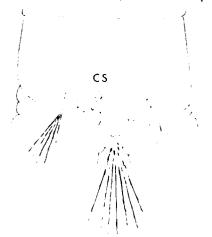


Fig. 615 — Lateral view of abdomival segment VIII · Ac. Fig. 617 — Lateral view of abdominal segment VIII · Ac. epactnes



Fig. 618 — Dorsal view of head and antennae - Ae. diantaeus

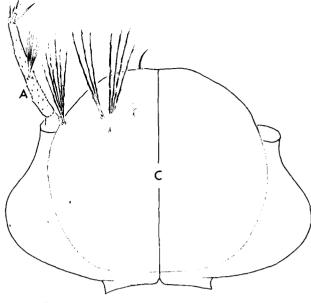


Fig. 619 — Dorsal view of head and antennae - Ae. vexans



Fig. 620 — Dorsal view of head and antennae Ac. diantaeus



Fig. 622 — Dorsal view of head and antennae - Ae. aurifer

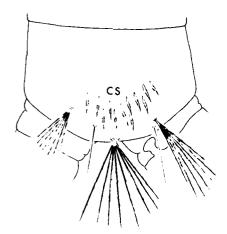


Fig. 621 — Lateral view of abdominal segment VIII - Ae. diantaeus

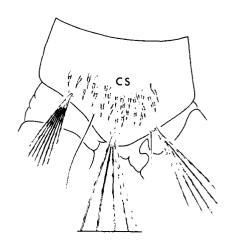


Fig. 623 — Lateral view of abdominal segment VIII - Ae. aurifer

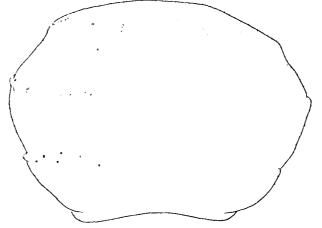


Fig. 624 - Dorsal view of thorax - Ae. s. spencerii

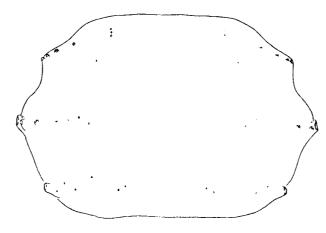


Fig. 625 — Dorsal view of thorax - Ae. campestris

30(29). Comb scales 13 or fewer (Fig. 626); median spine of comb scale broad at base (Fig. 627) s. spencerii (Plate 21)

Comb scales 14 or more (Fig. 628); median spine of comb scale narrow at base (Fig. 629) s. idahoensis (Plate 21)

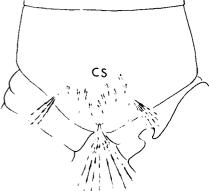


Fig. 626 — Lateral view of abdominal segment VIII - Ac. s. spencerii



Fig. 628—Lateral view of abdominal segment VIII - Ac. s. idahoensis



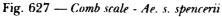




Fig. 629 — Comb scale - Ae. s. idahoensis

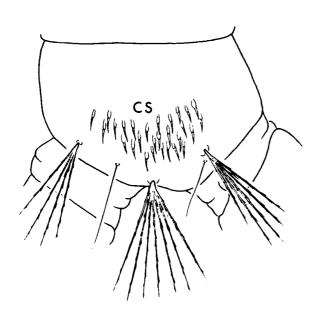


Fig. 630 — Lateral view of abdominal segment VIII - Ae. excrucians

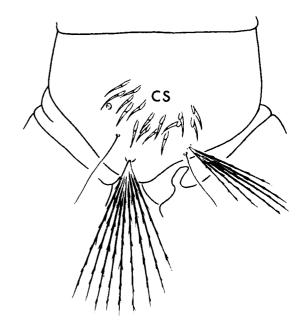


Fig. 631 — Lateral view of abdominal segment VIII - Ae. intrudens

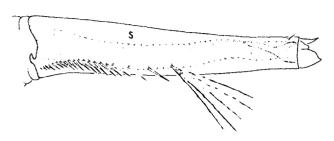


Fig. 632 - Lateral view of siphon - Ae. excrucians

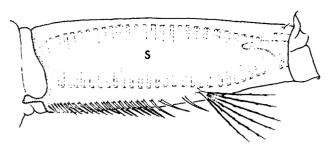


Fig. 634 - Lateral view of siphon - Ae. campestris

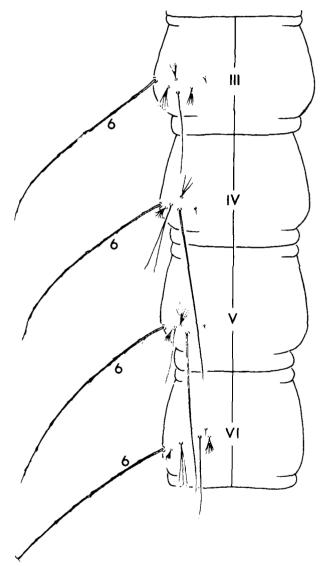


Fig. 633 — Dorsal view of abdominal segments III-VI - Ae. excrucians

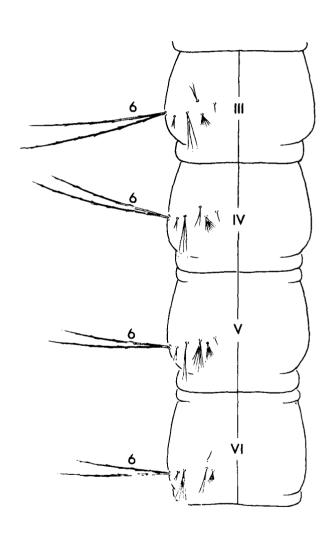


Fig. 635 — Dorsal view of abdominal segments III-VI - Ae. campestris

33(32) Pecten reaching distal to middle of siphon (Fig. 636); seta 1-M longer than antenna (Fig. 637) (in part) campestris (Plate 12)

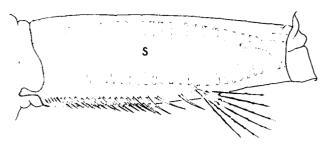


Fig. 636 - Lateral view of siphon - Ae. campestris

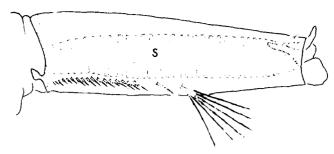
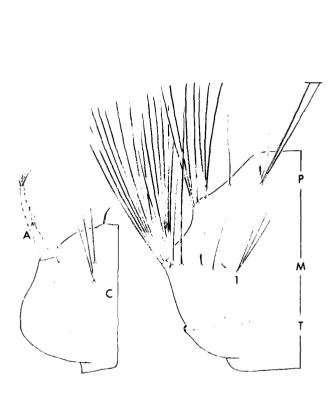


Fig. 638 - Lateral view of siphon - Ae. flavescens



A

Fig. 637 — Dorsal view of thorax and head - Ae, campestris

Fig. 639 - Dorsal view of thorax and head. Ac, flaveseens

34(33). Siphon index 3.5-4.0 (Fig. 640); body integument glabrous (Fig. 641) (in part) flavorens (Plate 12)

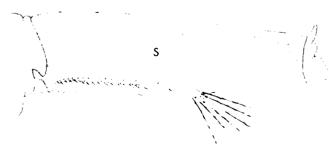
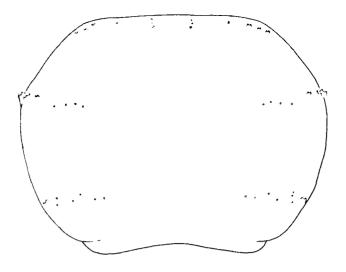


Fig. 640 - Lateral view of siphon Ar. flavesiens



Fig. 642 — Lateral view of siphon—Ac, aloponotum



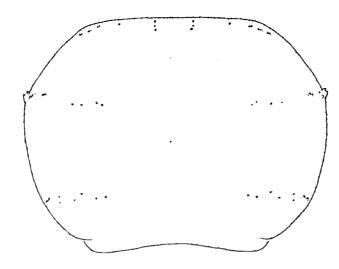
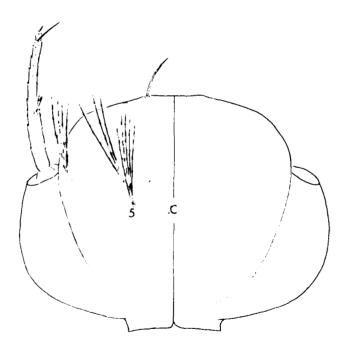


Fig. 641 - Dorsal view of thorax - Ae. flavescens

Fig. 643 - Dorsal view of thorax - Ae. aloponotum

 35(31). Seta 5-C with 3 or more branches (Fig. 644)
 36

 Seta 5-C single or double, rarely triple on both sides (Fig. 645)
 38



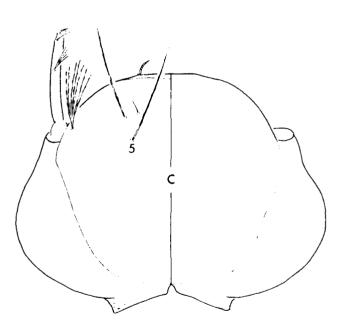


Fig. 644 — Dorsal view of head - Ae, intrudens

Fig. 645 - Dorsal view of head - Ac. niphadopsis

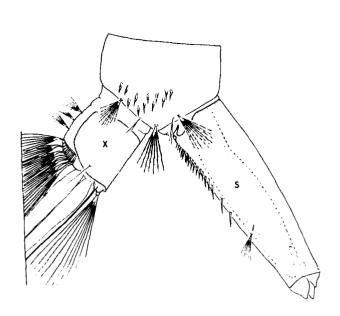
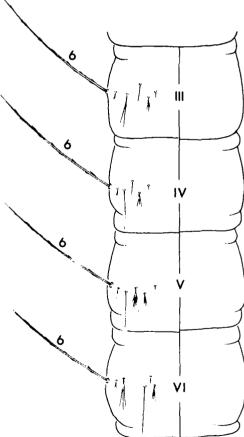


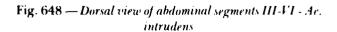
Fig. 646—Lateral view of abdominal segments VIII-X - Ae. vexans

Fig. 647 — Lateral view of abdominal segments VIII-X - Ae. euedes

37(36). Seta 6 usually single on III-VI (Fig. 648); seta 1-S with 4 or more branches (Fig. 649)intrudens (Plate 9)

Seta 6 usually double on III-VI (Fig. 650); seta 1-S double or triple (Fig. 651) (in part) euedes (Plate 15)





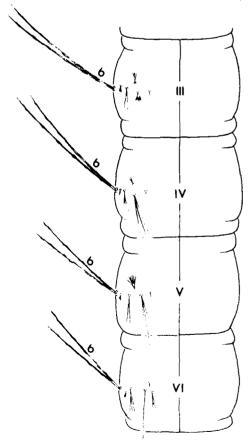


Fig. 650 — Dorsal view of abdominal segments III-V1 - Ae. eucles

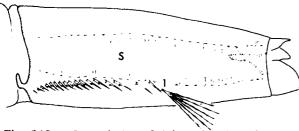


Fig. 649 — Lateral view of siphon - Ae. intrudens

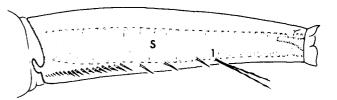


Fig. 651 — Lateral view of siphon - Ae. euedes

(Plate 17)



Fig. 652 - Dorsal view of head - Ae. decticus

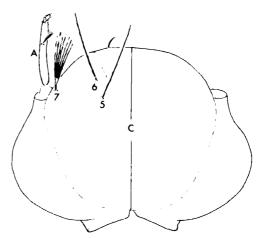


Fig. 653 - Dorsal view of head - Ae. niphadopsis



Fig. 654 — Comb scale - Ae, niphadopsis



Fig. 657 — Comb scale - Ae. riparius

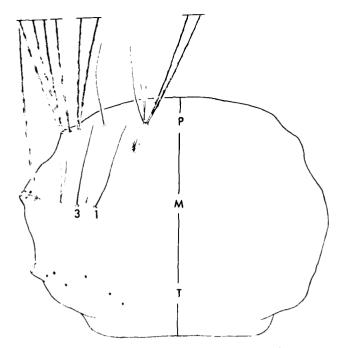


Fig. 655 - Dorsal view of thorax - Ae. niphadopsis

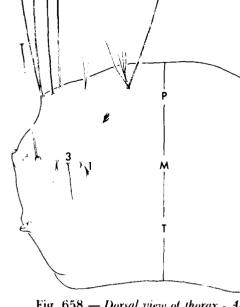


Fig. 658 - Dorsal view of thorax - Ae. riparius



Fig. 656 - Lateral view of siphon - Ae. niphadopsis

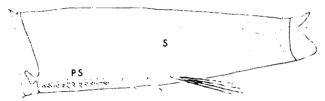


Fig. 659 - Lateral view of siphon - Ae. riparius

40(39). Comb with 12 or more scales; pecten on siphon with 18 or more spines (Fig. 660) (in part) euedes (Plate 15)



Fig. 660 — Lateral view of abdominal segment VIII - Acenedes

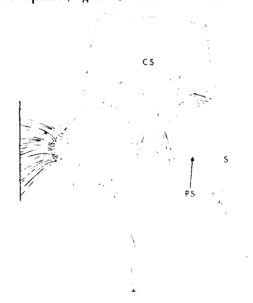
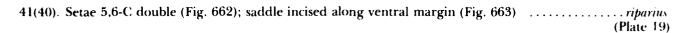


Fig. 661 — Lateral view of abdominal segment VIII - Acventrocattis



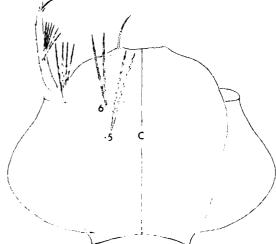


Fig. 662 — Dorsal view of head - Ae. riparius

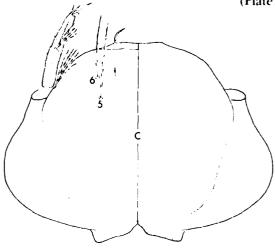


Fig. 664 — Dorsal view of head - Ae. ventrovittis

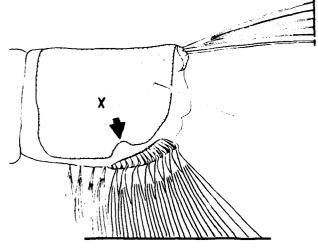


Fig. 663 — Lateral view of abdominal segment X - Ac. ribarius

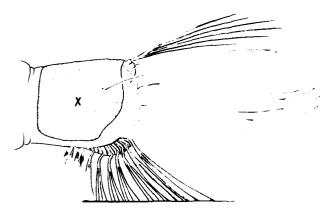


Fig. 665 — Lateral view of abdominal segment $X \cdot Ae$.



Fig. 666 — Dorsal view of antenna - Ae, triseriatus



Fig. 667 — Dorsal view of antenna - Ae, fitchir



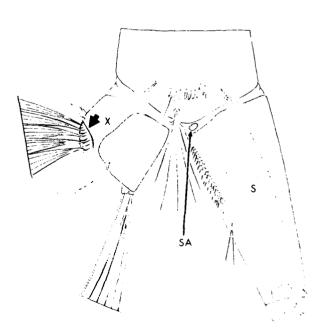


Fig. 668 — Comb scale - Ae, purpureipes

Fig. 669 — Comb scale - Ae. triseriatus

44(43). Boss of ventral brush weakly sclerotized; siphon without acus (Fig. 670)papago (Plate 24) Boss of ventral brush strongly sclerotized or brush arising from grid; siphon with acus





papago

Fig. 670 — Lateral view of abdominal segments VIII(X). Ac. Fig. 674 — Lateral view of abdominal segments VIII(X). Ac.

45(44). Integument of thorax and abdomen aculeate (Fig. 672); with 3-7 comb scales (Fig. 673) purpurupes

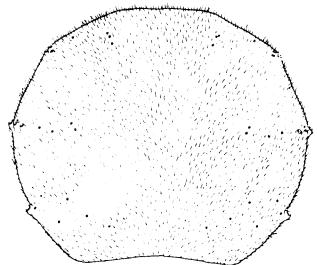


Fig. 672 - Dorsal view of thorax - Ae. purpureipes

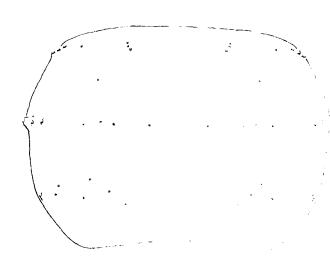


Fig. 674 — Dorsal view of thorax Ae. argspti

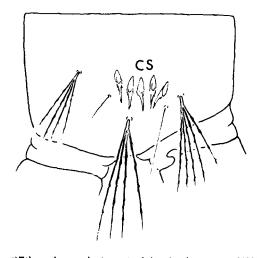


Fig. 673 — Lateral view of abdominal segment VIII - Acpurpureipes

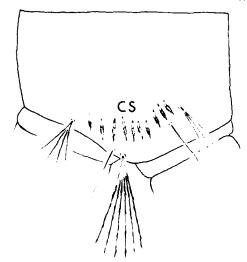


Fig. 675 — Lateral view of abdominal segment VIII - Ac. aegypti



Fig. 676 — Comb scale - Ae. acgypti

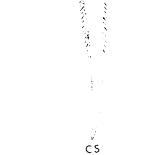


Fig. 678 - Comb scale Ac. muelleri

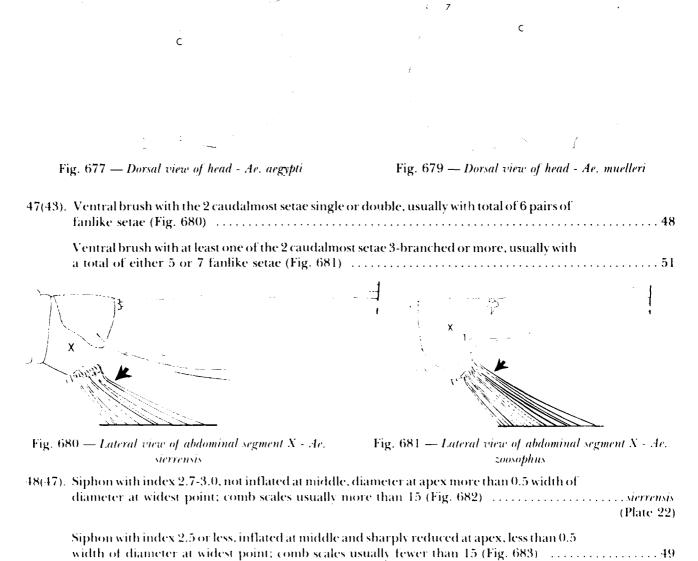
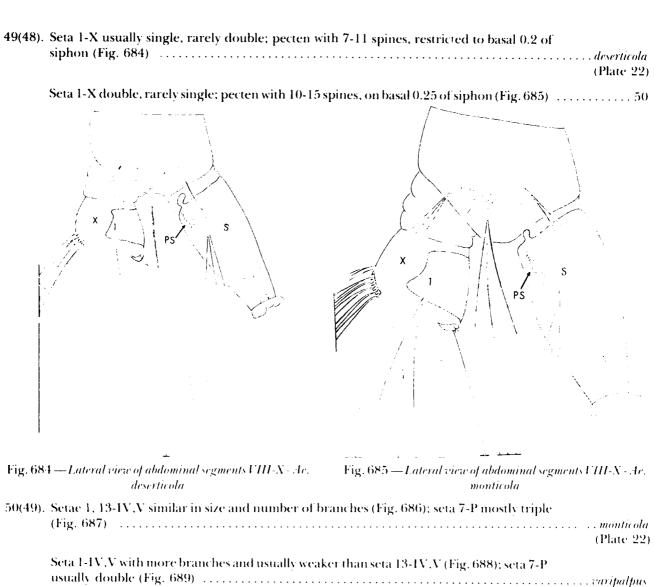


Fig. 682 — Lateral view of abdominal segments VIII X (Ac.) Sierrensis

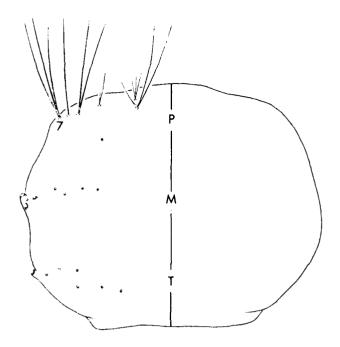
Fig. 683 — Lateral view of abdominal segments VIII-X (Ac. monticola





** (so coal segments Fig. 688 -- Dorsal ventral view of abdominal segments IV V ** Ac. varipalpus

(Plate 22)



P M

Fig. 687 — Dorsal view of thorax - Ae. monticola

Fig. 689 — Dorsal view of thorax - Ae. varipalpus

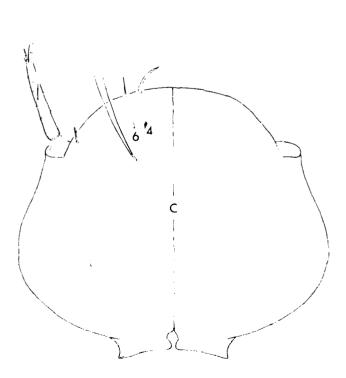


Fig. 690 - Dorsal view of head - Ae, burgeri

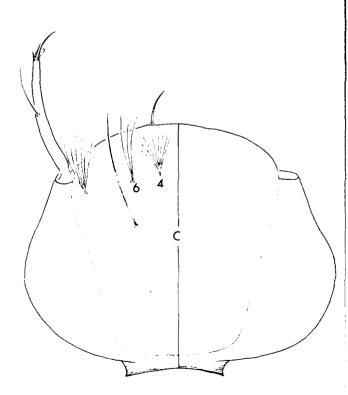
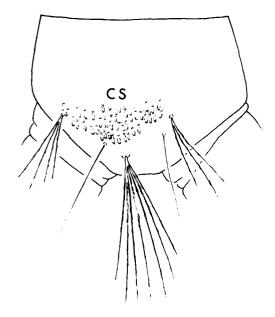


Fig. 692 — Dorsal view of head - Ac. triseriatus



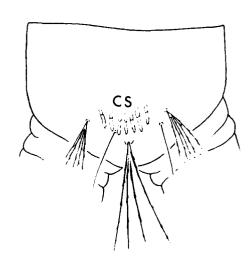


Fig. 691 — Lateral view of abdominal segment VIII - Ae. burgeri

Fig. 693 — Lateral view of abdominal segment VIII - Ae. triseriatus

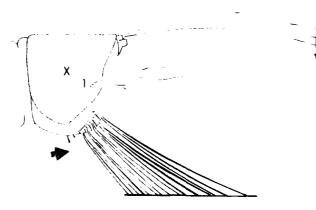


Fig. 694 — Lateral view of abdominal segment X - Ac. zoosophus

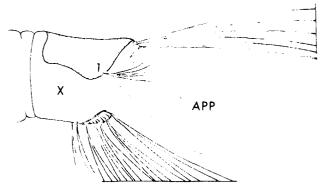


Fig. 695 — Lateral view of abdominal segment X - Ae.

Ventral brush with 5 pairs of fanlike setae (Fig. 698); acus detached and removed from base of siphon (Fig. 699); both pairs of anal papillae about same length, bulbous (Fig. 698) 54

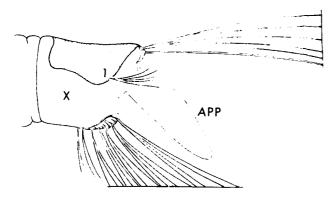


Fig. 696 — Lateral view of abdominal segment X – Ae. triseriatus

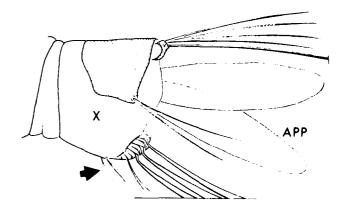


Fig. 698 — Lateral view of abdominal segment X - Ae. hendersoni

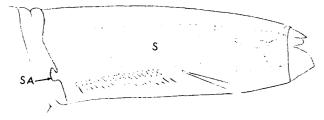


Fig. 697 — Lateral view of siphon - Ac. triscriatus

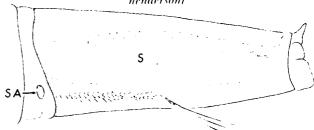


Fig. 699 — Lateral view of siphon - Ae, hendersoni

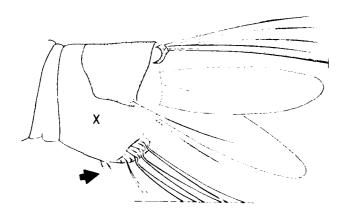


Fig. 700 — Lateral view of abdominal segment X—Ac. hendersoni

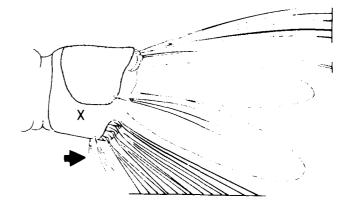


Fig. 701 — Lateral view of abdominal segment X · Ac.



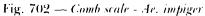




Fig. 703 — Comb scale - Ae. cantator

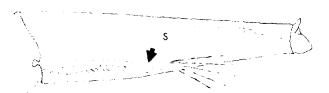


Fig. 704 - Lateral view of siphon - Ae, fitchii

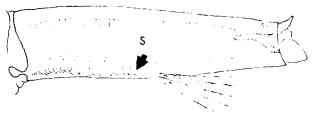


Fig. 705 - Lateral view of siphon - Ac. c. canadensis

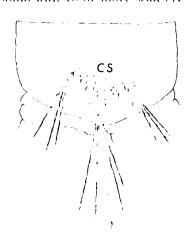


Fig. 706 — Lateral view of abdominal segment VIII. Acimboses

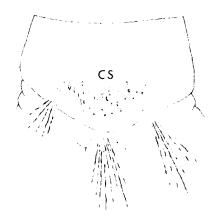


Fig. 707 — Lateral view of abdominal segment VIII—4e stimulars

58(57). Saddle extending near to midventral line; anal	•
,,	(Plate 25)
Saddle extending only about 0.5 to midventral lin	ne; anal papilla-saddle index 2.0 or more
(Fig. 709)	impiger (Plate 22)
7	(Flate 22)
X APP	
A The state of the	X
	APP
Fig. 708 — Lateral view of abdominal segment X - Ae. punctodes	Fig. 709 — Lateral view of abdominal segment X - Ae. impiger
•	60
	65
Seta 1-X longer than sattle (Fig. 717)	
The state of the s	
X	x 1-/-
The state of the s	
Fig. 710 — Lateral view of abdominal segment X - Ae. stimulans	Fig. 711 — Lateral view of abdominal segment X - Ae. aboriginis
sommans 50(59). Setae 5,6-C single, rarely double (Fig. 712)	
Seta 5-C with 2-4 branches, seta 6-C usually do	ouble (Fig. 713)
	the second second
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	<u> </u>
Fig. 712 - Dorsal view of head - Ac, melanimon	Fig. 713 — Dorsal view of head - Ae, sticticus

61(60).	Seta 1 attached distad to middle of siphon (Fig. 714); seta 1-M about equal to seta 2-M in		
	length (Fig. 715)	part)	melanimon
			(Plate 23)

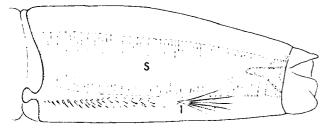


Fig. 714 — Lateral view of siphon - Ae, melanimon

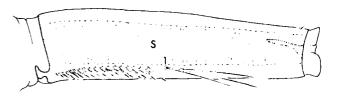


Fig. 716 — Lateral view of siphon - Ae. stimulans

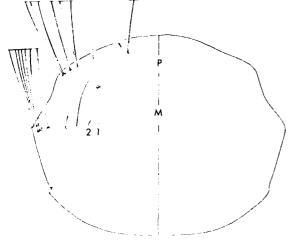


Fig. 715 — Dorsal view of thorax - Ae. melanimon

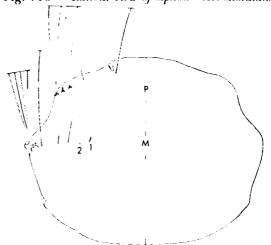


Fig. 717 — Dorsal view of thorax - Ae. stimulans

(Plate 11)



Fig. 718 — Comb scale - Ae, nevadensis



Fig. 720 — Comb scale - Ae. stimulans

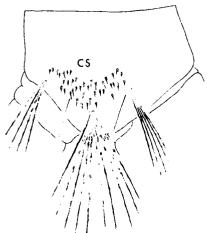


Fig. 719 — Lateral view of abdominal segment VIII - Ae.
nevadensis

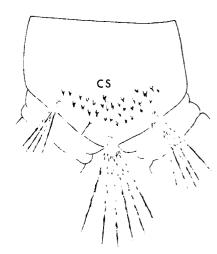


Fig. 721 - Lateral segment of abdominal segment VIII - Ae. stimulans

Seta 1-M shorter than seta 3-M and seta 5-C (Fig. 724, 725)

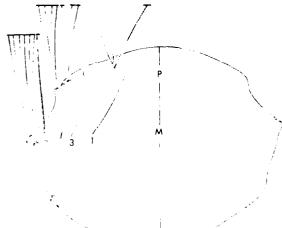


Fig. 722 — Dorsal view of thorax - Ae. mercurator

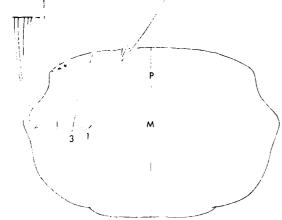


Fig. 724 — Dorsal view of thorax - Ae. sticticus



Fig. 723 — Dorsal view of head - Ae, mercurator

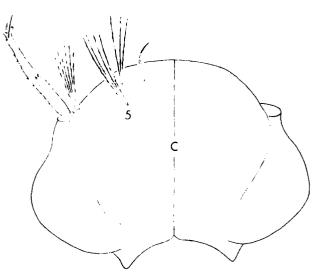


Fig. 725 — Dorsal view of head - Ac. stictious

64(63). Siphon index 3.2-4.0 (Fig. 726); comb scale with stout, subapical spinules (Fig. 727) (in part) flavescens (Plate 12)

Siphon index 2.5-3.0 (Fig. 728); comb scale with subapical spinules weak (Fig. 729)sticticus (Plate 25)

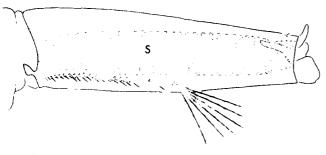


Fig. 726 - Lateral view of siphon - Ae. flavescens

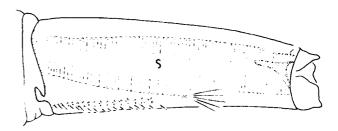


Fig. 728 — Lateral view of siphon - Ae. sticticus



Fig. 727 — Comb scale - Ac. flavescens

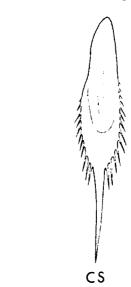


Fig. 729 — Comb scale - Ac. sticticus

65(59). Posterior border of saddle aciculate (Fig. 730); seta 1-M with 3-6 branches (Fig. 731) (Plate 26)

Posterior border of saddle without aciculae (Fig. 732); seta 1-M single (Fig. 733) aboriginis (Plate 9)



Fig. 730 — Lateral view of abdominal segment X - Ac. schizopinax

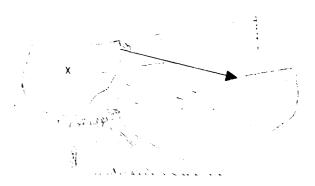
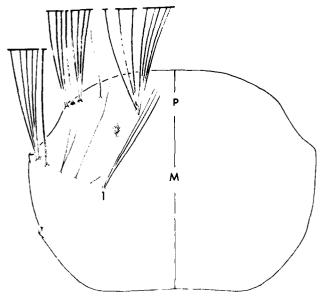


Fig. 732 — Lateral view of abdominal segment X - Ae, aboriginis



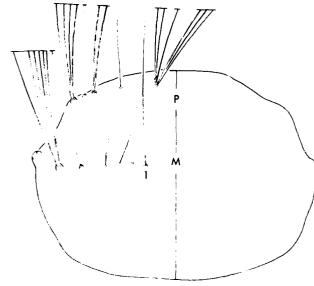
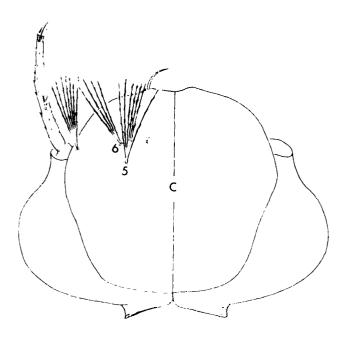


Fig. 731 — Dorsal view of thorax - Ae. schizopinax

Fig. 733 — Dorsal view of thorax - Ae. aboriginis



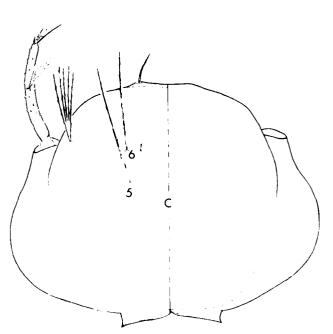


Fig. 734 — Dorsal view of head - Ae. pullatus

Fig. 735 — Dorsal view of head - Ae. dorsalis



Fig. 736 — Dorsal view of thorax - Ae. pullatus

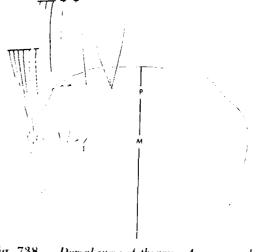


Fig. 738 - Dorsal view of thorax - Ae. c. canadensis

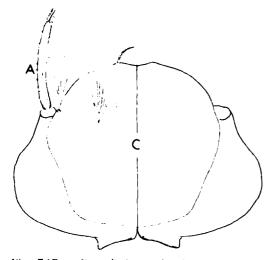


Fig. 737 — Dorsal view of head - Ac. pullatus

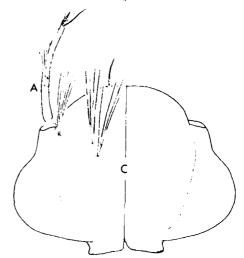


Fig. 739 — Dorsal view of head - Ae. c. canadensis

Seta 3-P double or triple (Fig. 742); with 60 or fewer comb scales (Fig. 743)

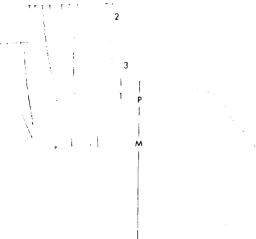
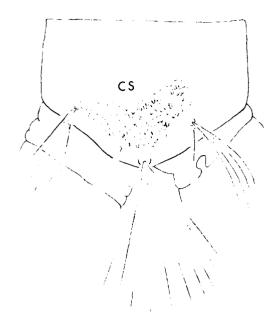


Fig. 740 - Dorsal view of thorax - Ar. promps



Fig. 742 — Dorsal view of thorax - Ae, pullatus



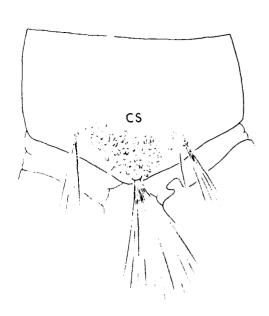


Fig. 741 — Lateral view of abdominal segment VIII - Ae. pionips

Fig. 743 — Lateral view of abdominal segment VIII - Ae. pullatus

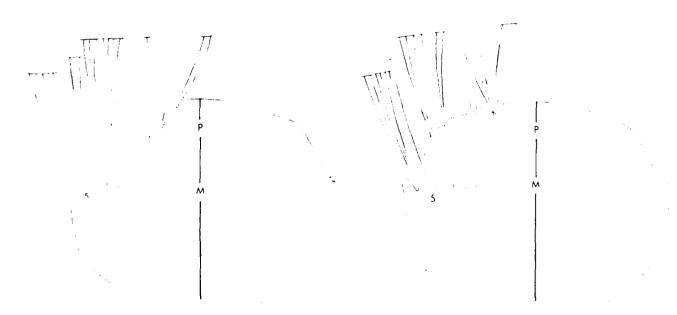
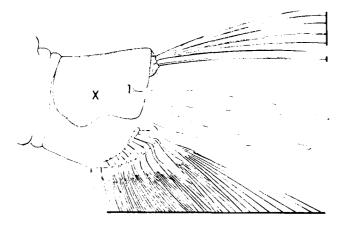


Fig. 744 - Dorsal view of thorax - Ac. pullatus

Fig. 745 - Dorsal view of thorax - Ae, cantator



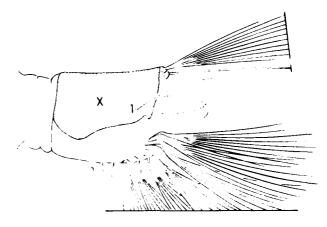
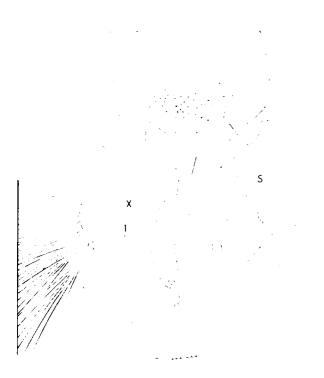


Fig. 745 — Lateral view of abdominal segment X - Ae. pullatus

Fig. 747 — Lateral view of abdominal segment X - Ae. cantator



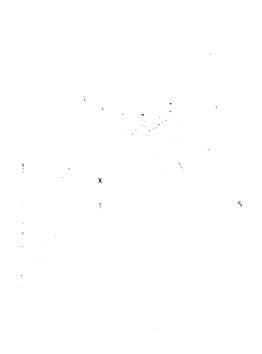


Fig. 748—Lateral www of abdominal segments VIII-X (Ac.)

Fig. 749 -- Lateral view of abdominal segments VIII-X - Ac. c. canadensis

Comb scale fringed with subequal spinules (Fig. 752); seta 6-1,11 double (Fig. 753) e. canadensis c. mathesau (Plates 14, 18)



Fig. 750 — Comb scale - Ae. thibaulti



Fig. 752 — Comb scale - Ae. c. canadensis



thibaulti

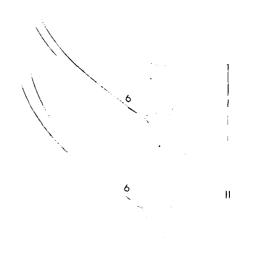


Fig. 751 — Dorsal view of abdominal segments I-II - Ae. Fig. 753 — Dorsal view of abdominal segments - I-II - Ae. c. canadensis

(Plate 27)

Seta 1-X shorter than saddle (Fig. 755)



Fig. 754 — Lateral view of abdominal segment X - Ae. squamiger

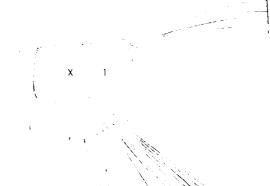
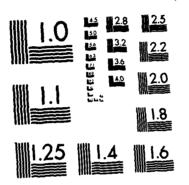


Fig. 755 — Lateral vii v of abdominal segment X - Ae communis

IDENTIFICATION AND GEOGRAPHICAL DISTRIBUTION OF THE MOSAUITOES OF NORTH AMERICA NORTH OF MEXICO(U) HALTER REED ARMY INST OF RESEARCH HASHINGTON DC F DARSIE ET AL 01 AUG 81 F/G 6/3 AD-A125 975 3/4 UNCLÁSSIFIED NL



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73(72). Seta 1-M about equal to length of antenna, or le	onger (Figs. 756, 757)74
Seta 1-M starte chan antenna (Figs. 758, 759)	76
Fig. 756 — Dorsal view of thorax - Ae. dorsalis	Fig. 758 — Dorsal view of thorax - Ae. increpitus
	F

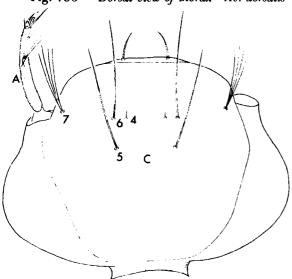


Fig. 757 - Dorsal view of head - Ae. dorsalis

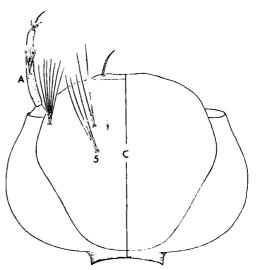


Fig. 759 — Dorsal view of head - Ae. increpitus

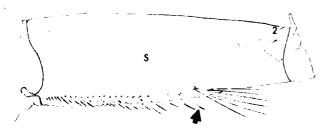


Fig. 760 - Lateral view of siphon - Ae. campestris

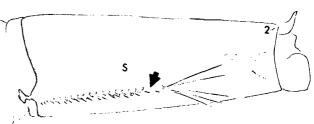


Fig. 761 - Lateral view of siphon - Ae. dorsalis

75(74).	Seta 1-X about 0.5 length of saddle (Fig. 762); the 4 setae 5,6-C usually single or the total		
	single and branches of branched setae rarely more than 7 (Fig. 763)		
	(Plate 18)		
	Seta 1-X almost equal to length of saddle (Fig. 764); the 4 setae 5,6-C usually branched, the		
	total single and branches of branched setae usually 10, not fewer than 8 (Fig. 765) grossbecki		

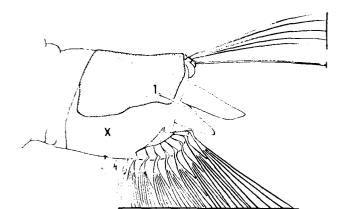
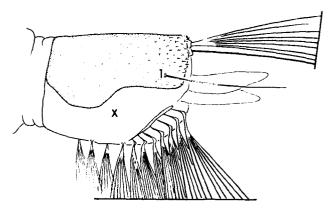


Fig. 762 — Lateral view of abdominal segment X - Ae. dorsalis



(Plate 22)

Fig. 764 — Lateral view of abdominal segment X - Ae. grossbecki

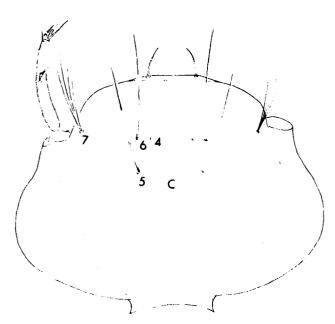


Fig. 763 - Dorsal view of head - Ae. dorsalis

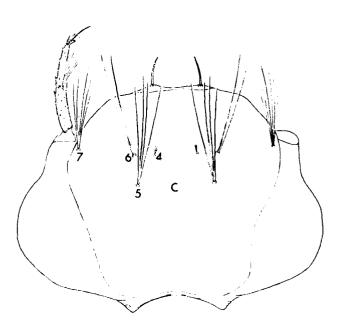


Fig. 765 — Dorsal view of head - Ae. grossbecki

76(73). Comb scales 36 or more, with median spine no stouter than subapical spinules (Figs. 766,	
767)	communis* churchillensis (Plates 17, 20)
Comb scales fewer than 35, with median spine stouter than subapical spinules on at least some scales (Figs. 768, 769)	77

^{*}For provisional separation of communis and churchillensis, see Ellis and Brust (167).

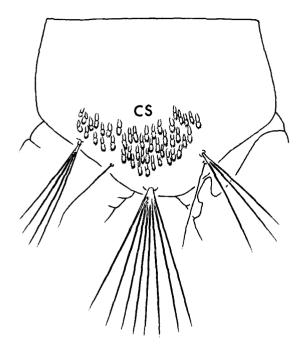


Fig. 766 — Lateral view of abdominal segment VIII - Ae. communis

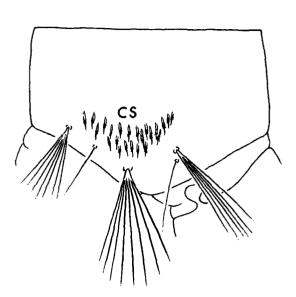


Fig. 768 — Lateral view of abdominal segment VIII - Ae. melanimon



Fig. 767 — Comb scale - Ae. communis



Fig. 769 — Comb scale - Ae, melanimon

77(76). F	Pecten extending distal to middle of siphon (Fig. 770); seta 1-IV,V short, multibranched	
(1	(Fig. 771)(in par	t) <i>melanimon</i> (Plate 23)
	Pecten confined to basal 0.5 of siphon (Fig. 772); seta 1-IV,V long, single to triple (Fig. 773)	78

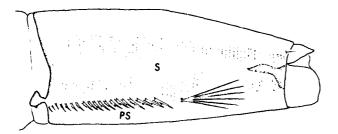


Fig. 770 — Lateral view of siphon - Ae. melanimon

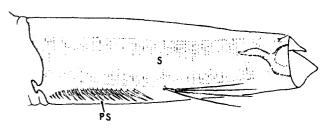


Fig. 772 — Lateral view of siphon - Ae. increpitus

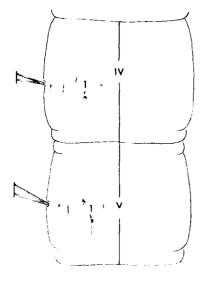


Fig. 771 — Dorsal view of abdominal segments IV-V - Ae. melanimon

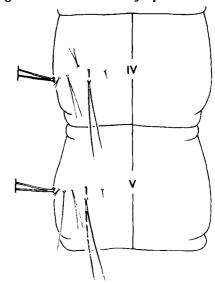


Fig. 773 — Dorsal view of abdominal segments IV-V - Ae. increpitus

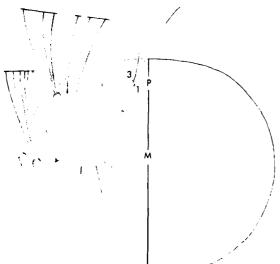


Fig. 774 — Dorsal view of thorax - Ae. implicatus

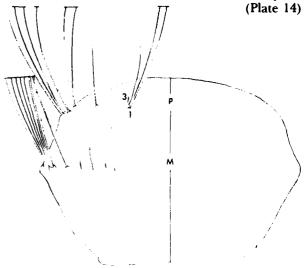
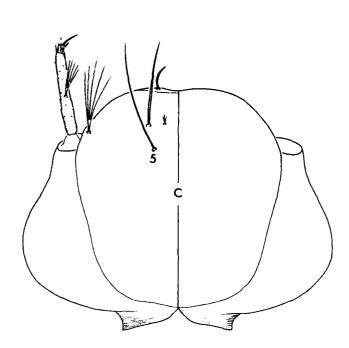
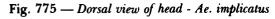


Fig. 777 — Dorsal view of thorax - Ae. increpitus





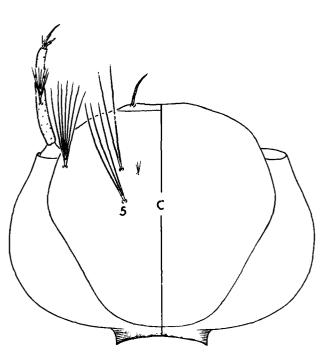


Fig. 778 — Dorsal view of head - Ae. increpitus

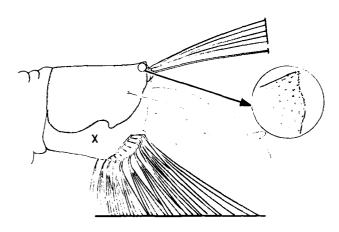


Fig. 776 — Lateral view of abdominal segment X - Ae. implicatus

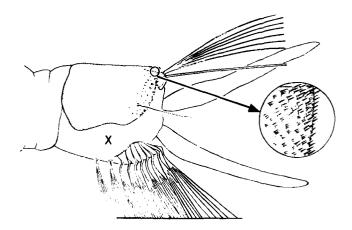


Fig. 779 — Lateral view of abdominal segment X - Ae. increpitus

KEY TO FOURTH STAGE LARVAE OF THE GENUS ANOPHELES

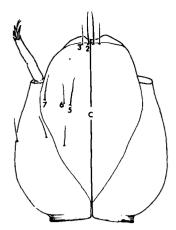


Fig. 780 — Dorsal view of head - An. judithae

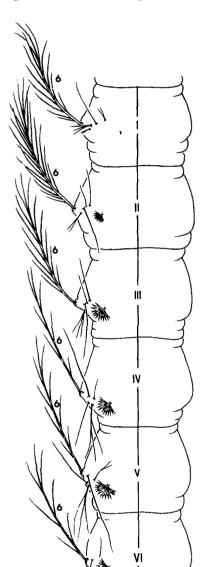


Fig. 781 — Dorsal view of abdominal segments I-VI - An. judithae

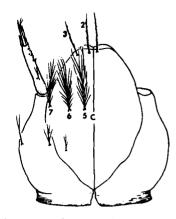


Fig. 782 — Dorsal view of head - An. albimanus

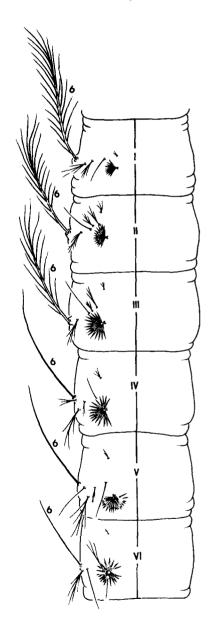


Fig. 783 — Dorsal view of abdominal segments I-VI - An. albimanus

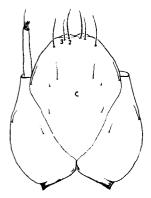


Fig. 784 - Dorsal view of head - An. barberi

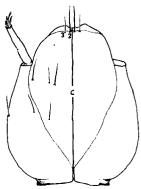


Fig. 786 - Dorsal view of head - An. judithae

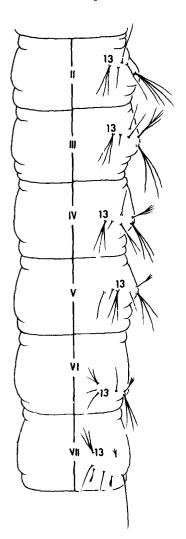


Fig. 785 — Ventral view of abdominal segments II-VII - An. barberi

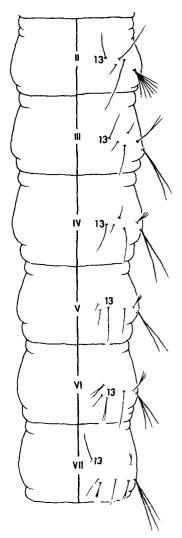
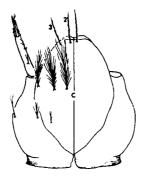
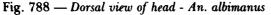


Fig. 787 — Ventral view of abdominal segments II-VII - An. judithae

 3(1).
 Seta 3-C unbranched (Fig. 788)
 4

 Seta 3-C with 5 or more branches (Fig. 789)
 6





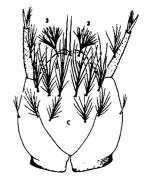


Fig. 789 — Dorsal view of head - An. quadrimaculatus

Seta 1 palmate on III-VII, leaflets with serrate margins (Fig. 792); setae 2,3-C smooth (Fig. 793)

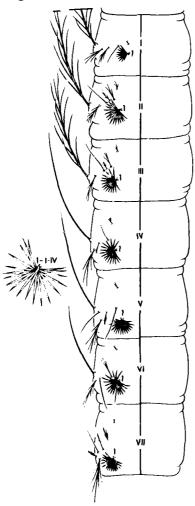


Fig. 790 — Dorsal view of abdominal segments I-VII - An. albimanus

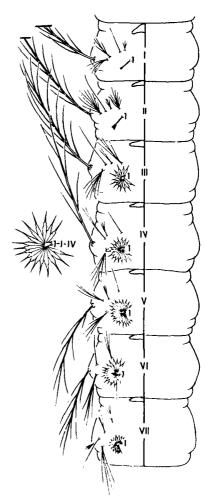


Fig. 792 — Dorsal view of abdominal segments I-VII - An. pseudopunctipennis

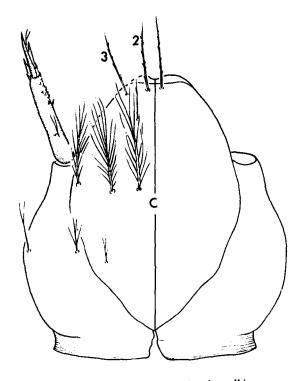


Fig. 791 - Dorsal view of head - An. albimanus

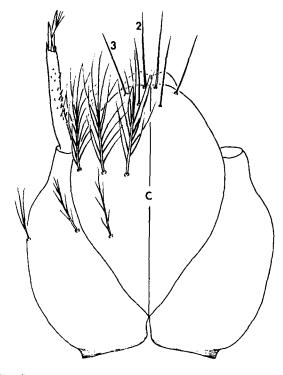


Fig. 793 — Dorsal view of head - An. pseudopunctipennis

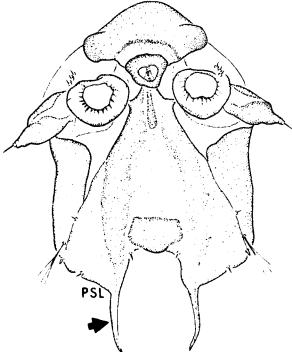


Fig. 794 — Spiracular apparatus - An. pseudopunctipennis

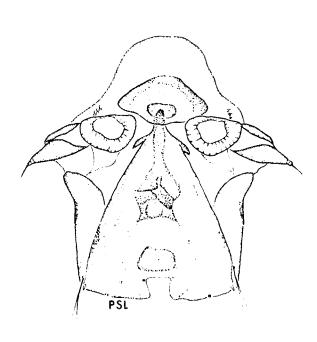
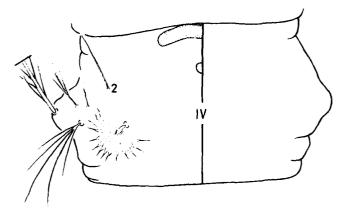


Fig. 796 — Spiracular apparatus - An. franciscanus



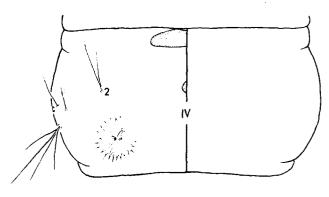


Fig. 795 — Dorsal view of abdominal segment IV - An. pseudopunctipennis

Fig. 797 — Dorsal view of abdominal segment IV - An. franciscanus

Seta 3-C dentritic, densely branched (Fig. 799)

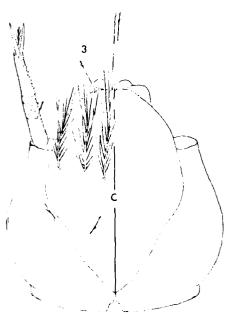


Fig. 798 — Dorsal view of head - An. atropos

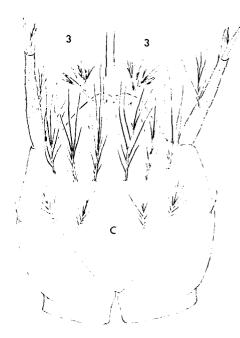
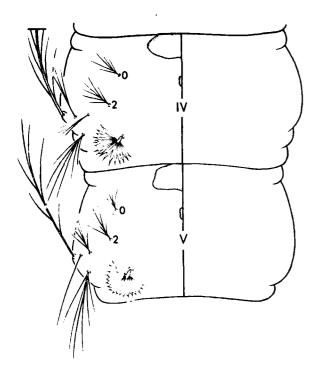


Fig. 799 - Dorsal view of head - An, quadrimaculatus

(Plate 31)



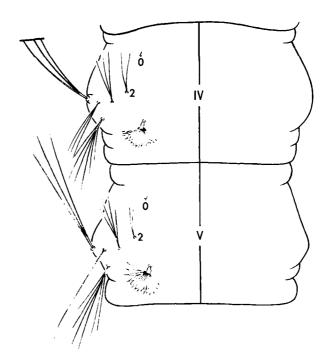


Fig. 800 — Dorsal view of abdominal segments IV-V - An. crucians

Fig. 801 — Dorsal view of abdominal segments IV-V - An. punctipennis

Seta 2-C simple, sparsely aciculate toward apex (Fig. 802); seta 1-P with 3-5 strong 8(7) branches from near base (Fig. 803)walkeri (Plate 28) Seta 2-C simple or forked in outer 0.5, without aciculae (Fig. 804); seta 1-P weak, single or

branches in outer 0.5 only (Fig. 805)9

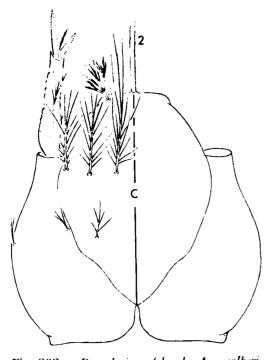


Fig. 802 - Dorsal view of head - An. walkeri

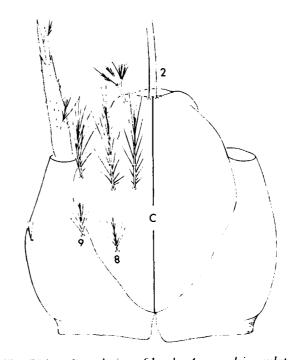
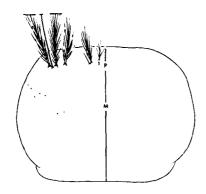


Fig. 804 - Dorsal view of head - An. quadrimaculatus



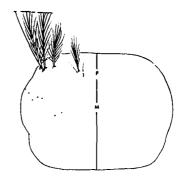
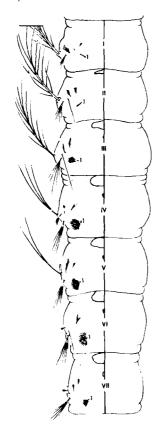
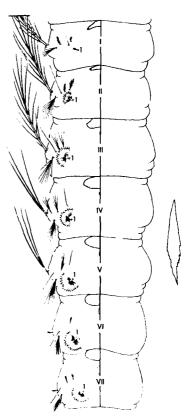


Fig. 803 — Dorsal view of thorax - An. walkeri

Fig. 805 — Dorsal view of thorax - An. quadrimaculatus

Seta 1-IV-VI fully palmate, 1-III and VII not more than 0.7 as large, leaflets usually with 9(8). marginal serrations fine (Fig. 806)10 Seta 1-III-VII fully palmate, apical 0.5 of leaflets with coarse marginal serrations (Fig.





bradleyi

Fig. 806 — Dorsal view of abdominal segments I-VII - An. Fig. 807 — Dorsal view of abdominal segments I-VII - An. quadrimaculatus

10(9). Seta 1-III better developed palmate seta than 1-I (Fig. 808); seta 5-II usually with fewer than 9 branches (Fig. 809)bradleyi (Plate 29) Seta 1-III not much better developed palmate seta than 1-I (Fig. 810); seta 5-II with 9 or more branches (Fig. 811) georgianus (Plate 30)

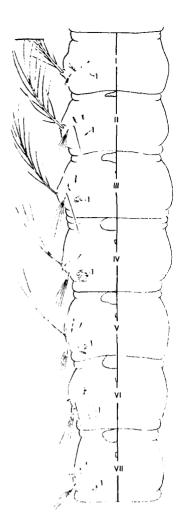


Fig. 808 — Dorsal view of abdominal segments I-VII - An. bradleyi

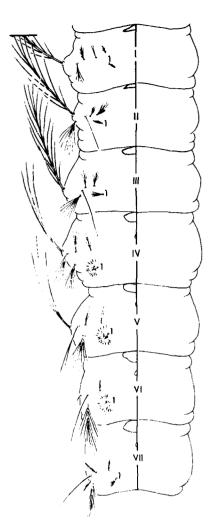


Fig. 810 — Dorsal view of abdominal segments I-VII - An. georgianus

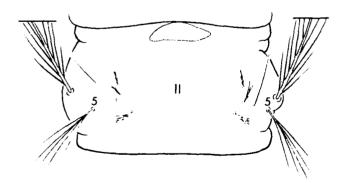


Fig. 809 — Dorsal view of abdominal segment II - An. bradleyi

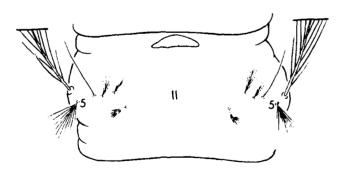
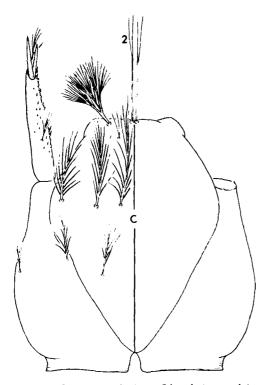
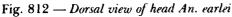


Fig. 811 — Dorsal view of abdominal segment II - An. georgianus





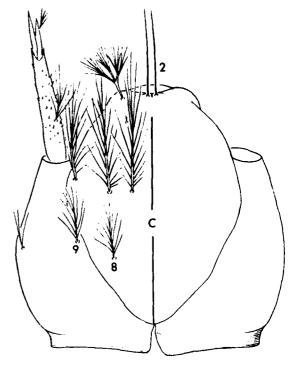


Fig. 813 - Dorsal view of head - An. quadrimaculatus

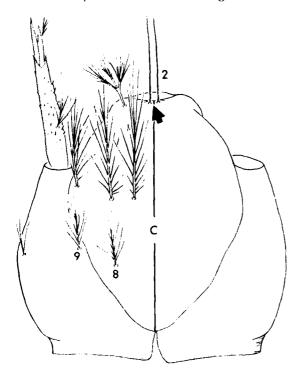


Fig. 814 - Dorsal view of head - An. quadrimaculatus

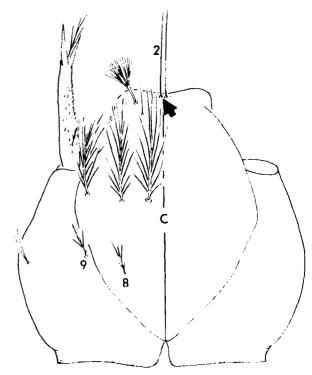
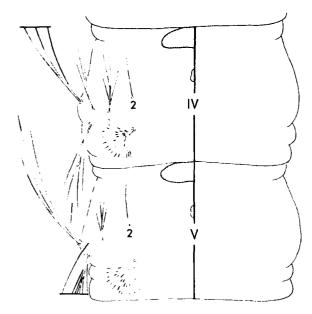


Fig. 815 - Dorsal view of head - An. punctipennis



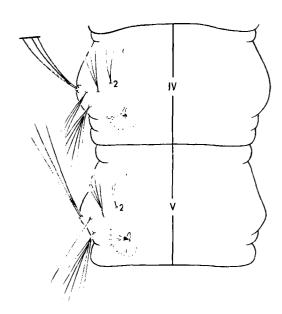
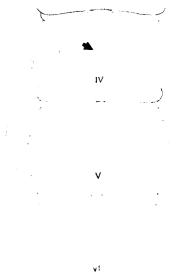


Fig. 816 — Dorsal view of abdominal segments IV-V - An. occidentalis

Fig. 817 — Dorsal view of abdominal segments IV-V- An. punctipennis

14(13). Segments IV-VI with 3 small, accessory, tergal plates (Fig. 818); seta 1-A attached at or distal to basal 0.33 of antenna; dorsal apotome with integument spotted (Fig. 819)freeborni (Plate 31)

Only 1 accessory, tergal plate on IV-VI (Fig. 820); seta 1-A attached within basal 0.33 of antenna; dorsal apotome with integument irregularly banded (Fig. 821)punctipennis (in part) perplexens



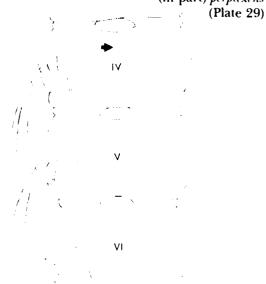


Fig. 818 — Dorsal view of abdominal segments IV-VI - An. freeborni

Fig. 820 — Dorsal view of abdominal segments IV-VI - An. punctipennis



Fig. 819 - Dorsal view of head - An. freeborni

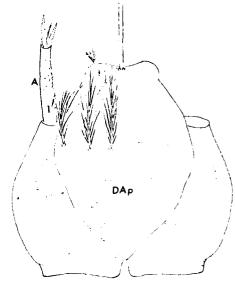


Fig. 821 - Dorsal view of head - An. punctipennis

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Fig. 822 — Dorsal view of head - Cx. pipiens



Fig. 823 — Dorsal view of head - Cx. territans

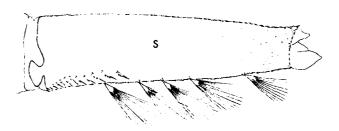


Fig. 824 — Lateral view of siphon - Cx. bahamensis

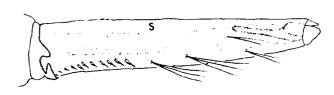


Fig. 826 — Lateral view of siphon - Cx. pipiens

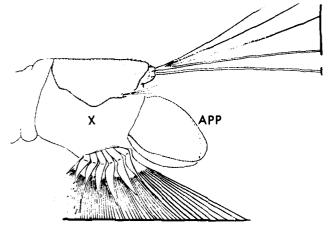


Fig. 825 — Lateral view of abdominal segment X - Cx. bahamensis

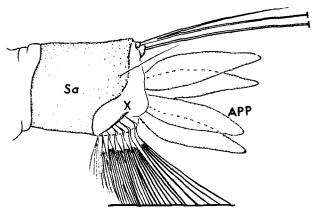


Fig. 827 — Lateral view of abdominal segment X - Cx. pipiens

3(2). Pecten reaching distal 0.75 of siphon, apical 4,5 spines large (Fig. 828)interrogator (Plate 32)



Fig. 828 — Lateral view of siphon - Cx. interrogator

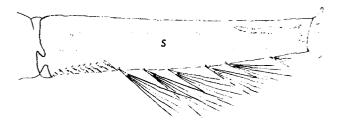


Fig. 829 - Lateral view of siphon - Cx. tarsalis



Fig. 830 — Lateral view of siphon - Cx. restuans

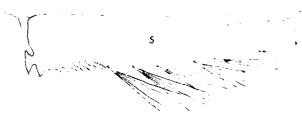
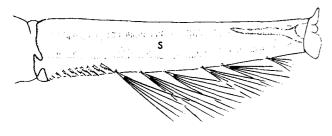


Fig. 831 — Lateral view of siphon - Cx. tarsalis

			a 1-A attached near middle (Fig. 832). of antenna, distal part more slender (Fig.	(Plate 38
Fig. 832 6(4). Sipl	2 — Dorsal view of a	entenna - Cx. restuans pines near apex (Fig. 834	Fig. 833 — Dorsal view of antenna	Cx. thriambus Coronator (Plate 35
Fig. 834	s — Lateral view of	iphon - Cx. coronator	Fig. 835 — Lateral view of siphon	- Cx. tarsalis
		traight line, usually with f setae not all in straight lin	5-9 pairs (Fig. 836)	8



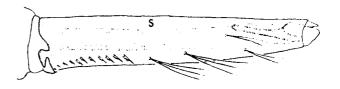


Fig. 836 — Lateral view of siphon - Cx. tarsalis

Fig. 837 — Lateral view of siphon - Cx. pipiens

8(7).	Siphon index 4.5-5.5, usually with 5 pairs of setae (Fig. 838)	tarsalis (Plate 34)
	Siphon index 8.0 or more, with 6-9 pairs of setae (Fig. 839)	chidesteri (Plate 34)

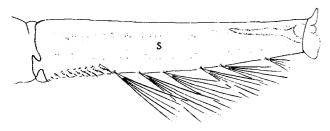




Fig. 838 - Lateral view of siphon - Cx. tarsalis

Fig. 839 — Lateral view of siphon - Cx. chidesteri

	Siphon with 3 pairs of setae (Fig. 840)	9(7).
)	Siphon with 4,5 pairs of setae (Fig. 841	





Fig. 840 - Lateral view of siphon - Cx. declarator

Fig. 841 - Lateral view of siphon - Cx. pipiens

10(9).	Siphon index 4.0-5.0 (Fig. 842)	 11
	Siphon index 6.0-8.0 (Fig. 843)	 12





Fig. 842 — Lateral view of siphon - Cx. pipiens

Fig. 843 — Lateral view of siphon - Cx. salinarius

11(10). Aciculae on dorsoposterior aspect of saddle m	nuch larger than those at dorsal middle (Fig.
844); seta 6-III,IV usually triple (Fig. 845)	
	(Plate 37)

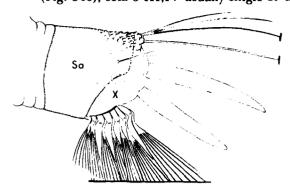


Fig. 844 — Lateral view of abdominal segment X - Cx. peus

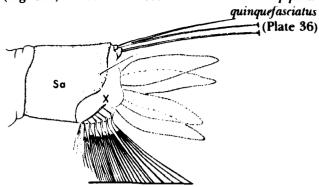


Fig. 846 — Lateral view of abdominal segment X - Cx. pipiens

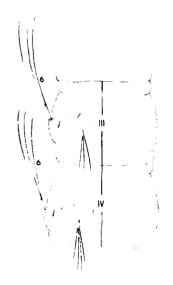


Fig. 845 — Dorsal view of abdominal segments III-IV - Cx. peus

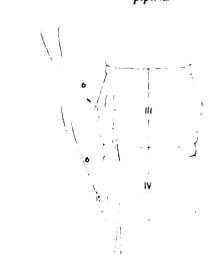


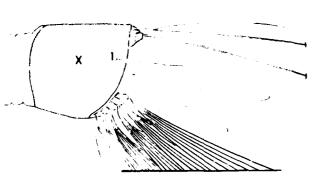
Fig. 847 — Dorsal view of abdominal segment III-IV - Cx. pipiens



Fig. 848 — Dorsal view of thorax - Cx. nigripalpus



Fig. 850 — Dorsal view of thorax - Cx. salinarius



X

Fig. 849 — Lateral view of abdominal segment X - Cx. nigripalpus

Fig. 851 — Lateral view of abdominal segment X - Cx. salinarius

13(12). Siphon usually with 5 pairs of setae, most often 2 pairs dorsally out of line (Fig. 852) ... erythrothorax (Plate 34)





Fig. 852 — Lateral view of siphon - Cx. erythrothorax

Fig. 853 — Lateral view of siphon - Cx. salinarius



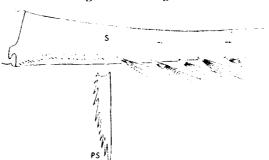


Fig. 854 — Lateral view of siphon - Cx. latisquama

Fig. 856 — Lateral view of siphon - Cx. peccator

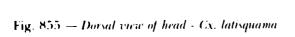
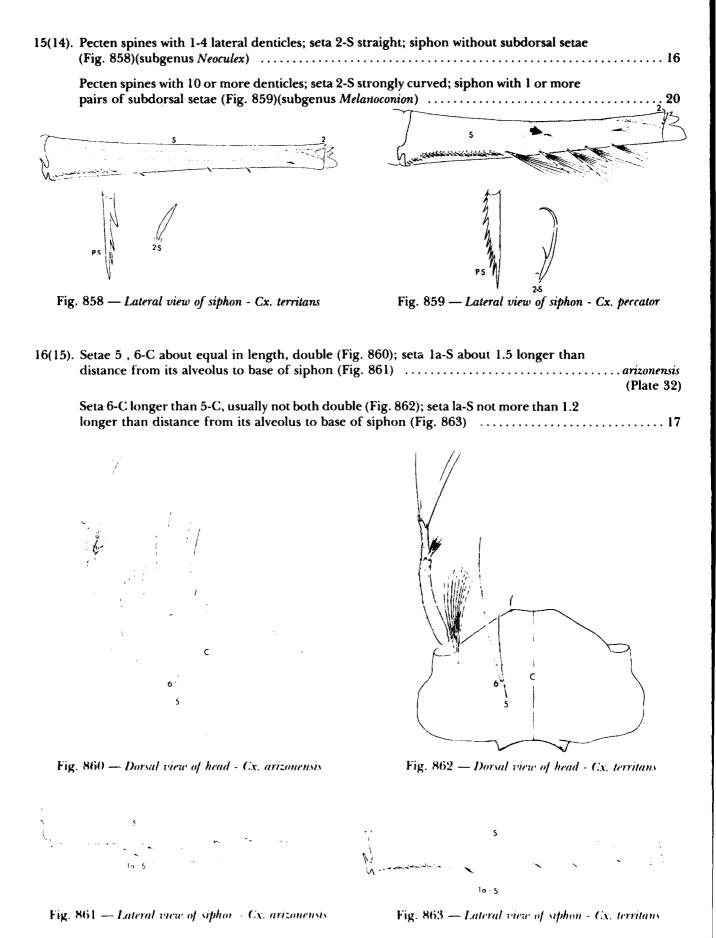
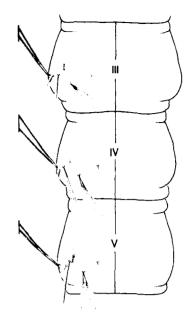


Fig. 857 — Dorsal view of head - Cx. peccator



Siphon less than 6.0 longer than basalmost seta,	index usually less than 7.0 (Fig. 865)
	7
/3	(3
With the territories with the second	hammer
Fig. 864 — Lateral view of siphon - Cx. apicalis	Fig. 865 — Lateral view of siphon - Cx. territans
17) Seta 5-C with 3 branches seta 6-C double (Fig. 8	366)reevesi
17). Seta 3-6 with 3 branches, seta 5-6 dodoic (Fig. 6	(Plate 33)
Seta 5-C single or double, seta 6-C usually single	e (Fig. 867)19
Fig. 866 — Dorsal view of head - Cx. reevesi 8). Seta 5-C single, occasionally double or triple (Fig. 8 pigmented (Fig. 869)	
pigmemen (Fig. 809)	
Seta 5-C double, rarely triple (Fig. 870); abdomin pigmented than IV (Fig. 871)	
	(Plate 35)
	· · · · · · · · · · · · · · · · · · ·



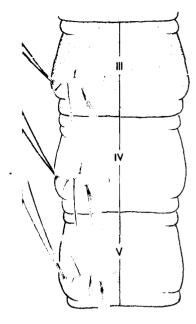


Fig. 869 — Dorsal view of abdominal segments III-V - Cx. territans

Fig. 871 — Dorsal view of abdominal segments III-V - Cx. boharti



Fig. 872 — Comb scale - Cx. pilosus



Fig. 873 — Comb scale - Cx. atratus

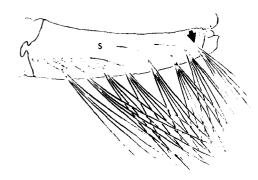


Fig. 874 — Lateral view of siphon - Cx. pilosus

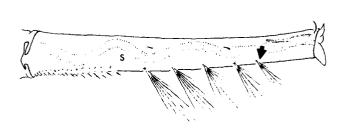


Fig. 875 — Lateral view of siphon - Cx. erraticus

 22(20). Siphon index more than 7.0 (Fig. 876)
 23

 Siphon index 7.0 or less (Fig. 877)
 24

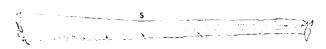


Fig. 876 — Lateral view of siphon - Cx. opisthopus



Fig. 877 - Lateral view of siphon - Cx. peccator

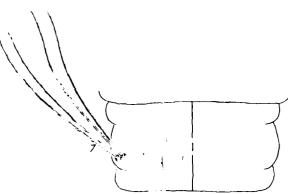


Fig. 878 — Ventral view of abdominal segment I - Cx. opisthopus

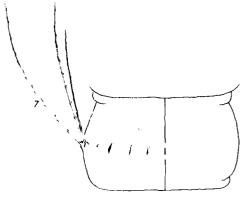


Fig. 880 — Ventral view of abdominal segment I - Cx. atratus



Fig. 879 — Lateral view of saddle - Cx. opisthopus



Fig. 881 - Lateral view of saddle - Cx. atratus

Comb scale long, with a narrow elongation in middle between base and apical fringed portion (Fig. 883) CS Fig. 882 — Comb scale - Cx. abominator Fig. 883 — Comb scale - Cx. iolambdis 5(24). Seta 5-C thin, much thinner and 0.5 or less length of seta 6-C, without aciculae (Fig. 884) Seta 5-G stout, about 0.75 length of 6-C, lightly aciculate (Fig. 885) Fig. 884 — Dorsal view of head - Cx. iolambdis 5(25). Seta 2-S without secondary tooth; pecten spine with 15 or more fine, lateral denticles (Fig. 886) (Plate Seta 2-S with secondary tooth; pecten spine with fewer than 12 coarser, lateral denticles (Plate Seta 2-S with secondary tooth; pecten spine with fewer than 12 coarser, lateral denticles	in middle (Fig. 882)	abomina (Plate
Fig. 882 — Comb scale - Cx. abominator Fig. 883 — Comb scale - Cx. iolambdis 5(24). Seta 5-C thin, much thinner and 0.5 or less length of seta 6-C, without aciculae (Fig. 884) Seta 5-C stout, about 0.75 length of 6-C, lightly aciculate (Fig. 885) Fig. 884 — Dorsal view of head - Cx. percutor Fig. 885 — Dorsal view of head - Cx. iolambdis 5(25). Seta 2-S without secondary tooth; pecten spine with 15 or more fine, lateral denticles (Fig. 886)		
Fig. 882 — Comb scale - Cx. abominator Fig. 883 — Comb scale - Cx. iolambdis 5(24). Seta 5-C thin, much thinner and 0.5 or less length of seta 6-C, without aciculae (Fig. 884) Seta 5-C stout, about 0.75 length of 6-C, lightly aciculate (Fig. 885) Fig. 884 — Dorsal view of head - Cx. peccator Fig. 885 — Dorsal view of head - Cx. iolambdis 6(25). Seta 2-S without secondary tooth; becten spine with 15 or more fine, lateral denticles (Fig. 886) (Plate		\wedge
Fig. 882 — Comb scale - Cx. abominator Fig. 883 — Comb scale - Cx. iolambdis 5(24). Seta 5-C thin, much thinner and 0.5 or less length of seta 6-C, without aciculae (Fig. 884) Seta 5-C stout, about 0.75 length of 6-C, lightly aciculate (Fig. 885) Fig. 884 — Dorsal view of head - Cx. peccator Fig. 885 — Dorsal view of head - Cx. iolambdis 6(25). Seta 2-S without secondary tooth; becten spine with 15 or more fine, lateral denticles (Fig. 886) (Plate	4	
Fig. 882 — Comb scale - Cx. abominator Fig. 883 — Comb scale - Cx. iolambdis 5(24). Seta 5-C thin, much thinner and 0.5 or less length of seta 6-C, without aciculae (Fig. 884) Seta 5-C stout, about 0.75 length of 6-C, lightly aciculate (Fig. 885) Fig. 884 — Dorsal view of head - Cx. peccator Fig. 885 — Dorsal view of head - Cx. iolambdis 5(25). Seta 2-S without secondary tooth: pecten spine with 15 or more fine, lateral denticles (Fig. 886) (Plate		
Fig. 882 — Comb scale - Cx. abominator Fig. 883 — Comb scale - Cx. iolambdis 5(24). Seta 5-C thin, much thinner and 0.5 or less length of seta 6-C, without aciculae (Fig. 884) Seta 5-C stout, about 0.75 length of 6-C, lightly aciculate (Fig. 885) Fig. 884 — Dorsal view of head - Cx. peccator Fig. 885 — Dorsal view of head - Cx. iolambdis 5(25). Seta 2-S without secondary tooth: pecten spine with 15 or more fine, lateral denticles (Fig. 886) (Plate		
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5(25). Seta 2-S without secondary tooth; pecten spine with 15 or more fine, lateral denticles (Fig. 886)		
5(25). Seta 2-S without secondary tooth; pecten spine with 15 or more fine, lateral denticles (Fig. 886)	· · · · · · · · · · · · · · · · · ·	
886)	Fig. 884 — Dorsal view of head - Cx, peccator	Fig. 885 — Dorsal view of head - Cx. iolambdis
(Plate		
	886)	
	Seta 2-S with secondary tooth: pecten spine with	

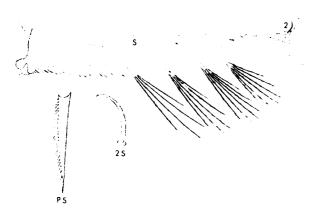


Fig. 886 — Lateral view of siphon - Cx. anips



Fig. 887 - Lateral view of siphon - Cx. peccator

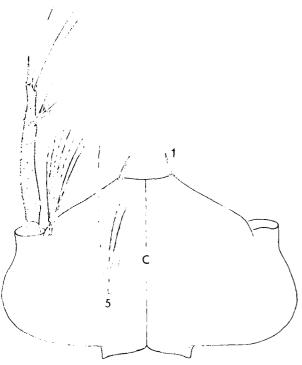


Fig. 888 — Dorsal view of head - Cx, iolambdis

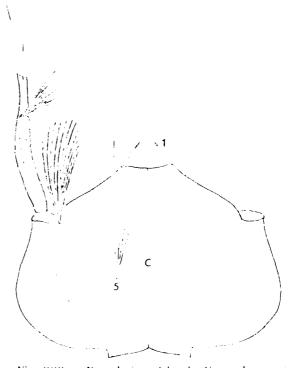


Fig. 889 — Dorsal view of head - Cx. mulrennani

KEY TO FOURTH STAGE LARVAE OF THE GENUS CULISETA

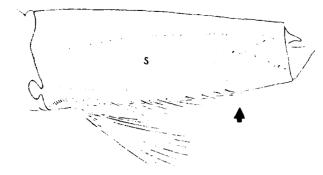


Fig. 890 — Lateral view of siphon - Cs. melanura

Fig. 891 — Lateral view of siphon - Cs. inornata



Fig. 892 - Dorsal view of head - Cs. morsitans

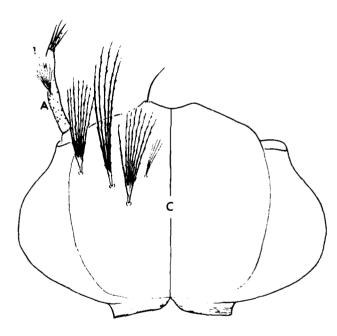
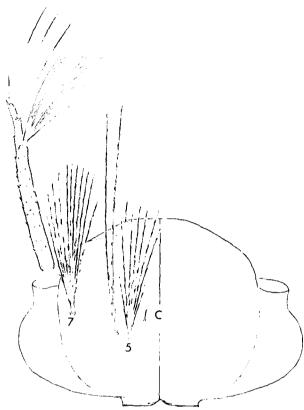


Fig. 894 — Dorsal view of head - Cs. inornata



Fig. 893 — Lateral view of siphon - Cs. morsitans

Fig. 895 - Lateral view of siphon - Cs. inornator



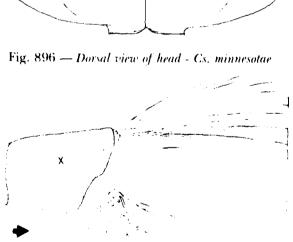
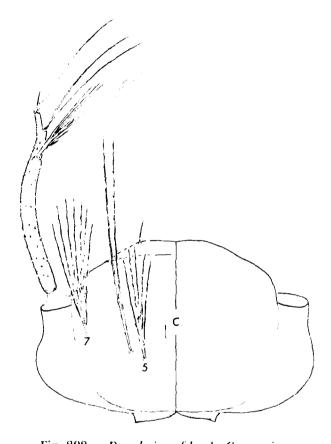


Fig. 897 — Lateral cuew of abdominal segment X - Cs. minnesotai



(Plate 48)

Fig. 898 — Dorsal view of head - Cs. morsitans



Fig. 899 — Lateral view of abdominal segment X - Cs.

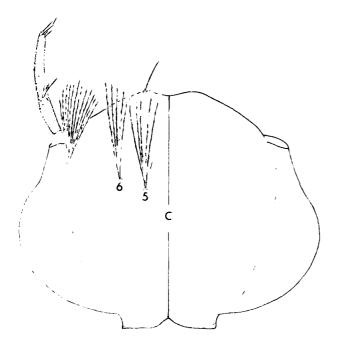
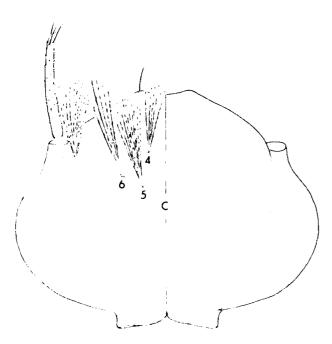
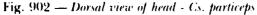


Fig. 900 - Dorsal view of head - Cs. impatiens

Fig. 901 - Dorsal view of head - Cs. inornata





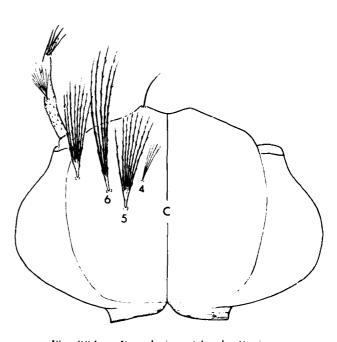


Fig. 904 — Dorsal view of head - Cs. inornata

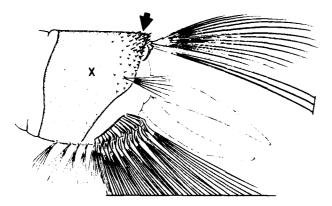


Fig. 903 — Lateral view of abdominal segment X - Cs. particeps

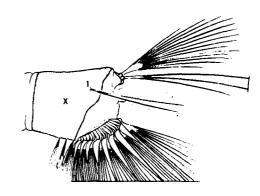


Fig. 905 — Lateral view of abdominal segment X - Cs. inornata

6(5). Seta 1-X with rather strong branches equal to length of saddle or longer (Fig. 906)inornata (Plate 44)

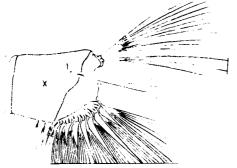


Fig. 906 — Lateral view of abdominal segment X - Cs. inornata

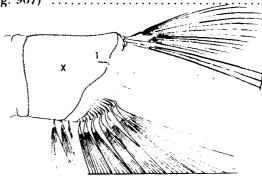


Fig. 907 — Lateral view of abdominal segment X - Cs. incidens

Antenna slender, 9.0 or more length of basal diameter, with fewer, fine spinules on distal 0.5 (Fig. 910); seta 1-M single, much longer than multibranched 2-M (Fig. 911)incidens

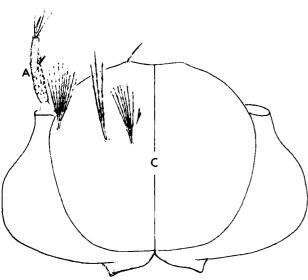


Fig. 908 - Dorsal view of head - Cs. alaskaensis

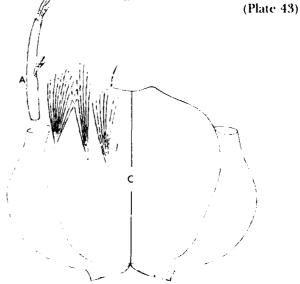


Fig. 910 — Dorsal view of head - Cs. incidens

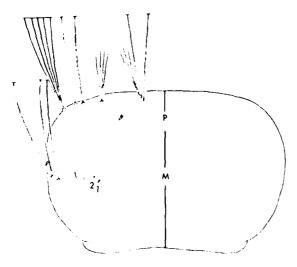


Fig. 909 — Dorsal view of thorax - Cs. alaskaensis

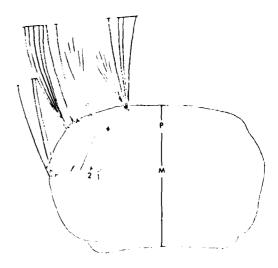


Fig. 911 - Dorsal view of thorax - Cs. incidens

KEY TO FOURTH STAGE LARVAE OF THE GENUS DEINOCERITES

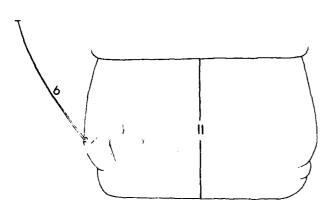


Fig. 912 — Dorsal view of abdominal segment II - De. mathesoni

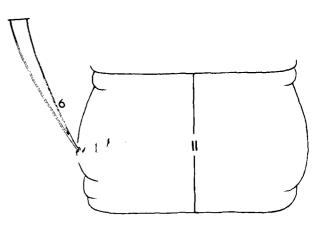


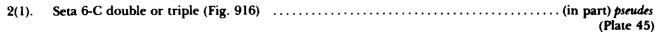
Fig. 914 — Dorsal view of abdominal segment II - De. pseudes



Fig. 913 — Lateral view of siphon - De, mathesom



Fig. 915 — Lateral view of siphon—De. pseudes



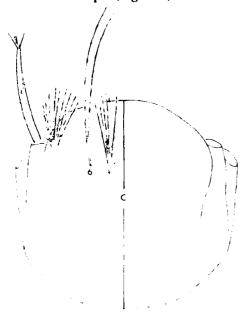


Fig. 916 - Dorsal view of head - De. pseudes

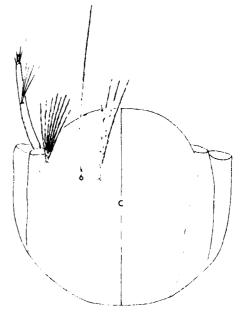


Fig. 917 — Dorsal view of head - De. cancer

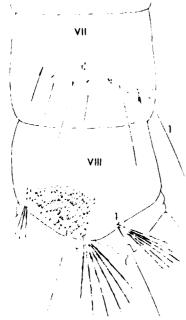


Fig. 918 — Lateral view of abdominal segments VII-VIII - De. pseudes

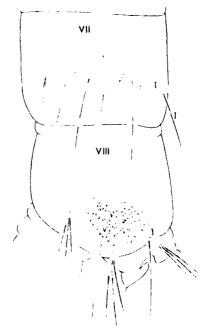
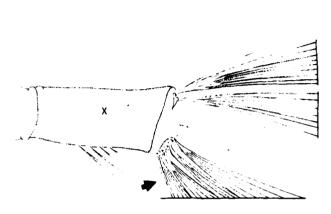


Fig. 919 — Lateral view of abdominal segments VII-VIII - De. cancer

KEY TO FOURTH STAGE LARVAE OF THE GENUS MANSONIA



X

(Plate 46)

Fig. 920 — Lateral view of abdominal segment X - Ma. titillans

Fig. 922 — Lateral view of abdominal segment X - Ma. dyari





Fig. 921 — Comb scale - Ma. titillans

Fig. 923 — Comb scale - Ma. dyari

KEY TO FOURTH STAGE LARVAE OF THE GENUS ORTHOPODOMYLA

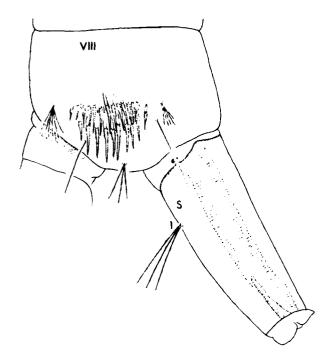


Fig. 924 — Lateral view of siphon and abdominal segment VIII - Or, alba

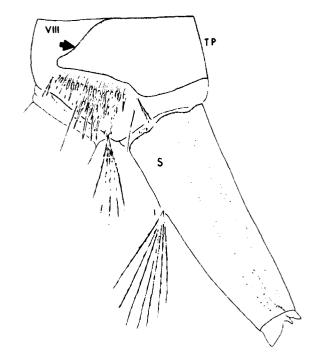


Fig. 925 — Lateral view of siphon and abdominal segment VIII - Or. signifera

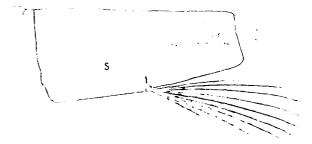


Fig. 926 -- Lateral view of siphon - Or. kummi

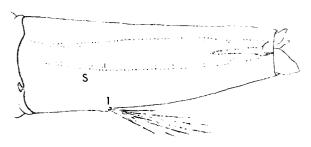


Fig. 928 — Lateral view of siphon - Or. signifera

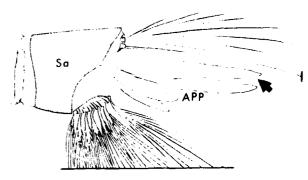


Fig. 927 — Lateral view of abdominal segment X - Or. kummi

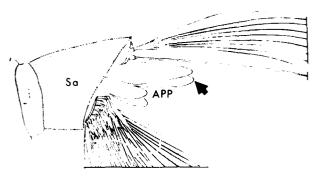


Fig. 929 — Lateral view of abdominal segment X - Or. signifera

KEY TO FOURTH STAGE LARVAE OF THE GENUS PSOROPHORA*

1.	Head capsule truncate anteriorly (Fig. 930); pecten with 12 or more filamentous spines (Fig. 931); antenna small, hardly reaching beyond anterior border of head (Fig. 930) (subgenus <i>Psorophora</i>)
	Head capsule rounded anteriorly (Fig. 932); pecten with fewer than 10 pecten spines, not produced into filaments (Fig. 933); antenna reaching well beyond anterior border of head
	(Fig. 939)

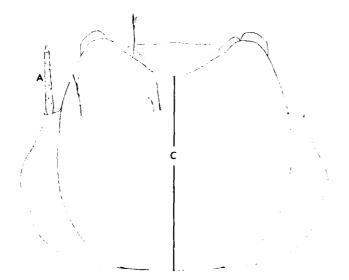


Fig. 930 — Dorsal view of head - Ps. ciliata



Fig. 932 - Dorsal view of head - Ps. discolor



Fig. 931 — Lateral view of siphon - Ps. howardii

Fig. 933 - Lateral view of siphon - Ps. columbiae

2(1).	Seta 1-X with 3,4 branches from near base (Fig. 934)	<i>ciliata</i> ite 43)	
	Seta 1-X single or branched some distance distal to base (Fig. 935)	wardii ne 48)	

^{*}The larva of Ps. mexicana is unknown.

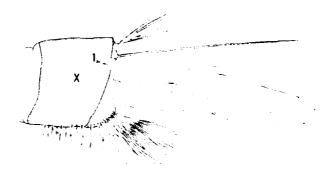


Fig. 934 — Lateral view of abdominal segment X - Ps. ciliata

Fig. 935 — Lateral view of abdominal segment X - Ps. howardii

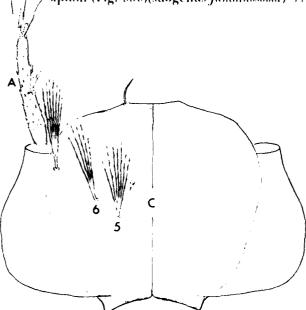


Fig. 936 - Dorsal view of head - Ps. columbiae

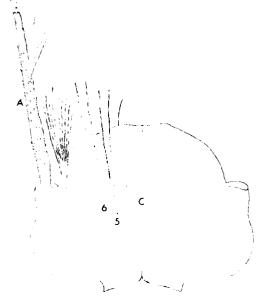


Fig. 938 — Dorsal view of head - Ps. ferox



Fig. 937 — Lateral view of siphon - Ps. discolor

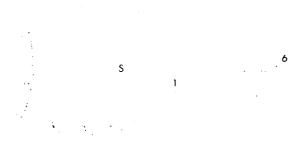


Fig. 939 — Lateral view of siphon - Ps. cyanescens

4(3).	Antenna longer than head, sinuate, somewhat inflated in distal 0.5 (Fig. 940); seta 1-S very
	large, with some branches at least equal to length of siphon (Fig. 941)discolor
	(Plate 45)
	Antenna shorter than head, slightly curved, not inflated (Fig. 942); seta 1-S much shorter
	than length of siphon (Fig. 943)

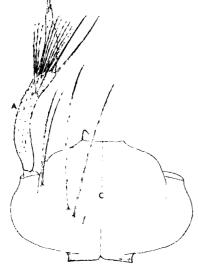


Fig. 940 - Dorsal view of head - Ps. discolor

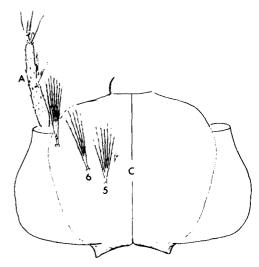


Fig. 942 — Dorsal view of head - Ps. columbiae



Fig. 941 - Lateral view of siphon - Ps. discolor



Fig. 943 — Lateral view of siphon - Ps. columbiae

Setae 5 , 6-C about equal to length of antenna, or longer, single to triple (Fig. 945) 6



Fig. 944 — Dorsal view of head - Ps. columbiae

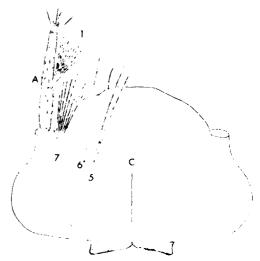
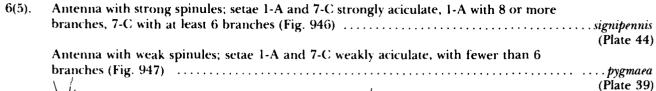


Fig. 945 — Dorsal view of head - Ps. significants



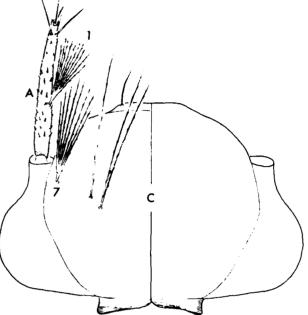


Fig. 946 - Dorsal view of head - Ps. signipennis

Fig. 947 — Dorsal view of head - Ps. pygmaea

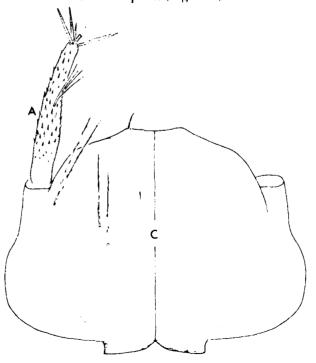


Fig. 948 — Dorsal view of head - Ps. cyanescens

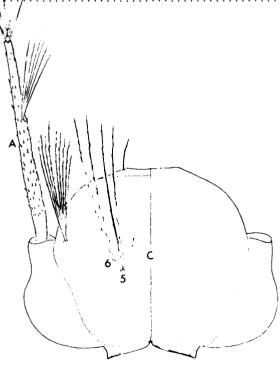


Fig. 950 — Dorsal view of head - Ps. ferox

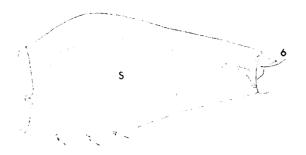


Fig. 949 — Lateral view of siphon - Ps. cyanescens

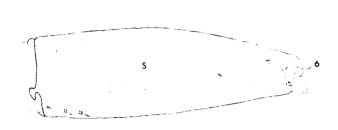


Fig. 951 — Lateral view of siphon - Ps. ferox

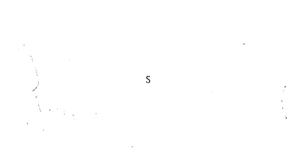


Fig. 952 — Lateral view of siphon - Ps. johnstonii

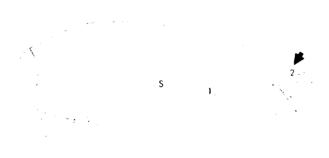


Fig. 954 — Lateral view of siphon - Ps. horrida



Fig. 953 — Lateral view of abdominal segment X - Ps. johnstonn

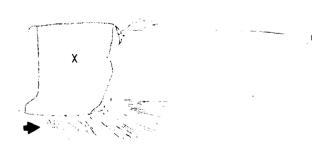


Fig. 955 — Lateral view of abdominal segment X - Ps. horrida

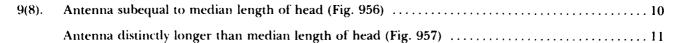




Fig. 956 — Dorsal view of head - Ps. horrida



Fig. 957 — Dorsal view of head - Ps. ferox

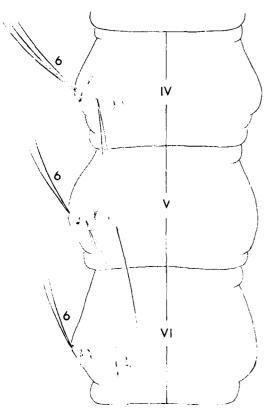


Fig. 958 — Dorsal view of abdomen Ps. horrida

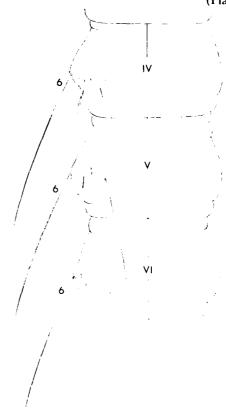
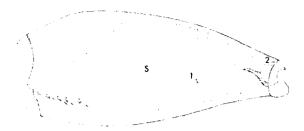


Fig. 960 — Dorsal view of abdomen - Ps. mathesoni



s

Fig. 959 - Lateral view of siphon - Ps. horrida

Fig. 961 - Lateral view of siphon - Ps. mathesoni

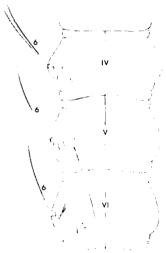


Fig. 962 — Dorsal view of abdominal segments IV-VI - Ps.

ferox

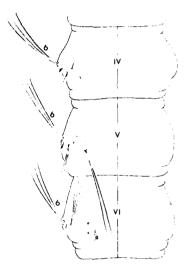


Fig. 964 — Dorsal view of abdominal segments IV-VI - Ps. longipalpus



Fig. 963 — Dorsal view of head - Ps. ferox



Fig. 965 — Dorsal view of head - Ps. longipolpus

KEY TO FOURTH STAGE LARVAE OF THE GENUS URANOTAENIA

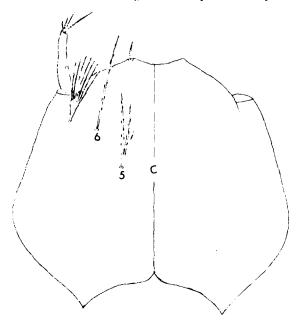


Fig. 966 - Dorsal view of head - Ur. a. syntheta

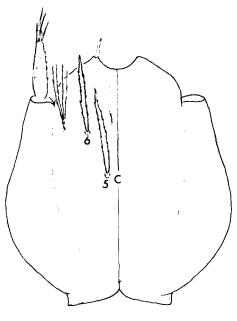


Fig. 967 - Dorsal view of head - Ur. sapphirina

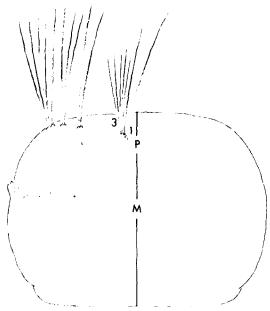


Fig. 968 - Dorsal view of thorax - Ur. lowii



Fig. 970 - Dorsal view of thorax - Ur. sapphirina

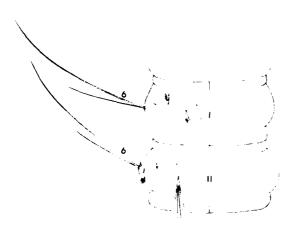


Fig. 969 — Dorsal view of abdominal segments I-II -Ur. lowii

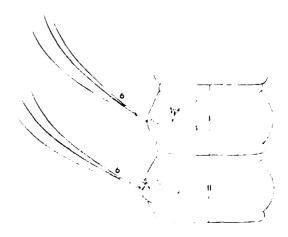


Fig. 971 — Dorsal view of abdominal segments I-II - Ur. sapphirina

KEY TO FOURTH STAGE LARVAE OF THE GENUS WYEOMYIA



Fig. 972 — Lateral view of abdominal segment X - Wy, mitchellii

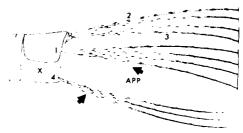


Fig. 974 — Lateral view of abdominal segment X - Wy. smithii

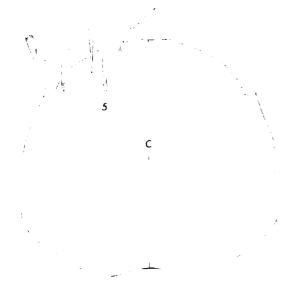


Fig. 973 — Dorsal view of head - Wy, mitchellii

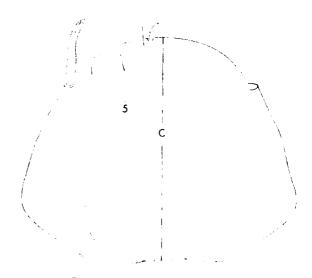


Fig. 975 — Dorsal view of head - Wy, smithii

2(1).	Siphon index about 6.0 (Fig. 976); seta 4-X with 1,2 long and 3,4 short branches (Fig. 977); several setae on siphon double or triple (Fig. 976)vana (Plat		
	Siphon index about 4.0-5.0 (Fig. 978); seta 4-X with 2,3 long, subequal branches (Fig. 979);	2	

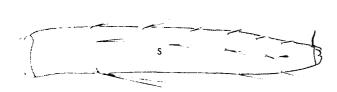


Fig. 976 — Lateral view of siphon - Wy. vanduzeei

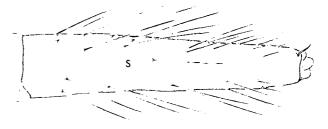


Fig. 978 — Lateral view of siphon - Wy. smithii



Fig. 977 — Lateral view of abdominal segment X - Wy. vanduzeei

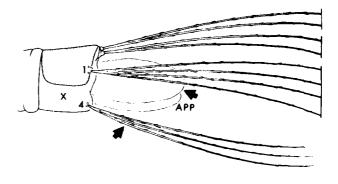


Fig. 979 — Lateral view of abdominal segment X - Wy.

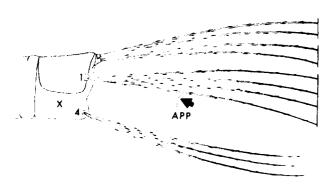


Fig. 980 — Lateral view of abdominal segment X - Wy. smithii

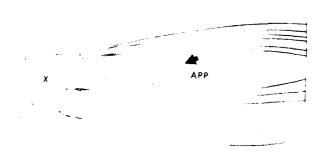


Fig. 982 — Lateral view of abdominal segment X - Wy. haynei

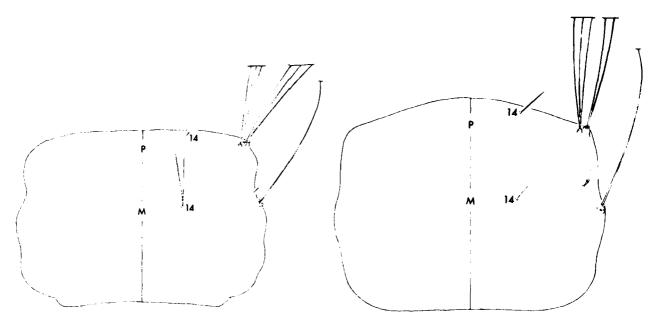


Fig. 981 - Ventral view of thorax - Wy. smithii

Fig. 983 - Ventral view of thorax Wy. haynei

GEOGRAPHICAL DISTRIBUTION OF THE CULICIDAE OF NORTH AMERICA. NORTH OF MEXICO

Closely associated with the identification of any taxon is its geographical distribution. The process of identification will be greatly influenced and assisted by knowing the limits of dispersion of the fauna with which you are working. Obviously, if you determine that a specimen is a particular species and, by checking its distribution, discover that you collected it outside of its known range, you will check it again. In this publication, identification and distribution have been linked, so that the user can check one against the other.

Tables 2-4 list the species and subspecies and the states/provinces from which they have been reported. Table 2 registers 101 species and subspecies from the 24 eastern states of the USA and the District of Columbia; Table 3, 142 species and subspecies from the 24 western states; and Table 4, 79 species and subspecies from Canada and Alaska. Of the 167 taxa, 75 occur in the conterminous 48 states and in Canada/Alaska (the latter two are lumped because they share a similar mosquito fauna); moreover, 87 taxa are found in the 48 states, but not in Canada/Alaska, and 5 are restricted to the latter area. Comparing Canada with Alaska, each has three species not found in the other area, i.e., Canada: Ae. churchillensis, Ae. rempeli and Ae togoi; Alaska: Ae. punctodes, Ae. ventrovittis and Cs. particeps. Ae. churchillensis is a sibling species of Ae. communis (167). Ae. rempeli has also been reported from the Union of Soviet Socialist Republics (154) so has wider distribution than the Canadian records would indicate. Therefore both of these could occur in Alaska. All three Alaskan taxa are potential members of the Canadian fauna (505). As far as Ae. togoi is concerned it belongs to the fauna of the Oriental biogeographical region and is apparently a recent introduction into British Columbia.

Of the 162 taxa distributed in the 48 states, 82 are disseminated in both the eastern and western blocks of states as listed in Tables 2, 3. Only 19 are restricted to the eastern states, while three times that many (61) are confined to the West. In all, 14, or 74%, of those found only in the East, are limited to peninsular Florida with a few extending into southern Georgia. The western states, on the other hand, with their plains, high mountains and deserts offer a wide variety of weather conditions and habitats which has resulted in the development of a diverse mosquito fauna, unique to the West. In fact, three western states, Texas, Arizona and California, have three species each not occurring in any other political unit of the region.

There follows in this section maps which depict the distribution in North America, north of Mexico, of all 167 taxa now known in the culicid fauna, except *Ps. varipes*, for which at present specific distribution is indefinite (see ref. 33). They have been drawn as accurately as possible within the limits of available information on occurrence in the states and provinces of the region, also referred to as political units. Because of the paucity of information about some, the distributional limits of the taxa necessarily had to be estimated. Many are depicted as having discontinuous distribution based on available records and whether it is real or imagined remains to be determined by further study. No attempt has been made to delineate within political units exactly in which parts such widespread species, as *Ae. vexans.* have been found.

In studying the biogeographical distributional patterns exhibited by the mosquito fauna in North America, north of Mexico, two paths of dispersal are very evident. Southern taxa, such as Ae. thibaulti and Ps. ferox (Plates 23, 44), have apparently spread northward using the lowlands of the valley of the Mississippi River and its tributaries and the coastal plain of the Atlantic Ocean. On the other hand, northern species, such as Ae. communis and Ae. hexodontus (Plates 17, 20), have dispersed southward along the high ranges of the western Rocky Mountains. It appears that the eastern Appalachian range lacks either sufficient altitude or possibly favorable breeding sites at the higher elevations to support the northern fauna. However, the Appalachian area has not been studied in depth and its mosquito fauna is not well documented.

Another noteworthy dispersal pattern is the avoidance of the southwestern states by such species as Ae. stieticus, Ae. hendersoni and Cx. restuans; see Plates 25, 16, 38. They have been able to colonize large areas of North America but not the southwest. Apparently, they simply cannot tolerate the dry climate and types of habitats found there. On the other hand, widespread species, such as Ae. dorsalis, Ae. vexans and Cx. tarsalis (see Plates 18, 26, 34) are well adapted and have successfully thrived in the dry areas of the southwestern USA.

One species, Ae. aegypti, presents a peculiar distributional problem. Christophers (531) stated that temperature limits its dispersal, and in North America it is restricted to that part south of a January isotherm of 1.8°C (35°F) and a July isotherm of 23.9°C (75°F). That agrees quite well with our limits indicated on Plate 10 as the extreme range. Rozeboom (532) spoke of three zones in relation to its limit of distribution: the zone of continuous breeding, the zone of egg survival in overwintering diapause, and the temporary summer zone with incursions during the warm months and complete winter dieoff. On Plate 10, then, the usual range marks the limits of continuous breeding, while the extreme range would include the latter two categories. In preparing the map we also considered the records of Morland and Tinker (318) and Tinker and Hayes (467) whose distribution of aegypti was based on actual surveys.

The study of Wood et al. (505) and our own review of the specimens in the U.S. National Museum have lead to the conclusion that An. occidentalis Dyar and Knab does not occur in Canada nor Alaska, as depicted by references 106, 135, 190, 448. Gjullin et al. (192) had previously stated that Alaskan records for An. occidentalis referred to An. earlei. Although we list it for Washington State, there is some doubt that it really is found there (see 505).

An analysis of the known distribution of the North American mosquitoes in other parts of the world reveals that only 48, or 28.7%, are indigenous, not found outside of the region. The number and proportion in each biogeographical region, area or specific country are given below.

REGION	NO. OF SPECIES	PERCENT OF TOTAL	REGION	NO. OF SPECIES	PERCENT OF TOTAL
Indigenous	48	28.7			
Palearctic	271	16.2	Caribbean	5	3.0
Neotropical	36	21.5	& Mexico		
Oriental	1	0.6	Caribbean	6	3.5
Cosmotropical	2	1.2	only Mexico only	40	23.9
Worldwide	1	0.6	Cuba only	1	0.6

⁴Includes Cx. pipieus which also occurs in the Southern Neotropical and Southern Ethiopian regions and two species, Ae. dorsalis and Ae. staticus which likewise are distributed in Mexico.

For the general distribution of each taxon outside our region, if applicable, see the Systematic Index section, page 2. The sources for this dispersal information have been principally Knight and Stone (519) and Knight (518).

In all, 83 species found in North America, north of Mexico, also are distributed in Mexico. Of the total, 33 taxa occur elsewhere in the Neotropical biogeographical region, 2 are cosmotropical, 5 share the Caribbean islands with Mexico, 3 are Holarctic or worldwide and extend into Mexico, and 40 are known only from Mexico outside the target area. The central highlands of Mexico and Baja California are considered part of the Nearctic biogeographical region, while the lowlands are included in the Neotropical zone. Of the 40 taxa, 18 species have Nearctic distribution in Mexico; and the other 22 are dispersed in the Neotropical lowlands although some also may occur in the highlands. In reality then 58, or 34.7% of taxa occurring in North America, north of Mexico, are also part of the Neotropical fauna. The works of Vargas (533) and Vargas and Martínez-Palacios (534) have been helpful in understanding the distribution of the Mexican culicid fauna. Vargas (loc. cit.) also reported that Ae. punctor, Ae. impiger, Ae. spenceru, and Cs. impatiens have been collected in Mexico; but their known distributions are so far removed from Mexico that the records need further confirmation; see Plates 13, 21, 22, 47.

For 34 of the taxa, distributional maps have been previously published and have been used as the basis for those shown here. Their sources have been acknowledged in the captions. The other 136 delimitations shown on the maps are originals, except that the northern extremes of 72 of the 75 Canadian taxa were delimited with the help of maps and information given by Wood et al. (505). The captions of each of the succeeding plates have been organized in the following manner: 1. States of the United States of America, using the official United States Post Office Department two letter abbreviations, tabulated in alphabetical order according to the spelling of the state. 2. Provinces of Canada, using the general accepted abbreviations; 3. In most instances, a reference to taxonomy listed as "Tax.", is followed by the numbers of the references in the bibliography

which contain information on some aspect of the taxonomy of that species. The numbers in parentheses within the lists of states and provinces indicate the bibliographic reference which first cited the occurrence of that species in that political unit.

It must be understood that the starting point for this publication was the monograph of Carpenter and La Casse (106), and that the references, which they cited verifying the occurrence of a species in a given political unit are not repeated here, but the user is referred to their treatise. Additionally, where applicable, there is a notation with substantiating reference if a species was previously reported to have been found in a political unit and subsequently determined that it did not actually occur there. The meaning of the abbreviations for the states and provinces will be found listed on the inside cover of the publication. Listed under the name of each species included in the identification keys is a plate number. This refers to the map on which appears the geographical distribution of that species.

In preparing a presentation of geographical distribution a nagging question to face is the problem of doubtful records. All mosquito specialists who have been responsible for mosquito records in particular political units have an obligation to preserve in an acceptable manner voucher specimens for each species known to occur within its boundaries. Published reports of species found in states/provinces ought to be verified by sample specimens. In the years since 1955 a large number of doubtful records have been settled (see captions on Plates 9-49) and those responsible must be commended. Some still remain in doubt and the following 17 records have not been included either because they are quite far removed from the known range of the species or a specimen may have been collected many years ago and no further evidence exists that the species is indeed a part of the fauna.

SPECIES	POLITICAL UNIT	REFERENCE
Ac. aboriginis	Michigan	229
Ae. canadensis mathesoni	Michigan	481
Ae, canadensis mathesoni	Newfoundland	505
Ae, fulvus pallens	Indiana	392
Ae. increpitus	Manitoba	505
Ae. nigromaculis	Kentucky	106
Ae. pullatus	Michigan	229
Ae. triseriatus	Manitoba	505
Ae. trivittatus	Nova Scotia	505
Cx. apicalis	Illinois	221
Cx. pipiens	Alberta	505
Cx. pipiens	Manitoba	505
Cx. territans	Arizona	382
Cs. incidens	Michigan	229
Cs. incidens	Newfoundland	177
Ma. titillans	Arkansas	225
Ps. mathesoni	Iowa	249

TABLE 2. SYNOPSIS OF THE OCCURRENCE OF MOSQUITO SPECIES IN THE EASTERN UNITED STATES

Mosquito Species	Alabama (55)	Connecticut (42)	Delaware (51)	District of Columbia (36)	Florida (68)	Georgia (58)	Indiana (52)	Kentucky (52)	Maine (36)	Maryland (57)	Massachusetts (45)	Michigan (57)	Mississippi (53)	New Hampshire (44)	New Jersey (57)	New York (58)	North Carolina (53)	Ohio (57)	Pennsylvania (51)	Rhode Island (31)	South Carolina (55)	Tennessee (50)	Vermont (27)	Virginia (51)	West Virginia (24)
Ae. abserratus		*					*	\vdash	*	*	*	*		*	*	*		*	*	*			*		
Ae. aegypti	*	^_	\dashv	*	*	*	*	*	Ĥ	*	Ĥ	Ţ	*			*	*	*			*	*	П	*	
Ae. atlanticus	 		*		<u>^</u>		*	*	1	*			*	Н	*	*	*		\vdash		*	*		*	
Ae. atropalpus	1	*	Ĥ	*		*		*	*	*	*	*	_	*	*	*	*	*	*	*	*	*	*	*	*
Ae. aurifer		*	*		\vdash		*		*	*	*	*	\vdash	*	*	*	T	*	*	*	П		*		
Ae. campestris	-	-	\exists		\vdash	\vdash		t^{-}		<u> </u>		*	\vdash	H	_		\vdash	Г	\vdash		М		Г		П
Ac. c. canadensis	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ae. c. mathesoni	→	-	H		*	*	Ė	 	Ť		T	☆		П			Г	☆			*		Π	\Box	Г
Ae. cantator	-	*	*		-		\vdash	*	*	*	*			*	*	*	T	*	*	*	Г		Π	*	
Ae. cinereus	→	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	
Ae. communis		<u> </u>	Н				 	┢	*	┢	*	*		*	*	*	T		*				Г		П
Ae. decticus	+						\vdash	T	*		*	*		*		*	T	Γ	*	1			Π		
Ae. diantaeus	+				T		1	T	*	1	*	*	Τ_	*		*		T	*			Γ	*		
Ae. dorsalis	+	*	*	┢╌	1	┪	*	T	十	*	*	*	<u> </u>	o	*	*	T	*	*	1	Ī				
Ae. dupreei	*	T	*	\vdash	*	*	*	*	\top			*	*	П	*		*	*			*	*	Π	*	
Ae, euedes	 				1	十	T		1			*		1					Ţ						
Ae. excrucians	1	*	*		T		*	Τ	*	*	*	*		*	*	*		*	*	*			*	oxdot	
Ae. fitchii		*	*		T		*		*	*	*	*		*	*	*		*	*	*	$oxed{oxed}$		*	oxdot	L
Ac. flavescens				Γ	Γ		*					*		*	*	*		*	L			L	上	*	
Ae. fulvus pallens	*				*	*	☆	*		*			*				*			<u> </u>	*	*		*	$oldsymbol{ol}}}}}}}}}}}}}}}}}}$
Ae. grossbecki		*	*		T		*	*	T	*			*		*	*		*	*	_	*	*	*	*	<u></u>
Ac. hendersoni	*	*	*	*	T	*	*	*	*	*	*	*	*	*	*	*	*	*	*	<u> </u>	*	*		*	*
Ae. impiger			T		Τ			T	T			*		*							$oldsymbol{ol}}}}}}}}}}}}}$	L	L		
Ae. implicatus					Τ	T		Π	*	\mathbf{L}^{-}	*	*		*	*	*	L	*		_	$oxedsymbol{oldsymbol{oldsymbol{oldsymbol{eta}}}$	\perp	上	ot	<u> </u>
Ac. infirmatus	*		*		*	*	*	*		*		I_{-}	*				*	<u> </u>			*	*	\perp	<u></u> ★	
Ae. intrudens		*	T	1	Τ	1		T	*		*	*		*	*	*			*	*	$oldsymbol{ol}}}}}}}}}}}}}}$	<u> </u>	1_	$oldsymbol{ol}}}}}}}}}}}}}}}}}}$	<u> </u>
Ae. mitchellae	*		*	*	*	*		*		*		*	*	1_	*	*	*	*	*	_	*	*	上	*	_
Ac. pionips	-	T			T	T	T	Т	*	T		*							L	_	上	\perp	丄	$oldsymbol{ol}}}}}}}}}}}}}}}}}}$	
Ac. provocans		*		T	1	T	Τ		*		*	*		*	*	*			*	4—	\perp	1_	<u></u>		1_
Ae. punctor						\prod	*		*		*	*		*	*	+	+-	ot	*	_	ot	\perp	*	\perp	↓_
Ae. riparius			Γ			\prod						*	4		L	*	ا	*	╃	_	$oldsymbol{\perp}$	<u> </u>	\perp	丄	\downarrow
Ae. sollicitans	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	-	*	+-	*	*	*	*	\perp	*	↓_
Ac. s. spencerii					Ι	Ι	Ι	Ι	\prod			*		L	Ĺ	*	-	*	_	\perp	\perp	1	\perp	\downarrow	↓_
Ae. sticticus	*	*	*	*	*	*	*	★	*	*	*	*	*	*	*	*	*	*	*		<u></u>	*	<u></u> ★	<u></u>	*

^{*} Confirmed Record * Doubtful Record

——————————————————————————————————————		_	_		_	_	_			_		_		г				_			_	_			_
Mosquito Species	Alabama	Connecticut	Delaware		Florida	Georgia	Indiana	Kentucky	Maine	Maryland	Massachusetts	Michigan	Mississippi	New Hampshire	New Jersey	New York	North Carolina	Ohio	Pennsylvania	Rhode Island	South Carolina	Tennessee	Vermont	Virginia	West Virginia
Ae. stimulans		*	*		L		*	*	*	*	*	*	*	*	*	*		*	*	*			*	*	L
Ae. taeniorhynchus	*	*	*	*	*	*				*	*		*	*	*	*	*		*	*	*			*	
Ae. thelcter					*																				
Ae. thibaulti	*	*	*		*	*	*	*		*			*			*	*	*			*	*	Г	*	
Ae. tormentor	*		*		*	*		*		*			*				*	*			*	*			
Ae. tortilis	1				*												T			<u> </u>		\Box	Г	Г	Г
Ae, triseriatus	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ae, trivittatus	*	*	*	*		*	*	*	*	*	*	*		*	*	*	*	*	*	*	*	*		*	*
Ae, vexans	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
An, albimanus					*																	Г			Г
An. atropos	*		Г		*	*				*			*		*		*			Г	*		Г	*	
An, barberi	*		*	*	*	*	*	*		*		*	*		*	*	*	*	*	Г	*	*	Г	*	*
An. bradlevi	*		*		*	*	†		-	*			*	T	*	*	*			┢	*	⇈	一	*	Т
An. crucians	*	*	*	*	*	*	*	*		*	*	*	*	T	*	*	*	*	*	*	*	*	\vdash	*	一
An. earlei	1	*	\vdash		┢	T	t		*		*	*		*	*	*	\vdash			╁		┢	*	\vdash	┢
An. georgianus	*	\vdash	H		*	*	T	\vdash	\vdash		\vdash		*	t			*			 	*	┢	┢─	<u> </u>	\vdash
An. perplexens	*	┢		\vdash	*	*	\vdash		┢╌			\vdash		t			*	*	*	 	-	*		H	_
An. pseudopunctipennis	+~	├	-		╁	├	-	-	├		H	\vdash	*	\vdash		_	┢			\vdash	<u> </u>	*	-	\vdash	_
An. punctipennis	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
An. quadrimaculatus	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	ı	*	*	*	*	*		*	*
An, walkeri	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	Ë
Cq. perturbans	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Cx. atratus	+	├─	H		*	╁	\vdash	\vdash	\vdash		 	 	-	 			\vdash		-	H		\vdash	H		Ë
Cx. bahamensis	+-	\vdash	\vdash		*	\vdash			├	\vdash	\vdash	-	\vdash	-			\vdash	├-		\vdash		\vdash	\vdash	\vdash	_
Cx. erraticus	*	\vdash	*	*	*	*	*	*	-	*		*	*	-	*		*	*	*		*	*		*	*
Cx. iolambdis	+	┢	H		*	<u> </u>			-	-	<u> </u>		<u> </u>	-			 			\vdash		╫	\vdash		Ĥ
Cx. latisquama	+-	\vdash	\vdash		*	 	-	-	<u> </u>	\vdash			 				 	-				╁─	<u> </u>	H	\vdash
Cx. mulrennani	+-	├	Н		*	-	_	-	 	-	\vdash		 	\vdash			\vdash	-		\vdash		╁		-	 -
Cx. nigripalpus	*	├─	Н		₩	*	-	*	 	\vdash	\vdash	-	*				*				*	*		-	-
Cx. opisthopus	+~	 	Н		*	<u> </u>		<u> </u>	<u> </u>	\vdash		 	<u> </u>				 					├-		-	-
Cx. peccator	*	┢	Н		<u>^</u>	*		*	<u> </u>	-		*	*	-	\vdash		*	\vdash	_	-	*	*		*	H
Cx. pilosus	*	-	-	_	*	*	-	*	 	├	\vdash	Ĥ	*	\vdash		_	*				*	┢	H	1	-
Cx. pipiens	*	•	*	*	1	*	*	*	*	*	+	*	*	+	*	+	*	*	*	•	<u>^</u>	*	*	*	*
Cx. quinquefasciatus	*	\vdash	H	*	*	*	<u> </u>	*	<u> </u>	*	Ĥ	<u> </u>	*	Ĥ	~	^	*	*	~	-	*	*	Ĥ	*	*
Cx. restuans	 ^	*	*		*	*	*	*	*	*	*	*	*	*	*	*	*	^ ★	*	*	*	\leftarrow	*	*	*
Cx. restuans Cx. salinarius	 ^	*	^ ★		*	*	*	*	×	<u>^</u>	<u>~</u>	*	*	*	<u>^</u>	<u>^</u>	*	<u>^</u>	*	*	*	*	*	*	*
Cx. tarsalis	 *	<u>~</u>	$\vdash \exists$		*	*	-	×	_	 	Ĥ	*	-	\vdash	× *	_	 ^ -	× ★	*		*	*	Ĥ	Ĥ	ŕ
	<u>^</u>	*	*		*	*	L.	*	4	*				*		*	-	*		*	*	ш	*	┰	4
Cx. territans	<u> </u>		<u> </u>		<u> </u>	×	×	×	×	×	×	×	<u> </u>	×	×	×	<u> </u>	×	×	×	×	<u> </u>		×	×

TABLE 2 (CONTINUED)

											·tts			shire			dina		iia	nd	dina				nia
	Alabama	Connecticut	Delaware	District of Columbia	Florida	Georgia	Indiana	Kentucky	Maine	Maryland	Massachusetts	Michigan	Mississippi	New Hampshire	New Jersey	New York	North Carolina	Ohio	Pennsylvania	Rhode Island	South Carolina	Tennessee	Vermont	Virginia	West Virginia
Cs. impatiens		*							*		*	*		*		*			*				*		L
Cs. inornata	*	*	*	*	*	*	*	*		*	*	*	*	*	*	*	*	*	*		*	*		*	*
Cs. melanura	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*		*	L
Cs. morsitans		*	*				*	*	*	*	*	*		*	*	*		*	*	*			*		L
Cs. minnesotae		*	*				*			*	*	*		*	*	*		*						L	L
De. cancer					*													_	L	<u> </u>			L	<u> </u>	\vdash
Ma. dvari					*	*							<u> </u>	_					L		Ш	_	<u> </u>	igspace	L
Ma. titillans					*								_			_			_	<u> </u>			╙	<u> </u>	╄
Or, alba	*		*	*	*	*	*	*		*	L	*	*		*	*	*	*	*	-		*	ota	*	L
Or, signifera	*	*	*	*	*	*	*	*		*	*	*	*	*	*	*	*	*	*	-	*	*	<u> </u>	*	*
Ps. ciliata	*	*	*	*	*	*	*	*	L	*	*	*	*	*	*	*	*	*	*	*	*	*	↓_	*	*
Ps. columbiae	*		*	*	*	*	*	*		*	*	L	*	<u> </u>	*	*	*	*	*	↓	*	*	↓_	*	*
Ps, cyanescens	*	_	*		*	*	*	*	L	*	<u> </u>	<u> </u>	*	igspace	*	<u> </u>	*	*	_		*	*	₩	*	╀
Ps. discolor	*		*	*	*	*	*	*	L	*	_		*	_	*	<u> </u>	*	*	L	₩	*	*	↓_	*	Ļ
Ps. ferox	*	*	*	*	*	*	*	*	L	*	*	*	*	*	*	*	*	*	*	+	*	*	igspace	*	*
Ps. horrida	*		*	*	*	*	*	*	L	*		*	₩-	L	 _	L	*	*	*	↓_	*	*	igspace	*	\downarrow
Ps. howardii	*		*	*	*	*	*	*	L	*	_	_	*	↓_	<u> </u>	<u> </u>	*	*	Ļ	1	*	*	—	*	╄
Ps. johnstonii					*			L	Ĺ	L		_	_	L	<u> </u>	L_		<u> </u>	L	$oldsymbol{oldsymbol{oldsymbol{eta}}}$	<u> </u>	L	igspace	\downarrow	\downarrow
Ps. mathesoni	*		*		*	*	*	*	L.	*	L		*	<u> </u>	*	*	*	*	<u> </u>	\perp	*	*	↓_	*	\downarrow
Ps. pygmaca					*	L	L			L			\perp	<u> </u>		_	<u> </u>	ــــ	_	↓_	<u> </u>	Ļ	\perp	 	ot
Ps. signipennis					\perp	L	L	*	L.	$oldsymbol{ol}}}}}}}}}}}}}}}}}}$	<u> </u>		↓_	$oldsymbol{ol}}}}}}}}}}}}}}}}}}$	1_	Ļ	_	↓_	ļ_	 	<u> </u>	*	igspace	igaplus	\downarrow
Tx. r. rutilus		_		L	*	*	<u> </u>	1	_	\perp	ļ	<u> </u>	↓_	\perp	$oxed{\bot}$	<u> </u>	ļ.,	 	1	\perp	*	<u> </u>	↓_	+	
Tx. r. septentrionalis	*	*	*	*	*	*	*	*		*	<u> </u>		*	\perp	*	*	*		*	↓_	*	*	↓_	*	*
Ur, lowii	*	1		L.	*	*	1	↓_	1	\downarrow	<u> </u>		*	\perp	↓_	<u> </u>	*	+	↓_	+	*	 		+	+
Ur. sapphirina	*	*	*	*	┷	*	+	*	_	*	+	*	*	*	*	*	╄		*	 *	*	*	↓ *	*	+
Wy. haynei	*				*	*	+		\perp	*	_	\downarrow	\downarrow	\downarrow	_	_	*	_	↓_	\perp	*	_	\perp	*	\perp
Wy. mitchellii					*	*				1_	_	\perp	1	\perp	_	<u> </u>		-	\downarrow	 	 	\vdash	↓_	\perp	\downarrow
Wy. smithii		*	*	_	<u>_</u>	L	*	_	*	*	*	*	\downarrow	*	*	*	 	*	*	*	┼	\vdash	4	+	+
Wy. vanduzeci		1		1	*										1			\perp	L	\perp	$oldsymbol{ol}}}}}}}}}}}}}}}}}}$	1_	$oldsymbol{oldsymbol{oldsymbol{oldsymbol{\bot}}}$	上	丄

TABLE 3. SYNOPSIS OF THE OCCURRENCE OF MOSQUITO SPECIES IN THE WESTERN UNITED STATES

	1	_	ı		<u> </u>				Г		Г			_						<u> </u>			ı	_
															(2)	North Dakota (32)			South Dakota (40)			3)		
		(<u>[2</u>	6					စ္ခ	51)		(G)	6	_	(5	ta (59)		ta (4)	9	(91
	43)	(5,	F) 4	(4)	<u>(</u>	$\widehat{\mathbf{x}}$		6	<u>.</u>	a ((54	7	1	35)	cic	ŧko	я (47	ko	<u>2</u>		5) =	3 (4
	[E	sas	ii.	op	(46	(3)	48)	(E)	an	SOL	Ξ.	na	Ska Ka	(H)	le,	Di	Ξ.) [I	1	<u>x</u>	(47)	gu	nsi	ΙΞ̈́
	[5]	E	5	Ora	ho	ioi) e	ısa	isi	nıc	S	nta	EE	12	3	rth	lah	ာနှင့	Ę	ças	4	shi	9	
Mosquito Species	Arizona (43)	Arkansas (54		Colorado (42)	Idaho (49)	Illinois (58)	Iowa (48)	Kansas (52)	Louisiana (56)	Min	Missouri (54)	Montana (46)	Nebraska (49)	Nevada (35)	New Mexico (55)	No.	Oklahoma (59)	Oregon (47)	Sol	Texas (82)	Utah (47)	Washington (43)	Wisconsin (50)	Wyoming (46)
Ac. aboriginis		Ť			*									 				*				*	 	t
Ae. abserratus		-	 			*				*													*	
Ae. aegypti		*				*		*	*		*						*			*				
Ac. aloponotum		_																*				*		
Ae. atlanticus		*		Н		*		*	*		*						*			*		\Box		
Ae. atropalpus	-			\vdash	-				 	*	T	\vdash	\vdash									\vdash	*	
Ac. aurifer						*	*			*		Γ										Г	*	
Ae. bicristatus			*																					
Ac. bimaculatus																				*				
Ac. brelandi									Γ		Γ_			İ						*		Г		
Ac. burgeri	*										Г													
Ae. campestris			*	*	*		*			*		*	*	*	*	*		*	*	*	*	*	*	*
Ae. c. canadensis		*			*	*	*	*	*	*	*	*	*	T		*	*		*	*		*	*	*
Ae. cutaphylla	*		*	*	*							*		*	*			*			*	*		*
Ac. cinereus		*		*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*		*	*	*	*
Ae. communis			*	*	*					*		*		*	*			*			*	*	*	*
Ae, decticus										*														
Ac. deserticola			*			1																		
Ac. diantacus										*		*											*	*
Ac. dorsalis	*		*	*	*	*	*	*		*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ac. dupreer		*				*	*	*	*		*						*			*				
Ac. epactius	*	*		*				*	*		*				*		*			*	*			
Ac. cuedes						<u> </u>				*														
Ac. excrucians				*	*	*				*		*			*	*		*			*	*	*	*
Ac fitchii	*		*	*	*	*	*			*		*	*	*	*	*		*			*	*	*	*
Ac. flavescens			*	*	*	*	*	*		*	*	*	*			*		*	*		*	*	*	*
Ac. fulvus pallens		*				*			*		*						*			*				
Ac grossbecki		*	_			*			*		*									*			*	
Ac hemiteleus		<u> </u>	*															*						
Ac. hendersom		*		*	*	*	*	*	*	*	*	*	*		*		*	*	*	*	*		*	*
Ac hexodontus			*	*	*							*	Ī	*	*			*			*	*		*
Ac impiger				*	*							*						*			*	*		*
Ac implicatus	*		T	*	*		*	Γ		*		*	*		*			*			*	*	*	*

* Confirmed Record — Doubtful Record

	_				1						_	ι –	·	·					_	1	1	<u> </u>		
Mosquito Species	Arizona	Arkansas	California	Colorado	Idaho	Illinois	Iowa	Kansas	Louisiana	Minnesota	Missouri	Montana	Nebraska	Nevada	New Mexico	North Dakota	Oklahoma	Oregon	South Dakota	Texas	Utah	Washington	Wisconsin	Wyoming
Ae. increpitus	*		*	*	*							*	*	*	*			*	*		*	*		*
Ae. infirmatus		*		\Box		*			*		*									*				
Ae. intrudens				*	*					*		*		_		*		*	*		*	*	*	*
Ae. melanimon			*	*	*							*	*	*	*			*			*	*		*
Ae. mercurator	T				*							*												*
Ae. mitchellae		*	Г		Γ	*		*	*						*		*			*				Г
Ae. monticola	*	Γ		1								Ţ.			*									
Ae. muelleri	*	Γ			T									Г	*			Г		*				Г
Ae. nevadensis					*									*				*			*	*		*
Ae. nigromaculis	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*		*
Ae. niphadopsis			*		*									*				*		Γ	*			*
Ae. papago	*	Γ																						
Ae. pionips	1		\vdash	*	*					*		*				*		*				*		*
Ae. provocans				1	*					*		*										*	*	
Ae. pullatus	*	Γ	*	*	*							*		*	*			*		Π	*	*		*
Ae. punctor				*	*	*	*			*		*				*						*	*	*
Ae. purpurcipes	*					İ																		\Box
Ae. riparius							*			*	*	*				*				Γ			*	Г
Ae. scapularis		1				1														*				
Ae. schizopinax	\top	Ì	*	*	*							*		*	*			*			*			*
Ac. sierrensis			*		*							*	1	*				*			*	*		\vdash
Ae. sollicitans	*	*			1	*	*	*	*	1	*	<u> </u>	*	_	*	*	*		*	*	-	ļ		\vdash
Ac. s. idahoensis				*	*					_		*	*	*	*	*		*	*		*	*		*
Ae. s. spencerii	1			1		*	*	*		*		*	*			*	*		*				*	*
Ae. squamiger			*																					
Ae. sticticus		*	*	*	*	*	*	*	*	*	*	*	*			*	*	*	*	*	*	*	*	*
Ae. stimulans		<u> </u>		1	†	*	*	*		*	*	1	*						*	\Box		!	*	\top
Ae. taeniorhynchus	*	*	*	<u>† </u>				*	*							<u> </u>	*			*	-			\top
Ac. theleter	t	 		†		†			†-	<u> </u>			I^-		*	厂	*			*				\vdash
Ac. thibaulti	1	*	 -		<u> </u>	*		1	*		*			†						*				\vdash
Ac. tormentor		*		1	† –	*			*	1	*	† <u> </u>	†			\vdash	*			*	T		_	
Ac. triscriatus		*	 			*	*	*	*	*	*	一	*	<u> </u>			*			*		\vdash	*	_
Ac. trivittatus	*	*	1	*	*	*	*	*	*	*	*	*	*		*	*	*		*	*	*	1	*	*
Ac. varipalpus	*	<u> </u>		†	<u> </u>			<u> </u>				T			_						*			
Ac. ventrovittis	*	<u> </u>	*	\vdash	*	†	 	_				1						*			*	*	1	*
Ac. vexans	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ac. zoosophus		*		1				*	*			T					*			*				1
An. albimanus		I^-		t																*				

T. ADIA: 3 (CONTINCTIO)	_				-1						\neg	_		\neg	\neg		_							
Mosquito Species	Arizona	Arkansas	California	Colorado	Idaho	Illinois	Iowa	Kansas	Louisiana	Minnesota	Missouri	Montana	Nebraska	Nevada	New Mexico	North Dakota	Oklahoma	Oregon	South Dakota	Texas	Utah	Washington	Wisconsin	Wyoming
An. atropos									*											*				
An. barberi		*				*	*	*	*	*	*		*				*		*	*			*	
An. bradleyi									*											*				
An. crucians		*				*	*	*	*		*				*		*			*			*	
An. carlei				*	*		*	*		*		*	*	*		*			*		*	*	*	*
An. franciscanus	*		*	*				*					*	*	*		*	*		*	*			*
An. treeborni	*	T	*	*	*		Г					*		*	*			*		*	*	*		*
An. georgianus	1						Γ		*															
An. judithae	*														*					*				
An. occidentalis			*	<u> </u>														*				*		
An, pseudopunctipennis	T	*						*	*		*				*		*			*	L.			L
An. punctipennis		*	*	*	*	*	*	*	*	*	*	*	*		*	*	*	*	*	*		*	*	*
An. quadrimaculatus		*				*	*	*	*	*	*		*			*	*		*	*			*	
An. walkeri		*				*	*	*	*	*	*		*			*			*	*			*	
Cq. perturbans		*	*	*	*	*	*	*	*	*	*	*	*		*	*	*	*	*	*	*	*	*	*
Cx. abominator					Г															*		Π	П	
Cx. anips	1	T	*	t	1			T										Ī			\Box			
Cx. apicalis	*		*		Τ	☆		T						*	*		*	*		*	*			
Cx. arizonensis	*	1		Ì			Ì	\top																
Cx. boharti	†	1	*	1	*	Γ	1							*				*			Γ	*	Γ	Γ
Cx. chidesteri	T^-	1	1	t		1														*				
Cx. coronator	*	+	1	l	1	\top	1		1	†				Ì	*		1			*	T	\Box		
Cx. declarator	†~	\top	1	1	†	T	T	1		 	✝			1						*	T			
Cx. crraticus	1	*	1		1	*	*	*	*	*	*		*		*		*		*	*	T		*	
Cx crythrothorax	*	\top	*	*	*					1			T	*	*					*	*			
Cx. interrogator		1				T^-		1						I^-						*	Т	Π		
Cx mgripalpus	*	1		1	1	1			*	<u> </u>	1			<u> </u>			*			*				
Cx. peccator	T	*	T		1	*		*	*		*						*			*				
Cx. peus	*	1	*		1			T						*	*		*	*	Ţ	*	Т	*	П	
Cx. pilosus	1	T	1	1		T			*											*	Г	\prod		
Cx. pipicus		*	*	*	*	*	*	*		*	*	*	*			*	*	*	*		*	*	*	*
Cx quinquetasciarus	*	*	*		Ì	*	*	*	*		*		*	*	*		*	Ť.		*	*			
Cx reevesi			*			T																I^{-}		
Cx. restuans	*	*	*	*	*	*	*	*	*	*	*	*	*	i	*	*	*	*	*	*	*		*	*
Cx salmarius	1	*	1	*	*	*	*	*	*	*	*		*		*	*	*		*	*		T	*	*
Cx. tarsalis	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Cx territans	+	*	*	*	*	*	*	*	*	*	*	*	*	*			*	*	*	*	*	*	*	*

	_	г –	_	_		_	<u> </u>	_	_									·	г —		_	1	_	
Mosquito Species	Arizona	Arkansas	California	Colorado	Idaho	Illinois	Iowa	Kansas	Louisiana	Minnesota	Missouri	Montena	Nebraska	Nevada	New Mexico	North Dakota	Oklahoma	Oregon	South Dakota	Texas	tah	Washington	Wisconsin	★ Wyoming
Cs. alaskaensis	₹.	<	<u> </u>	<u>></u>	<u>1</u>	E		7	Ë	<u> </u>		*	<u> </u>	<u>≺</u>	4	_	\mathcal{C}	<u> </u>	S		1	<u> </u>	₽	
Cs. impatiens		-	*	*	*		*	\vdash			*	*	*	<u>^</u>	*			*	*		*	*	*	*
Cs. incidens	*		<u>^</u>	*	*			_		_	^	<u>^</u>	*	<u>^</u>	*	*	*	<u>^</u>	<u>^</u>	*	<u>^</u>	<u>^</u>	Ĥ	*
Cs. inornata	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	<u>^</u>	<u>^</u>	*	*	*	*	*
Cs. melanura	<u> </u>	*	Ĥ	_	_	*	*	*		*	*	_	*	-		^	*	-	Î	*	Ĥ	_	*	Ê
Cs. minnesotae	-	1	_		*	*	*	Ĥ	<u> </u>	*	Ĥ	*					Ĥ	*		Â	*	*	*	\vdash
Cs. morsitans	-	-	-	_	*	*	*			*		*	\vdash	Н		*		<u>^</u>	*		*	_	*	╁
Cs. particeps	*	-	*			Ĥ	<u> </u>		-			<u> </u>	-				\vdash	*			Ĥ	*	1	\vdash
De. mathesoni	l ·		<u> </u>							-		-	-				\vdash	<u> </u>		*	 	<u> </u>	\vdash	\vdash
De. pseudes	╁	┢	-	-			_	\vdash				-		Н			Н	_		*			\vdash	
Hg. equinus			 -			_	┢	<u> </u>	\vdash			 	-	H				Н		*	<u> </u>			\vdash
Ma. titillans		\vdash	┝			\vdash	<u> </u>	-		-				\vdash				-	 	*	\vdash	-		\vdash
Or. alba		*	 	ļ		*	*	*	*		*	一	*	_	*		*	<u> </u>	 	*	┢		\vdash	\vdash
Or. kummi	*		 				H	<u> </u>				H		-	*			Т	_				\vdash	
Or. signifera	*	*	*			*	*	*	*	*	*	 	*		*		*	*	*	*	*	-	*	一
Ps. ciliata		*	-	-	-	*	*	*	*	*	*	-	*		*		*		*	*		\vdash	*	\vdash
Ps. columbiae	† "	*	*	*	-	*	*	*	*	*	*		*	*			*		*	*	\vdash	-	T	1
Ps. confinnis complex	*		会			<u> </u>				-				-	*						\vdash	_	\vdash	\vdash
Ps. cyanescens	†	*	<u> </u>		-	*		*	*	_	*	 	*	-	*		*		<u> </u>	*	<u> </u>		┢┈	
Ps. discolor	*	*	_	\vdash		*	*	*	*		*		*	\vdash	*	_	*		-	*	\vdash			
Ps. ferox		*			Г	*	*	*	*	*	*	<u> </u>	*	一			*		*	*			*	T
Ps. horrida		*		-		*	*	*	*	*	*		*	_			*		*	*			*	<u> </u>
Ps. howardii	*	*		i	_	*		*	*		*		*				*			*				
Ps. longipalpus		*						*	*	_	*		*				*		*	*			\Box	\top
Ps. mathesoni		*				*			*		*		· · · · ·				*			*			*	尴
Ps. mexicana	† –	Ì			<u> </u>				ļ	_										*				\vdash
Ps. signipennis	*	*	*	*			*	*			*	*	*	*	*	*	*		*	*	*	-	<u> </u>	*
1x. r. septentrionalis		*				*		*	*		*						*			*		1		
Tx, sp.	*					-																		Г
Ur. a. anhydor	*		*										<u> </u>	*										
Ur a syntheta	Ì	*										T -	<u> </u>		*		*			*				
Ur lown		*							*								*			*				
Ur sappliirina		*				*	*	*	*	*	*		*		*	*	*		*	*			*	Г
Wy smithii						*				*													*	

TABLE 4. SYNOPSIS OF THE OCCURRENCE OF MOSQUITO SPECIES IN CANADA AND ALASKA

Mosquito Species	Alaska (32)	Alberta (41)	British Columbia (46)	Labrador (26)	Manitoba (43)	New Brunswick (28)	Newfoundland (15)	Northwest Territories (28)	Nova Scotia (25)	Ontario (57)	Prince Edward Island (18)	Quebec (52)	Saskatchewan (40)	Yukon (26)
Ae. aboriginis	*		*											
Ac. abserratus				*	*	*	*		*	*	*	*		
Ac. aloponotum			*											
Ac. atropalpus				*						*		*		
Ac. aurifer						*				*		*		
Ac. campestris		*	*		*					*		*	*	*
Ac. c. canadensis	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ae. c. mathesoni							☆							
Ac. cantator				*		*	*		*		*	*		
Ac. cataphylla	*	*	*										*	*
Ae. churchillensis		*			*									
Ac. cinereus	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ac. communis	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ae. decticus	*	*		*	*			*		*		*		П
Ac. diantaeus	*	*	*	*	*			*	*	*		*	*	*
Ae. dorsalis		*	*		*	*				*		*	*	\Box
Ac. cuedes	*	*	*		*			*	*	*		*	*	
Ac. excrucians	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ae. fitchii	*	*	*	*	*	*	*	*	*	*	*	*	*	*
Ac. flavescens	*	*	*	*	*		Г	*		*		*	*	*
Ac. grossbecki										*				
Ac. hendersoni			*		*					*		*	*	\Box
Ac. hexodontus	*	*	*	*	*			*		*		*	_	*
Ac. impiger	*	*	*	*	*			*		*		*	*	*
Ac. implicatus	*	*	*	*	*			*		*	*	*	*	*
Ac. increpitus		*	*		: 7	-				Г		Г	*	_
Ac. intrudens	*	*	*	*	*	*	*		*	*	*	*	*	
Ac. melanimon		*	*						Г				*	
Ac. mercurator	*	*	*		*			*		*		*	*	*
Ac. nigripes	*		*	*	*		*	*		T		*		*
Ac. nigromaculis		*			*								*	
Ac. pionips	*	*	*	*	*			*		*		*	*	*
Ac. provocans	-	*	*		*	*		*	*	*	*	*	*	\Box
Ac. pullatus	*	*	*	*			*	*		1		*	_	*
					1		.	L						

* Confirmed Record — Doubtful Record

TABLE 4 (CONTINUED)

				_			_					_		
Mosquito Species	Alaska	Alberta	British Columbia	Labrador	Manitoba	New Brunswick	Newfoundland	Northwest Territories	Nova Scotia	Ontario	Prince Edward Island	Quebec	Saskatchewan	Yukon
Ae. punctor	*	*		*	*	*	*	*	*	*		*	*	*
Ae. rempeli		Г					\vdash	*	\vdash	*		*	_	
Ae. riparius	*	*	*		*	*	Г	*	*	*		*	*	*
Ae. schizopinax		*					H			H			┢	
Ae. sierrensis		H	*						\vdash	Г				
Ae. sollicitans		Г				*			*	*	*			
Ae. s. idahoensis			*			<u> </u>	Г		\vdash			\vdash		\vdash
Ae. s. spencerii		*	*		*				_	*			*	
Ae. sticticus		*	*	*	*	*	 		H	*		*	*	_
Ae. stimulans		 	 		*	*	*		*	*	*	*	_	
Ae. thibaulti		┢			\vdash				\vdash	*				t
Ae. togoi			*		 	\vdash				 		\vdash		\vdash
Ae. triseriatus	 	-			☆	*	Н			*	-	*		
Ae. trivittatus	<u> </u>			\vdash	*				☆	*		*	\vdash	
Ae. ventrovittis	*		 		-				-				 	<u> </u>
Ae. vexans	★	*	*		*	*	-		*	*	*	*	*	*
An. barberi			.	-		 	Н			*		*	 	H
An. earlei	*	*	*	*	*	*	\vdash	*	*	*	*	*	*	*
An. freeborni			*		-	H	-			-	<u> </u>			-
An. punctipennis		H	*		*	*	\vdash		*	*		*		┢
An. quadrimaculatus		\vdash					\vdash		-	*		*		\vdash
An. walkeri		\vdash	 	<u> </u>	*	*	-		*	*	_	*	*	
Cq. perturbans		*	*	 	*	*	\vdash		*	*	*	*	*	┢╌
Cx. pipiens		☆	*	-	公	*			*	*		*		
Cx. restuans		*	<u> </u>		*	*	┢		*	*		*	*	-
Cx. tarsalis		*	*		*	-		*		*		-	*	\vdash
Cx. territans	*	*	ь—	*	*	*	\vdash	*	*	*		*	*	*
Cs. alaskaensis	*	↓		*	*	<u> </u>		*			-	*	*	*
Cs. impatiens	*	*	L	*	*	*	*	*	-	*		*	*	*
Cs. incidens	*	*	*	-	-	-	☆		-	-		-	*	*
Cs. inornata		*	*		*	<u> </u>	H	*		*		*	*	*
Cs. melanura		-	H	┢╌		<u> </u>	-			*		*		<u> </u>
Cs. minnesotae	*	*	*		*	 		L	-	*		*	*	<u> </u>
Cs. morsitans	*	*	\vdash	*	*	*	*	*	*	*		*	*	*
Cs. particeps	<u>^</u>	┝	Ë	<u> </u>	-	 	Ĥ			<u> </u>	 ~	Ë	Ë	Ë
Or. alba		\vdash		\vdash			-		├	*		*	\vdash	\vdash
Or. signifera		\vdash			-	-	-		-	*		Ë	\vdash	├
Ps. ciliata		\vdash	Ļ			+-	\vdash		 	*		*		\vdash
Ps. columbiae		-		-	-	-			\vdash	<u>^</u>		-	<u> </u>	\vdash
			L	<u> </u>	L	<u> </u>	Ш			ட்		L	<u> </u>	

TABLE 4 (CONTINUED)

	Alaska	Alberta	British Columbia	Labrador	Manitoba	New Brunswick	Newfoundland	Northwest Territories	Nova Scotia	Ontario	Prince Edward	Quebec	Saskatchewan	Yukon
Ps. ferox		i								*			Π	
Ps. signipennis		Γ					Г					П	*	
Tx. r. septentrionalis							Г		Γ	*		Г	Г	П
Ur. sapphirina			\Box						Π	*		*		
Wy. smithii				*	*	*	*		*	*	*	*	*	

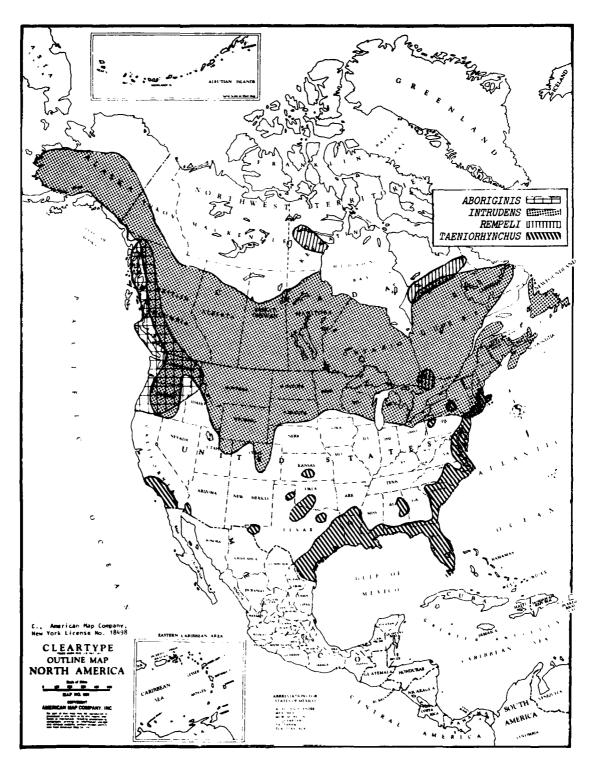


Plate 9. Distribution of Aedes aborigmis - USA: AK, ID, OR, WA (106); CANADA: BC (106), Not in SASK (380); Tax. 279, 505. Aedes intrudens - USA: AK, CO, CT, ID, ME, MA, MI, MN, MT, NH, NY, ND, OR, PA, RI, SD, UT, WA, WI, WY (106), NJ (133); CANADA: ALTA, BC, LAB, MAN, NB, NS, PEI (106), NFLD (360), ONT (27), PQ (155), SASK (380); Tax. 279, 505. Aedes rempeli - CANADA: NWT, PQ (106), ONT (505); Tax. 154, 279, 434, 505. Aedes taeniorhynchus - USA: AL, AR, CA, CT, DE, DC, FL, GA, LA, MD, MA, MS, NJ, NY, NC, PA, RI, SC, TX, VA (106), AZ (385), KS (344), NH (74), OK (215, 225), Not in NM (502); Map modified after Knight (248); Tax. 34, 279.

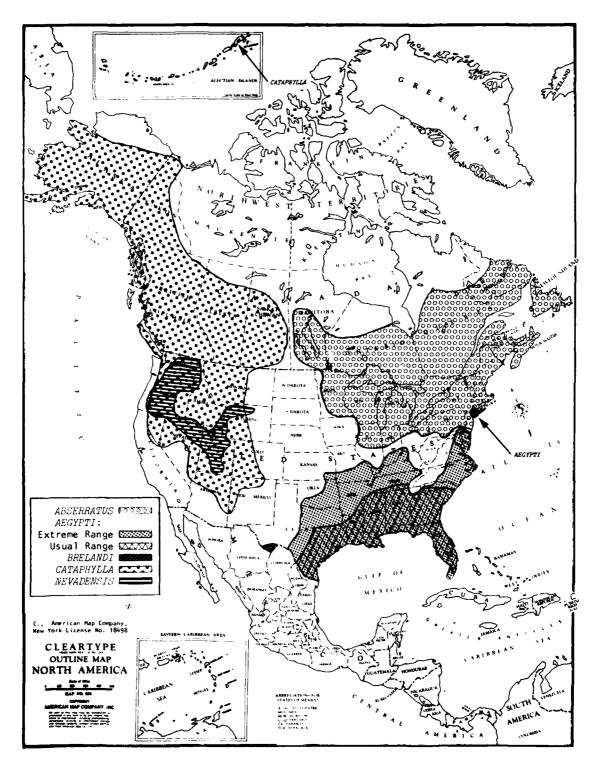


Plate 10. Distribution of Aedes abserratus - USA; CT, IL, ME, MA, MI, MN, NH, NJ, NY, OH, PA, RI, VT, WI (106), IN (415), MD (50); CANADA; LAB, NS, ONT, PEI (106), MAN (70), NB (529), NFLD (478), PQ (288); Tax. 279, 478, 505. Aedes aegypti - USA; AL, AR, DC, FL, GA, IL, IN, KS, KY, LA, MS, MO, NC, OK, SC, TN, TX, VA (106), MD (467), NY (40), OH (Berry & Parsons, in litt. 1978); Map modified after Morland & Tinker (318); Tax. 34, 279. Aedes brelandi - USA; TX (514); Tax. 514. Aedes cataphylla - USA; CA, CO, ID, MT, OR, UT, WA, WY (106), AK (192), AZ, NV (382), NM (340); CANADA; AL FA, BC, SASK, YUK (106); Tax. 279, 501, 505. Aedes nev adensis - USA; ID, OR, WA (191), NV, UT, WY (117); Map modified after Fllis & Brust (167); Tax. 117, 167, 191, 501.

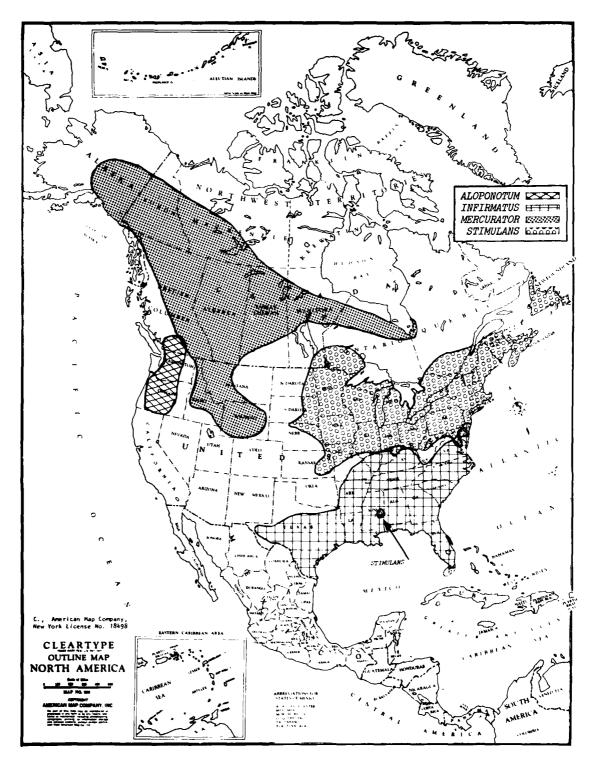


Plate 11. Distribution of Aedes aloponotum - USA: OR, WA (106); CANADA: BC (191); Tax. 191, 279, 505. Aedes infirmatus - USA: AL, AR, FL, GA, KY, LA, MS, MO, NC, SC, TN, TX (106), DE (252), IL (220), IN (217), MD (123), VA (7), Not in AZ (7); Map modified after Arnell (7); Tax. 7, 279. Aedes mercurator - USA: ID, MT, WY (106 as Ae, stimulans), AK (504); CANADA: ALTA, MAN, NWT, ONT, SASK, YUK (504), BC, PQ (505); Tax. 504, 505. Aedes stimulans USA: CT, DE, IL, IA, KS, ME, MA, MI, MN, MS, MO, NE, NH, NJ, NY, OH, PA, RI, SD, VT, WI (106), IN (414), KY (128), MD, VA (46), Not in CO (212) nor UT (338); CANADA: MAN, NB, NS, ONT, PEI, PQ (106), NFLD (360), Not in ALTA, BC, NWT, SASK, YUK (504); Tax. 279, 302, 504, 505.

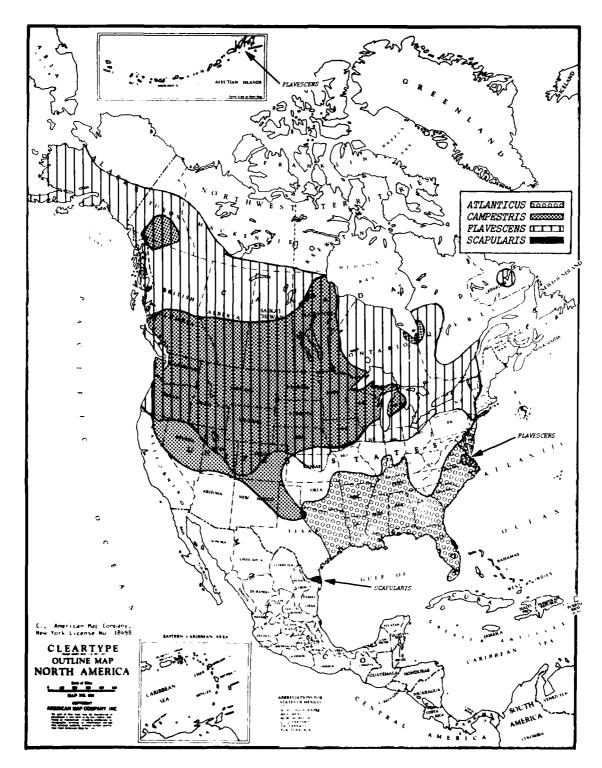


Plate 12. Distribution of Aedes atlanticus - USA: AL, AR, DE, DC, FL, GA, KS, LA, MD, MS, MO, NJ, NY, NC, OK, SC, TX, VA (106), IL (393), IN (422), KY (127), TN (61); Tax. 279. Aedes campestris - USA: CO, ID, IA, MI, MN, MT, NE, ND, OR, SD, TX, UT, WA, WI, WY (106), CA (203), NV (115), NM (460); CANADA: ALTA, BC, MAN, ONT, PQ, SASK, YUK (106); Tax. 279, 478, 505. Aedes flavescens - USA: AK, CA, CO, ID, IL, IA, KS, MI, MN, MO, MT, NE, NY, ND, OR, SD, UT, WA, WI, WY (106), IN (414), NH (Burger, in litt. 1977), NJ (132), OH (Berry & Parsons, in litt. 1978), VA (Skeeter 23: 2, 1978); CANADA: ALTA, BC, LAB, MAN, NWT, ONT, SASK, YUK (106), PQ (155); Tax. 279, 478, 505. Aedes scapularis - USA: TX (106), Not in FL (7) nor LA (95); Tax. 7, 279.

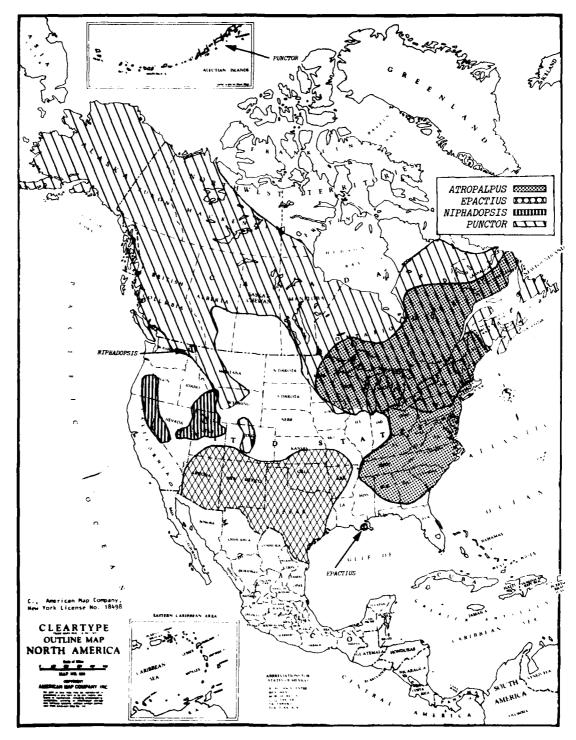


Plate 13. Distribution of Acdes atropalpus - USA; CT, DC, GA, ME, MD, MA, MN, NH, NJ, NY, NC, PA, RL SC, TN, VT, VA, WV, WI (106), AL, KY (514), MI (23), OH (Berry & Parsons, in litt 1978); CANADA; LAB, ONT, PQ (106); Map modified after Zavortink (514); Lax, 279, 347, 348, 505, 514. Acdes epactus USA; AZ, AR, CO, KS, MO, NM, OK, TN, UT (514), LA (144); Map modified after Zavortink (514); Tax, 347, 348, 514. Acdes inphadopsis - USA; ID, NV, OR, UT (106), CA (97), WY (114); CANADA; Not in ALTA (Pucat, in litt, 1979); Tax, 279, 378. Acdes punctor USA; AK, CO, H., ME, MA, MI, MN, MT, NH, NJ, NY, ND, VT, WI, WY (106), ID (190), IN (416), IA (361), PA (69), WA (325), Not in MD (46) nor UT (338); CANADA; ALTA, BC, LAB, MAN, NB, NWT, NS, ONT, PFT, PQ, SASK, YUK (106), NFLD (177); Tax, 279, 505.

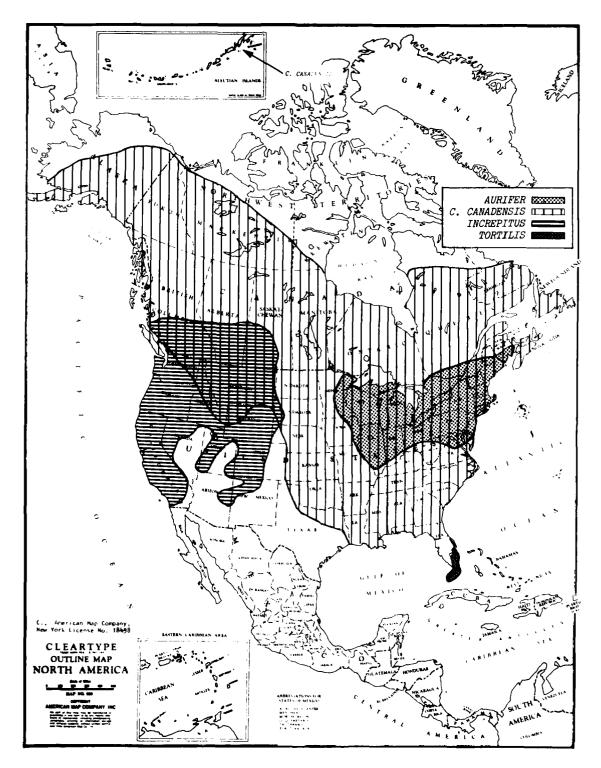


Plate 14. Distribution of Acdes aurifer - USA; CT, DE, H., IA, ME, MD, MA, MI, MN, NH, NJ, NY, OH, RI, VT, WI (106), IN (407), PA (492); CANADA; ONT, PQ (106), NB (314); Tax. 279, 505. Acdes e. canadensis - USA; AL, AR, CT, DE, DC, FL, GA, ID, IL, IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NY, NC, ND, OH, OK, PA, RI, SC, SD, TN, TX, VA, VT, WA, WI, WY (106), AK (130), WV (3), Not in NM (502); CANADA; ALTA, BC, LAB, MAN, NB, NFLD, NWT, NS, ONT, PEI, PQ, SASK, YUK (106); Tax. 28, 279, 505. Acdes increptins - USA; CA, CO, ID, MT, NV, NM, OR, UT, WA, WY (106), AZ (503), NE (376), SD (187); CANADA; BC, SASK (106), ALTA (370); Tax. 279, 501, 505. Acdes torthis - USA; FL (106); Map after Arnell (7); Tax. 7, 28, 34.

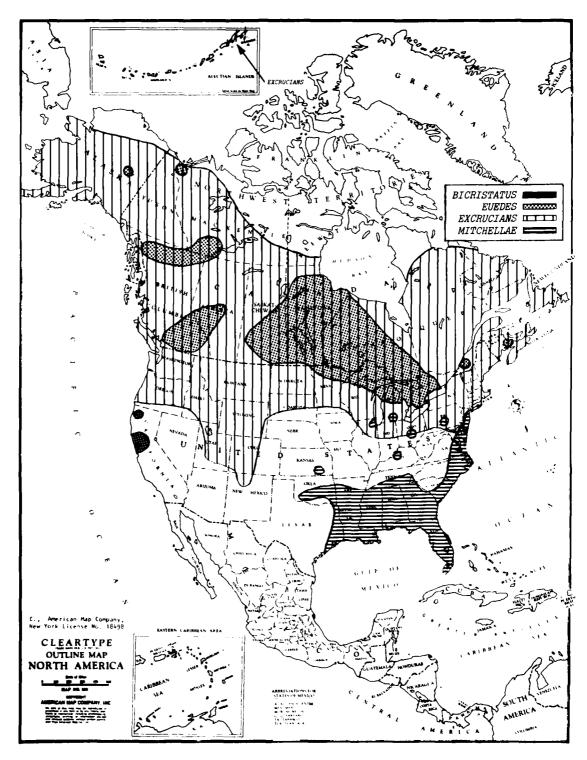


Plate 15. Distribution of Aedes hieristatus - USA: CA (106, 107); Tax. 279. Aedes eucdes - USA: AK (333), MI (23), MN (395); CANADA: ALTA (169), MAN (70), NS (529), ONT (444), PQ (155), BC, NW I, SASK (505); Tax. 138, 279, 395, 504, 505, 516. Aedes exerucians - USA: AK, CO, CT, ID, IL, ME, MA, MI, MN, MT, NH, NJ, NY, ND, OH, OR, PA, RI, UT, VT, WA, WI, WY (106), DE (252), IN (414), MD (50), NM (340); CANADA: ALTA, BC, MAN, NFLD, NWT, NS, ONT, PEI, PQ, SASK, YUK (106), LAB (219), NB (314); Tax. 136, 205, 279, 302, 501, 505. Aedes mitchellae - USA: AL, AR, DE, DC, FL, GA, IL, LA, MD, MS, NJ, NM (no specific locality), NY, NC, OK, SC, TN, TX, VA (106), KS (344), KY (127), MI (Newson, in litt. 1977), OH (352), PA (443); Tax. 279.

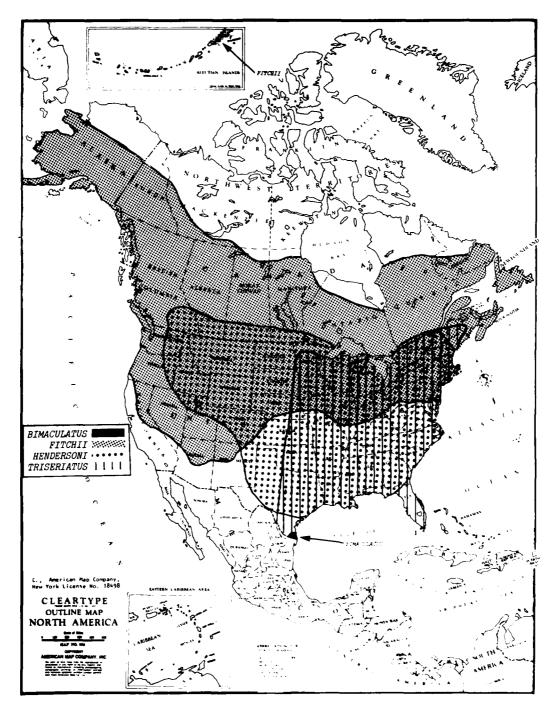


Plate 16. Distribution of Acdes bimaculatus - USA: TX (106). Acdes titchii - USA AK, CA, CO, CT, ID, H., IA, ME, MA, MI, MN, M1, NF, NH, NJ, NY, ND, OH, OR, RL, UT, VT, WA, WI, WY (106), AZ, NV (382), DF (251), IN (414), MD (46), NM (499), PA (396); CANADA: ALTA, BC, LAB, MAN, NFLD, NWT, ON LPFT, PQ, SASK, YUK (106), NB (214), NS (529); Tax. 279, 501, 505. Acdes hendersone - USA: AL, AR, CT, DF, DC, GA, IA, KY, ME, MD, MA, MN, MS, NH, NJ, NY, NC, OR, PA, SC, TN, UT, VA, WV (514), CO, TX (65), ID, WY (337), IL (222), IN, MI, OH (472), KS, NF (212), LA (116), MO (427), MT, NM, SD (332), OK (353), WI (268); CANADA: BC (514), MAN (470), ON T, PQ, SASK (505); Map after Zavortink (514): Tax. 65, 201, 211, 279, 505, 514. Acdes triscriatus - USA: AL, AR, CT, DF, DC, FL, GA, H, TN, TA, KS, KY, LA, MF, MD, MA, MI, MN, MS, MO, NE, NH, NJ, NY, NC, OH, OK, PA, RI, SC, TN, TX, VT, VA, WI (106), WV (3); CANADA: ON T, PQ (106), NB (529); Map modified after Zavortink (514): Tax. 65, 201, 211, 279, 505, 514.

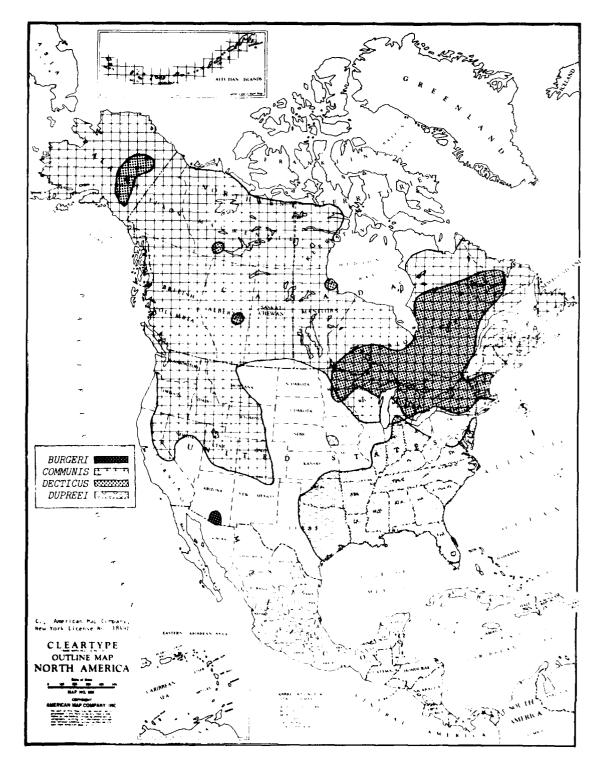


Plate 17. Distribution of Acdes biogeri - USA: AZ (514); Map after Zavortink (514); Lax, 73, 514, Icdes communis - USA: AK, CA, CO, MF, MA, MI, MN, MT, NH, NJ, NY, OR, PA, UT, WA, WI, WY (106), ID (321), NV (109), NM (340); CANADA, ATTA, BC, LAB, MAN, NB, NWT, NS, ONT, PFT, PQ, SASK, YUK (106), NFTD (167); Map modified after Fllis & Brust (167); Tax, 167, 279, 501, 505, Acdes decticus - USA: AK, MA, MI, NH, NY (106), MF (399), MN (19), PA (493); CANADA: LAB, ONT (106), ALTA, MAN (156), PQ (288), NWT, (505); Map modified after Bourassa et al. (55); Tax, 279, 505, Acdes duprice - USA: AL, AR, FL, GA, H., LA, KS, KY, UV, MS, MO, NJ, NC, OK, SC, TN, TX, VA (106); DF (250), IN (416), MI (Newson, in litt, 1978), OH (352), Not in MD (46); Tax, 279.

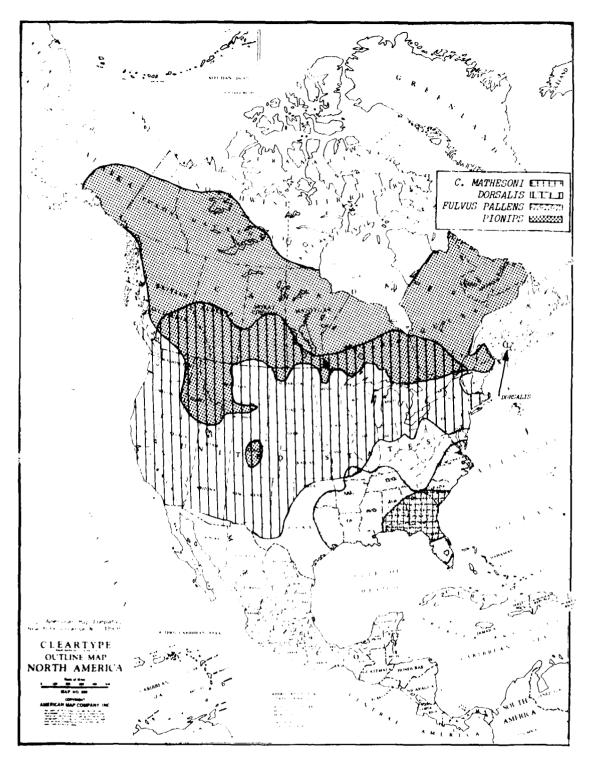


Plate 18. Distributes a of Ledes e. mathesom - USA: AL, FL, GA, SC (106), Not in OH (Parsons & litt, 1978); CANADA: NFLD (360, doubtful 505). Ledes dorsalis - USA: CA, CO, CT, Di , ID, IL, IA, KS, MA, MN, MO, MT, NF, NV, NM, NY, ND, OH, OK, OR, PA, SD, TX, UT, WA, WI, WY (106), AZ (382), IN (416), MD (292), MI (23), NJ (72); CANADA: ALTA, BC, MAN, ON 1, PQ, SASK (106), NB (314); Tax. 13, 53, 205, 279, 505, Ledes futions pallens - USA: AL, AR, FL, GA, HL, KY, LA, MD, MS, NC, OK, SC, TN, TX, VA (106), MO (294); Tax. 279, Ledes promps - USA: AK, CO, ID, MT, ND, WY (106), ME (301), MH (23), MN (16), OR, WA (190); CANADA: ALLA, BC, LAB, MAN, NWT, ON 1, PQ, SASK, YUK (106): Tax. 136, 205, 279, 505.

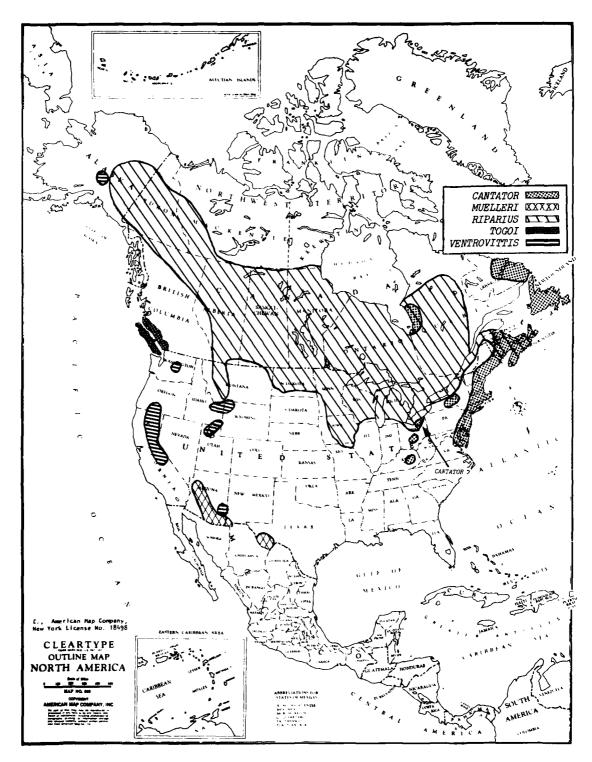


Plate 19. Distribution of Aedes cantator - USA: CT, DE, ME, MD, MA, NH, NJ, NY, PA, RI, VA (106), KY (Knapp, in litt. 1978), OH (352); CANADA: NB, NS, PEI (106), LAB, PQ (291), NFLD (478); Tax. 279, 505. Aedes muelleri - USA: AZ (106), NM (336), TX (64); Map modified after Zavortink (514); Tax. 305, 514. Aedes riparius - USA: AK, IA, MI, MN, MT, NY, ND, WI (106), MO (427), OH (352), Not in CO (212) nor WY (350); CANADA: ALTA, BC, MAN, NWT, ONT, SASK, YUK (106), NB (529), NS (505), PQ (155); Tax. 136, 205, 279, 505. Aedes togoi - CANADA: BC (437); Tax. 505. Aedes ventrovittis - USA: CA, ID, WA (106), AK (48), AZ (304), OR (190), UT (330), WY (331); Tax. 279, 331.

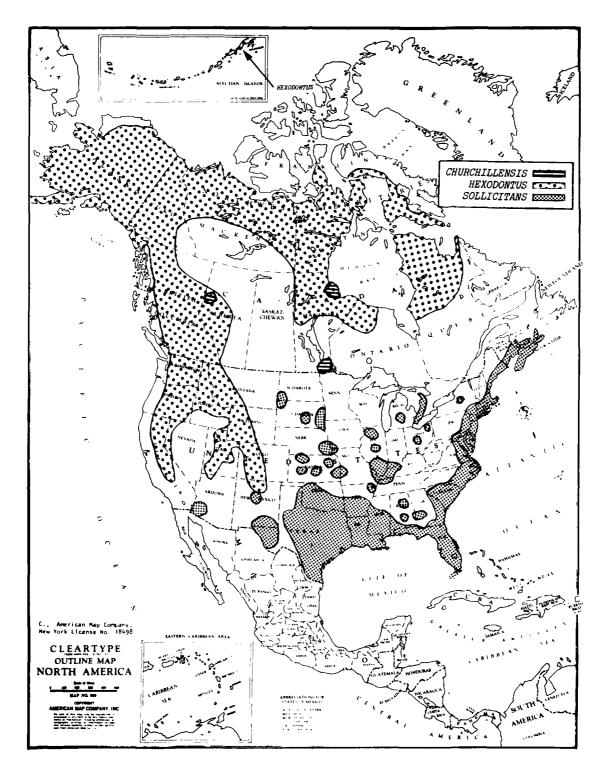


Plate 20. Distribution of Aedes churchillensis - CANADA: ALTA, MAN (167); Tax. 167, 505. Aedes hexodontus - USA: AK, CA, CO, ID, MT, OR, WA (106), NV (109), NM (340), UT (379), WY (350); CANADA: BC, MAN, NWT, PQ, YUK (106), ALTA (370), LAB, ONT (505); Tax. 279, 478, 501, 504, 505. Aedes sollicitans - USA: AL, AZ, AR, CT, DE, DC, FL, GA, H., IN, KS, KY, LA, ME, MD, MA, MS, MO, NE, NH, NJ, NM, NY, NC, ND, OH, OK, PA, RI, SC, TX, VA (106), IA (248), MI (Newson, in litt. 1977), SD (187), TN (438); CANADA: NB, NS, PEI (106); ONT (223); Map modified after Knight (248); Tax. 28, 34, 279, 505.

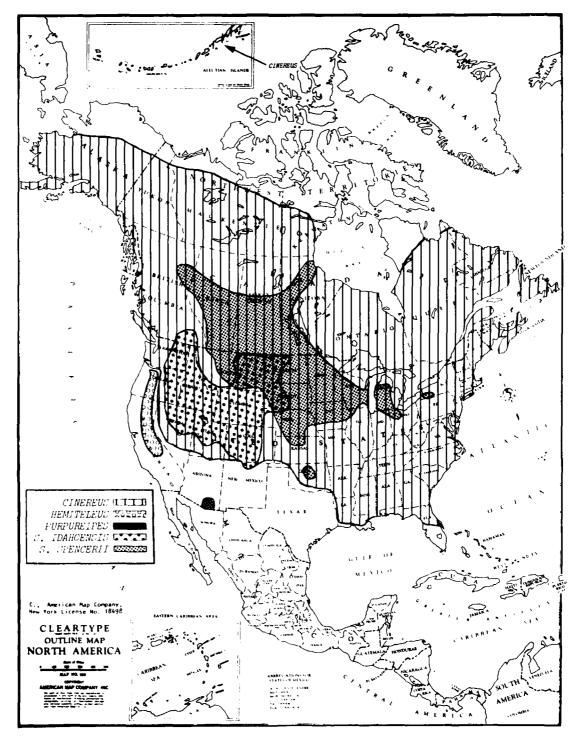


Plate 21. Distribution of Aedes emereus - USA: AL, AK, AR, CO, CT, DE, DC, FL, GA, ID, IL, IN, IA, KS, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NY, NC, ND, OH, OK, OR, PA, RI, SC, SD, TN, UT, VT, WA, WI, WY (106), KY (127), LA (116), NV (382), NM (503), VA (193); CANADA: ALTA, BC, LAB, MAN, NWT, NS, ONT, PEI, PQ, SASK, YUK (106); NB (314), NFLD (505); Tax. 54, 279, 356, 501, 505, Aedes hemiteleus - USA: CA, OR (54); Tax. 54, 356, Aedes purpureipes - USA: AZ (106); Tax. 279, 306, 514, Aedes s. idahoensis - USA: CO, ID, MT, NE, NV, ND, OR, UT, WA, WY (106), NM (212), SD (187); CANADA: BC (106); Tax. 338, 366, Aedes s. spencerii - USA: IL, TA, KS, MI, MN, MT, NF, NY, ND, SD, WI, WY (106), OH (95), OK (353); CANADA: ALTA, BC, MAN, SASK (106), ONT (505); Tax. 28, 279, 338, 366, 505.

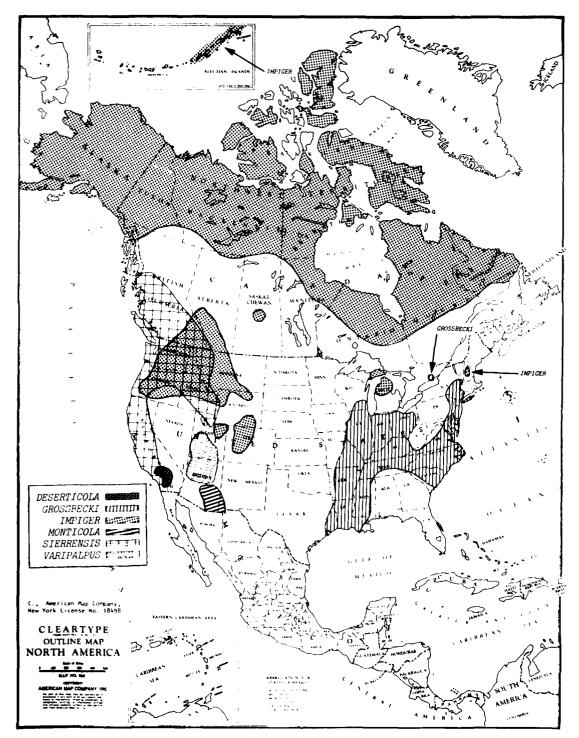


Plate 22. Distribution of Acdes descrite ola - USA; CA (510); Map after Arnell & Nielsen (9); Tax, 9, 510. Acdes grossbeekt - USA; AR, DF, H., KY, LA, MD, MS, MO, NJ, NY, OH, SC, TN, VT, VA (106), CT (483), IN (407), PA (493), TX (Harris Co. M.C.D., in litt. 1978), WI (466); CANADA; ONT (223); Tax. 279, 505. Acdes impiger - USA; AK, CO, ID, MT, OR, UT, WA, WY (106), MI (229), NH (235); CANADA; ALTA, MAN, NWT, ONT, PQ, SASK, YUK (106), BC (177), LAB (505); Tax. 28, 139, 279, 478, 505. Acdes monticola - USA; AZ (38), NM (336); Map after Arnell & Nielsen (9); Tax. 9, 38, 279. Acdes sierrensis - USA; CA (267), ID (80) MT (337), NV (112), OR (213), UT (332), WA (325); CANADA; BC (135); Map after Arnell & Nielsen (9); Tax. 9, 36, 38, 134, 279, 505. Acdes varipalfus - USA; AZ (38), UT (332); Map after Arnell & Nielsen (9); Tax. 9, 36, 38, 279.

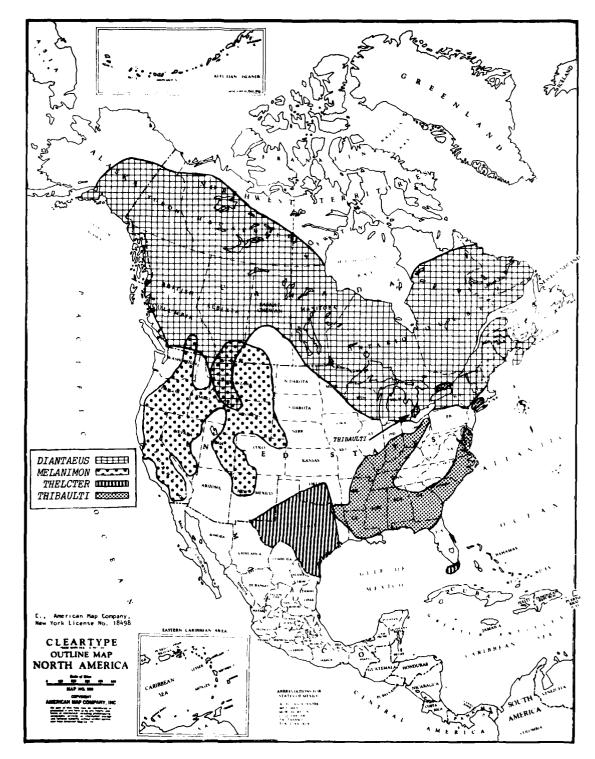


Plate 23. Distribution of Aedes diantaeus - USA; AK, ME, MA, MI, MN, MT, NH, NY, VT, WY (106), PA (493), WI (420); CANADA; BC, LAB, NWT, NS, ONT, PQ, YUK (106), ALTA (370), SASK (380), MAN (505); Tax. 205, 279, 505, Aedes melanime u USA; CA, CO, MT, NV (106), ID, NE, NM, UT, WA, WY (381), OR (190); CANADA; ALTA (76), BC (Belton, in litt. 1978), SASK (226); Tax. 13, 53, 279, 381, 505, Aedes theleter - USA; FL, OK, TX (106), NM (316); Map after Arnell (7); Tax. 7, 279, Aedes thibaulti - USA; AL, AR, FL, GA, IL, KY, LA, MS, MO, NC, OH, SC, TN, TX (106), CT, NY (485), DE (46), IN (408), MD (239), VA (46); CANADA; ONT (44); Tax. 279, 505.

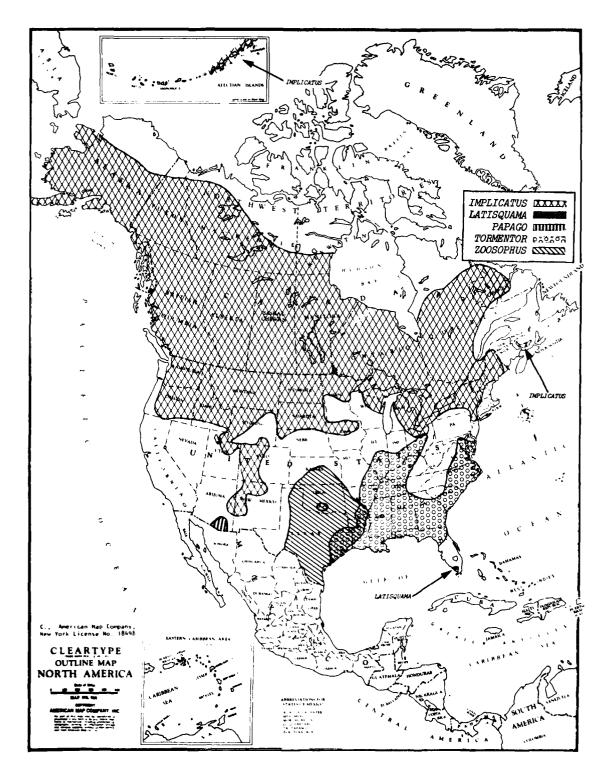


Plate 24. Distribution of Aedes implicatus - USA: AK, CO, ID, IA, MA, MI, MN, MT, NE, NH, NY, UT, WA, WY (106), AZ, NM (340), ME (301), NJ (72), OH (352), OR (190), WI (420); CANADA: ALTA, BC, MAN, NWT, ONT, PQ, SASK, YUK (106), LAB (505), PEI (529); Tax. 279, 478, 501, 505, Aedes papago - USA: AZ (513), Map after Zavortink (514); Tax. 513, 514, Aedes tormentor - USA: AL, AR, FL, GA, LA, MS, MO, NC, OH, OK, SC, TX (106), DE (Lake, in litt. 1972), HL (393), KY (397), MD (50), TN (61); Tax. 279, 388, Aedes zoosophus - USA: KS, OK, TX (106), AR (225), LA (236), Not in NM (502); Map after Zavortink (514); Tax. 514, Culex latisquama - USA: FL (454); Tax. 175.

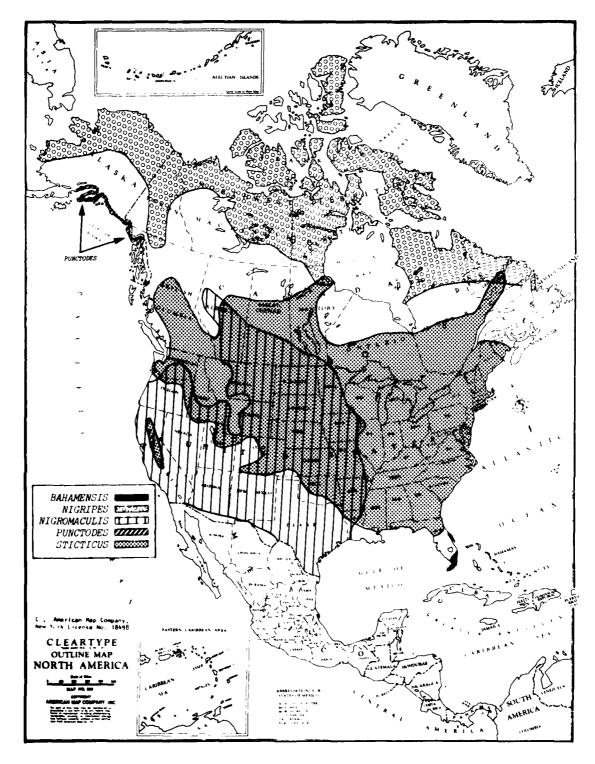


Plate 25. Distribution of Acdes nigropes - USA: AK (106, 463), CANADA: MAN, NWT, PQ, YUK (106), BC, LAB, NFLD (505); Tax. 139, 205, 279, 505. Acdes nigromaculis - USA: CA, CO, 1D, H., IA, KS, MN, MO, MT, NE, NM, ND, OK, OR, SD, TX, UT, WA, WY (106), AZ (382) AR, LA (225), NV (115); CANADA: AL IA, MAN, SASK (106); Tax. 279, 505. Acdes proceedes - USA: AK (106); Tax. 136, 279. Acdes staticus - USA: AL, AR, CA, CO, CT, DE, DC, FL, GA, ID, H., IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NY, NC, ND, OH, OK, OR, PA, SC, SD, TN, TX, UT, VI, VA, WA, WY (106), WV (3), WI (519); CANADA: ALTA, BC, MAN, NB, ONT, PQ, SASK (106), LAB (505); Tax. 205, 279, 505, 515. Culex bahamensis - USA: FL (106); Tax. 34, 57.

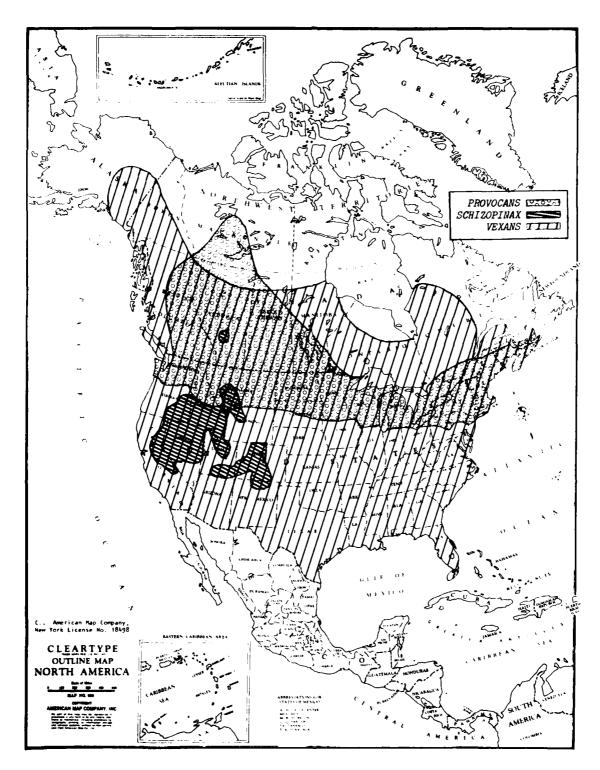


Plate 26. Distribution of Aedes provocans - USA; CT, ID, ME, MA, MI, MN, MT, NH, NY, RI, VT, WA, WI, (106, as Ae, trichurus), NJ (133), PA (398); CANADA; ALTA, BC, MAN, NB, NS, ONT, PEI, PQ, SASK (106, as Ae, trichurus), NWT (505); Tax, 279, 504, 505, Aedes schizopinax - USA; MT, WY (106), CA (382), CO, ID, OR (329), NV (109), NM (340), UT (379); CANADA; ALTA (169); Tax, 279, 501, 505, Aedes vexans - USA; AL, AZ, AR, CA, CO, CT, DE, DC, FL, GA, ID, IL, IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NM, NY, NC, ND, OH, OK, OR, PA, RI, SC, SD, TN, TX, UT, VI, VA, WA, WV, WI, WY (106), AK (441), NV (382); CANADA; ALTA, BC, MAN, NB, NS, ONT, PEI, PQ, SASK, YUK (106); Tax, 279, 505.

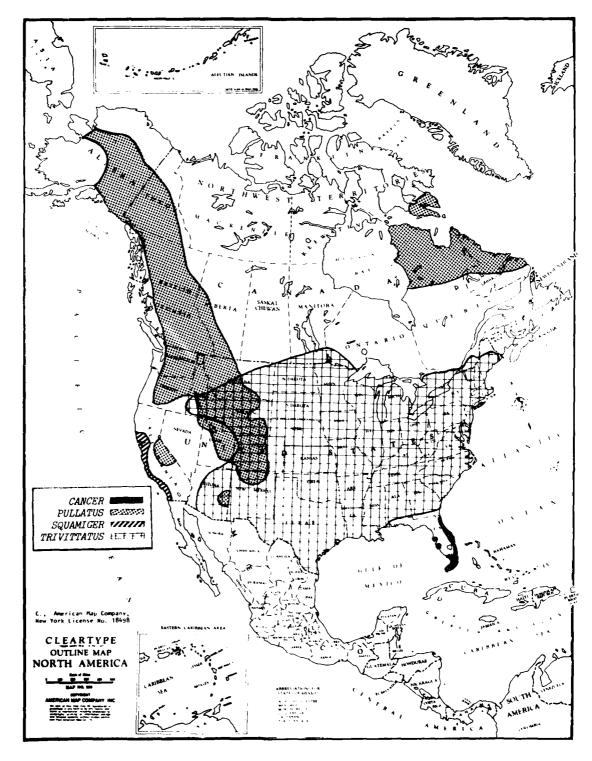


Plate 27. Distribution of Aedes pullatus - USA; AK, CA, CO, ID, MT, OR, UT, WA, WY (106), AZ (340), NV (109), NM (316), Not in MI (114); CANADA; ALTA, BC, NWT, PQ, YUK (106), LAB, NFLD (505); Tax. 136, 205, 279, 501, 505, Aedes squamiger - USA; CA (106); Tax. 279, Aedes trivittatus - USA; AR, CO, CT, DF, DC, GA, ID, H., IN, IA, KS, KY, LA, ME, MD, MA, MN, MO, MT, NE, NJ, NM, NY, NC, ND, OH, OK, PA, RI, SC, SD, TN, TX, VA, WV, WI, WY, (106), AL (61), AZ (382), MI (481), NH (52), UT (330); CANADA; ON F (106), MAN (470), PQ (265); Map after Arnell (7); Tax. 7, 279, 505, Democerites cancer - USA; FL (106); Tax. 1, 34, 35.

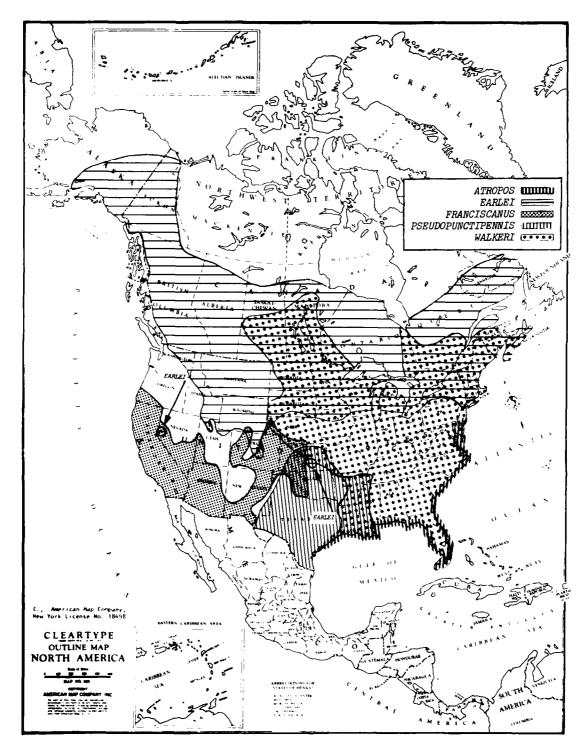


Plate 28. Distribution of Anopheles atropos - USA; AL, FL, GA, LA, MD, MS, NJ, NC, SC, TX, VA (106); Lax, 34. Anopheles earlei - USA; AK, CO, CT, ID, IA, ME, MA, MI, MN, MT, NE, NH, NY, ND, SD, VT, WI, WY (106), KS (344), NV (109), NJ (131), UT (339), WA (190); CANADA; AL, IA, BC, LAB, MAN, NB, NS, ONT, PQ, SASK (106), NWT, PEL, YUK (505); Lax, 488, 505. Anopheles franciscanus - USA; AZ, CA, CO, KS, NV, NM, OK, OR, IN, UT, WY (106), NF (375); Tax, 436, Anopheles pseudopunctipennis - USA; AR, KS, LA, MS, MO, NM, OK, TN, TX (106), Not in CO (242); Tax, 436, Anopheles walkeri - USA; AL, AR, CT, DF, DC, FL, GA, IL, IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, NE, NH, NJ, NY, NC, ND, OH, PA, RI, SC, SD, TN, TX, VT, VA, WI (106); ANADA; MAN, NB, NS, ONT, PQ (106), SASK (308), Not in BC (441), Tax, 28, 505.

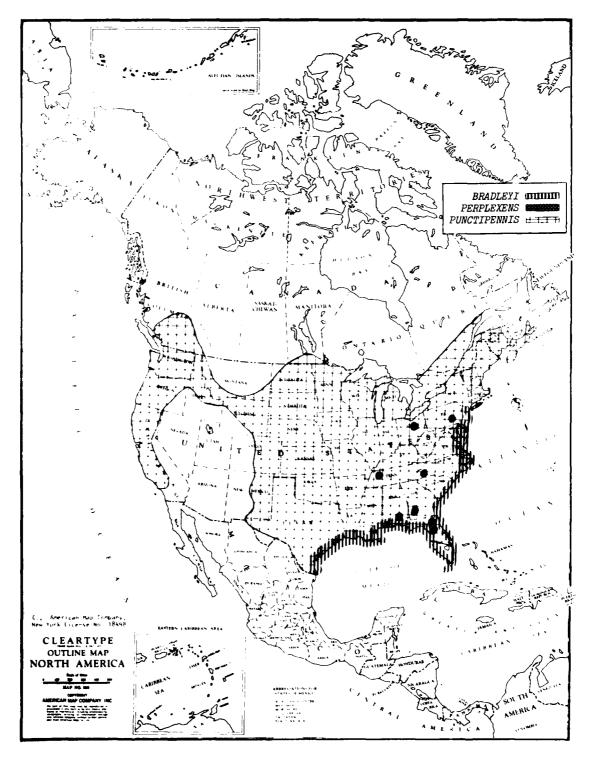


Plate 29. Distribution of Anopheles bradleyi - USA: AL, DE, FL, GA, LA, MD, MS, NJ, NY, NC, SC, TX, VA (106); Map after Floore et. al. (174); Tax. 174. Anopheles perplexens - USA: AL, FL (245), GA (41), NC, TN (394), OH (352), PA (272); Tax. 41. Anopheles punctipennis - USA: AL, AR, CA, CO, CT, DE, DC, FL, GA, ID, IL, IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NM, NY, NC, ND, OH, OK, OR, PA, RI, SC, SD, TN, TX, VT, VA, WA, WV, WI, WY (106); CANADA: BC, MAN, NS, ON L, PQ (106), NB (473); Tax. 39, 41, 505.



Plate 30. Distribution of Inopheles albimanus - USA: FL, TX (106); Tax. 34. Anopheles barberi - USA: AL, AR, DF, DC, FL, GA, HL, IN, IA, KS, KY, LA, MD, MS, MO, NE, NJ, NY, NC, OH, OK, PA, SC, TN, TX, VA (106), MI (Newson, in litt. 1977), MN (369), SD (161), WV (3), WI (363); CANADA: ON I (435), PQ (264); Map modified after Zavortink (511): Tax. 505, 509, 511. Anopheles georgianus - USA: AL, FL, GA, LA, MS, NC, SC (106); Map after Floore et, al. (174); Tax. 174. Inopheles pulithae - USA: AZ, NM (509), TX (511); Map after Zavortink (511): Tax. 509, 511. Anopheles occidentalis: USA: CA, OR, WA (106), Not in AK (192); CANADA: Not in BC (505) nor YUK (488); Tax. 488.

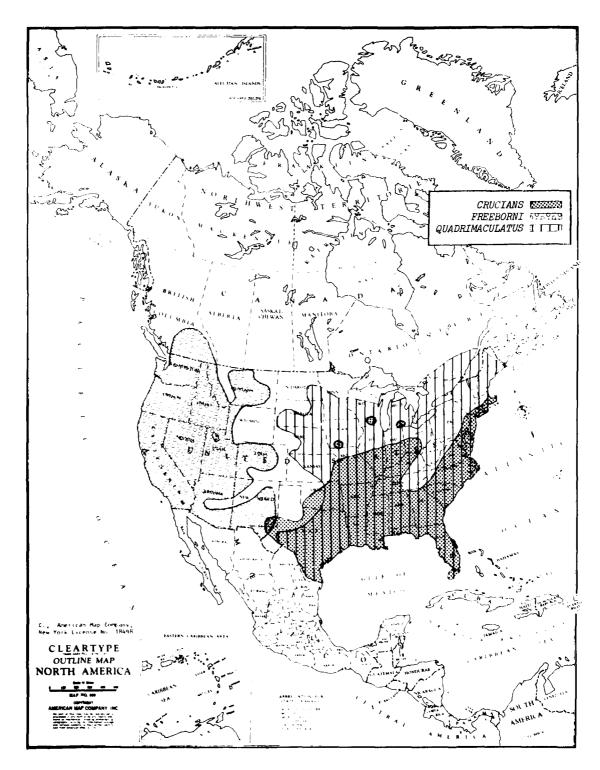


Plate 31, Distribution of Anopheles crucians - USA; AL, AR, CT, DE, DC, FL, GA, H., IN, IA, KS, KY, LA, MD, MA, MS, MO, NJ, NM, NY, NC, OH, OK, PA, RI, SC, TN, TX, VA (106), MI (Newson, in litt, 1977), WI (Dicke, in litt, 1979); Map after Floore et. al. (174); Tax. 34, 174, Anopheles freehorm - USA; AZ, CA, CO, ID, MT, NV, NM, OR, TX, UT, WA, WY (106); CANADA; BC (106); Tax. 488, 505, Anopheles quadrimaculatus - USA; AL, AR, CT, DF, DC, FL, GA, H., IN, TA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, NE, NH, NJ, NY, NC, ND, OH, OK, PA, RI, SC, SD, TN, TX, VT, VA, WI (106), WV (3); CANADA; ON T, PQ (106); Tax. 39, 505.

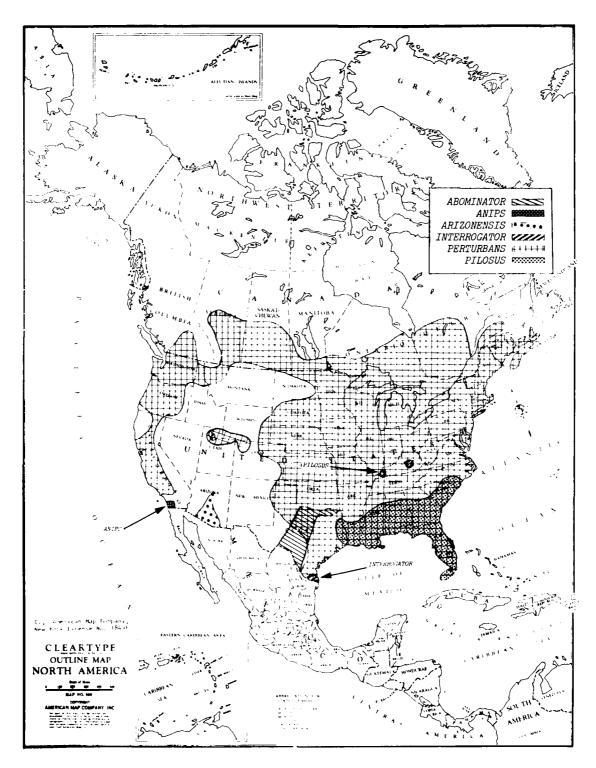


Plate 32. Distribution of Copollettidia perturbans - USA; AL, AR, CA, CO, CT, DE, DC, FL, GA, ID, IL, IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NY, NC, ND, OH, OK, OR, PA, RI, SC, SD, TN, TX, UT, VA, VT, WA, WI, WY (106), NM (502), WV (3); CANADA; BC, MAN, NS, ON F, PEI, PQ, SASK (106), ALTA (206), NB (314); Tax. 39, 389, 390, 505, Culex abominator - USA; TX (106), Not in LA (95); Map after Fournier & Suydev (176); Tax. 175, Culex amps - USA; CA (106); Tax. 54, 175, Culex arranensis - USA; AZ (106); Tax. 262, Culex interrogator - USA; TX (106); Tax. 57, Culex pilosus - USA; AL, 11 GA, KY, LA, MS, NC, SC (106), TX (498); Tax. 34, 175, 247.



Plate 33. Distribution of Culex apicalis -USA; AZ, CA (106), NV (115), NM (172), OK (353), OR (190), TX (62), UT (339); Tax, 261, 262, Culex review - USA; CA (106); Tax, 262, Culex territans - USA; AK, CA, FL, GA, ID, IA, LA, MD, MA, MI, MN, MS, MO, MT, NY, NC, OH, OK, OR, RI, TX, VT, VA, WA (106), AL, SC (245), AZ (382), AR (225), CO (12), CT (483), DE (144), II, (392), TN (414), KS (522), KY (127), ME (301), NE (164), NV (337), NH (52), NJ (71), PA (496), SD (187), TN (438), UT (339), WV (3), WI (147), WY (350); CANADA; BC (406), AL TA (370), LAB, NWT (177), MAN (310), NB (314), NS (473), ON I (242), PQ (155), SASK (380), YUK (505); Tax, 28, 39, 205, 262, 505, Psorophora polinstonn - USA; FL (106), Tax, 34.

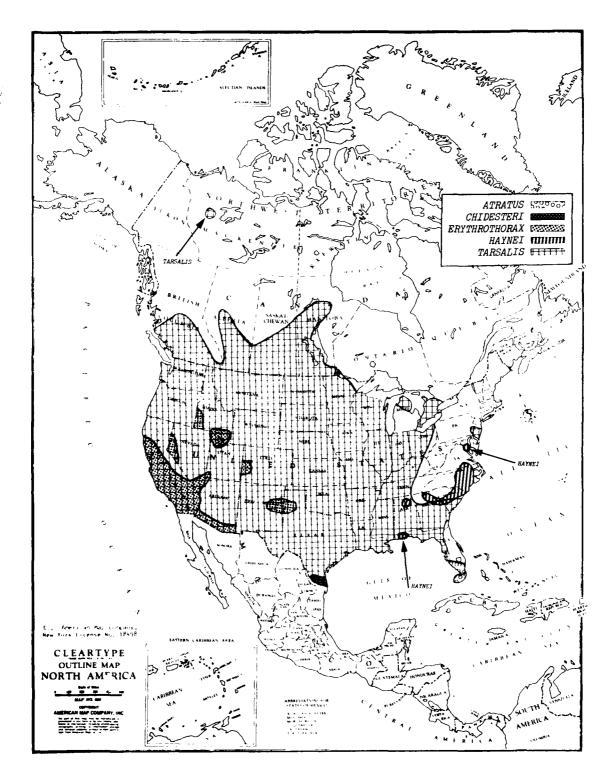
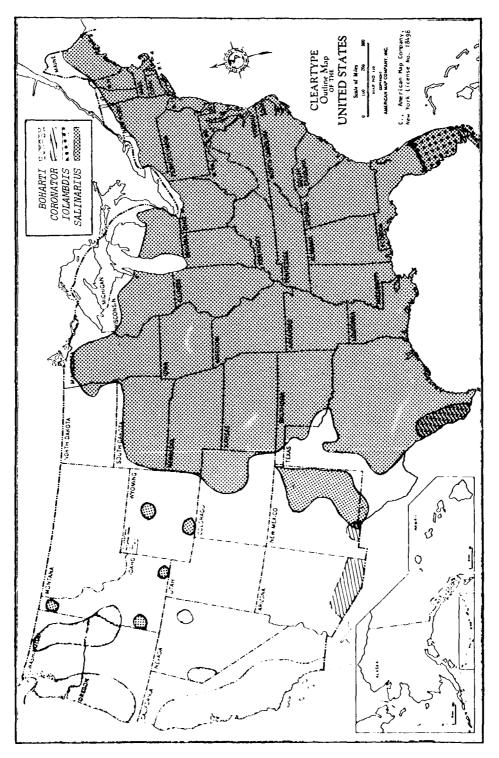


Plate 34, Distribution of Culex atratus - USA; FL (106); Tax. 28, 34, 175, 247, Culex chidesteri - USA; TX (106); Tax. 34, 57, Culex erythrothorax - USA; CA, ID, UT (106), AZ (382), CO (212) NV (115), NM (460), TX (312); Tax. 57, Culex tarsalis - USA; AL, AZ, AR, CA, CO, FL, GA, ID, H., IN, IA, KS, KY, LA, MI, MN, MS, MO, MT, NE, NV, NM, ND, OK, OR, SC, SD, TN, TX, UT, WA, WI, WY (106), NJ (256), OH (352), PA (66); CANADA; ALTA, BC, MAN, NWT, SASK (106), ONT (223); TAX. 54, 57, 505, Wycomym hayner - USA; AL, NC, SC (106), FL (58), GA (145), MD (51), VA (129); Tax. 145.



salimarius - USA: AL, AR, CO, CT, DE, DC, FL, GA, ID, H., IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MQ, NE, NH, NJ, NM, NY, NC, ND, OH, OK, PA, RI, SC, SD, TN, TX, VT, VA, WI, WY (106), WV, (3). Not in UT (330); CANADA: Not in NS (505); Tax. 57. (106), AZ (382), NM 3sK —?(. Not in LA (95); Tax. 57, Culex iolambdis - USA; FL (106); Tax, 34, 175, 247, Culex Plate 35. Distribution of *Culex boliatii* - USA:CA (106), ID,OR, WA (262), NV (382); Tax, 262, Culex caronator - USA; TX

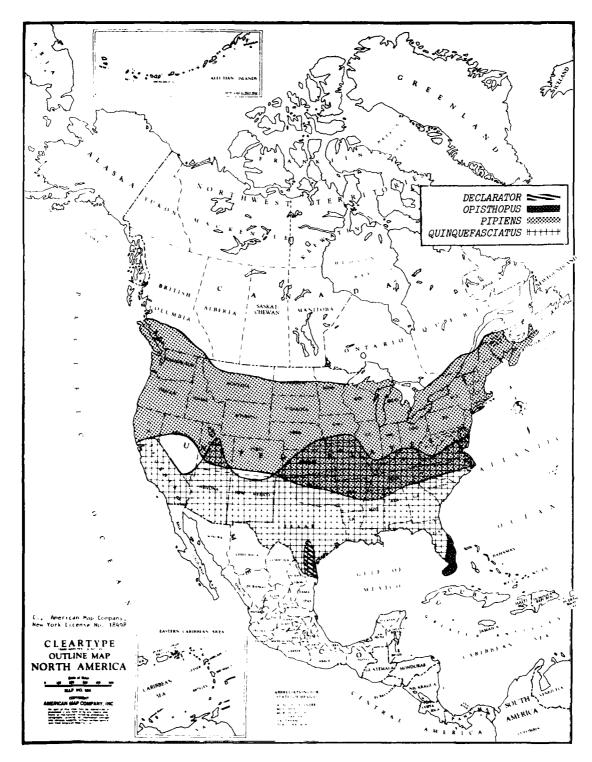
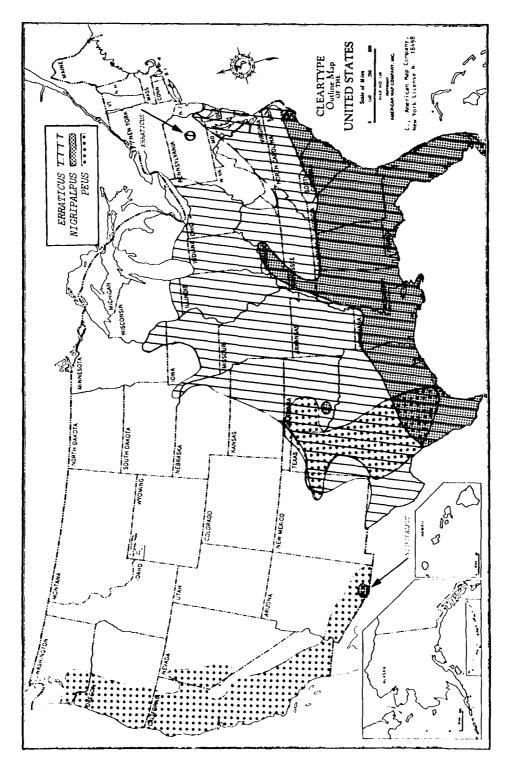


Plate 36. Distribution of Culex declarator - USA: TX (106); Tax. 57, 447. Culex pipiens - USA: AL, AR, CA, CO, CT, DE, DC, GA, ID, IL, IN, IA, KS, KY, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NY, NC, ND, OH, OK, OR, PA, RI, SC, SD, TN, UT, VT, VA, WA, WI, WY (106), WV (3), Not in NM (502); CANADA: BC, NB, NS, ONT, PQ (106); Tax. 14, 57, 505, 523. Culex opisthopus - USA: FL (106); Tax. 34, 175, 185, 247, 457. Culex quanquefasciatus - USA: AL, AZ, AR, CA, DC, FL, GA, IL, IA, KS, KY, LA, MS, MO, NE, NM, NC, OH, OK, SC, TN, TX, UT, VA (106), IN (327), MD (14), NV (118), WV (3); Iax. 14, 31, 34, 405, 523, 524.



(225), Not in AR, NM (95); Tax, 34, 57, Culty pens (USA; CA, OK, OR, TX, WA (106), AZ, NV (382), NM (460); Tax, 34, 57. Plate 37. Distribution of *Colles contiens* - USA: AL, AR, DE, DC, FL, GA, H., IN, IA, KS, KY, LA, MD, MI, MS, MO, NE, NC, OH, OK, SC, SD, TN, TX, VA (106), MN (16), NJ (132), NM (502), PA (396), WV (3), WI (Dicke, in litt, 1979); Tax. 34, 175, 247, Culex mgripalpus - USA; AL, FL, GA, LA, MS, NC, SC, TN, TN (106), AZ (304), KY (127), OK

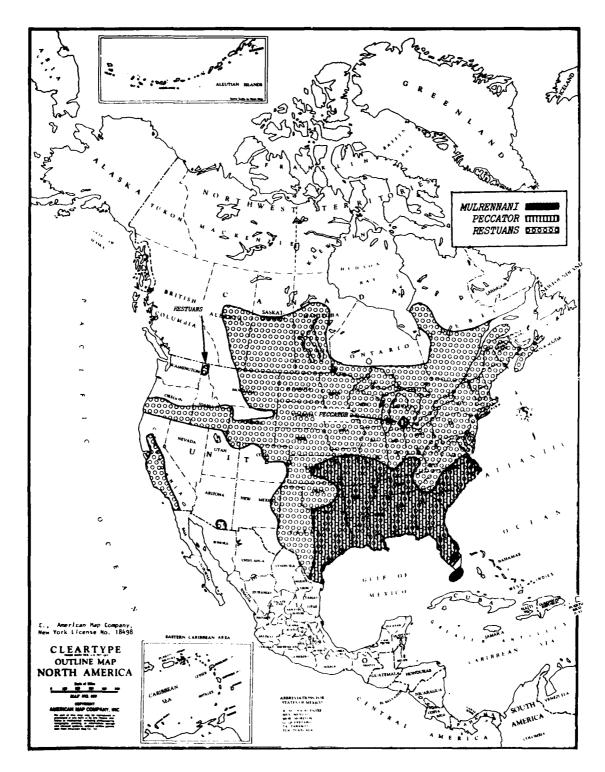


Plate 38, Distribution of Culex mulrennani - USA; FL (106); Tax. 175, 247, Culex peccator - USA; AL, AR, FL, GA, H., KS, KY, LA, MI, MS, MO, NC, OK, SC, TN, TX, VA (106), Not in DE (Lake, in litt. 1972); Tax. 175, 247, Culex restuans - USA; AL, AR, CA, CO, CT, DE, DC, FL, GA, ID, H., IN, IA, KS, KY, LA, ME, MD, MA, MI, MN, MS, MO, MT, NE, NH, NJ, NM, NY, NC, ND, OH, OK, PA, RI, SC, SD, TN, TX, UT, VT, VA, WV, WI, WY (106), AZ (382), OR (213); CANADA; MAN, NB, ONT, PQ, SASK (106), ALTA (371), NS (529); Tax. 57, 505.

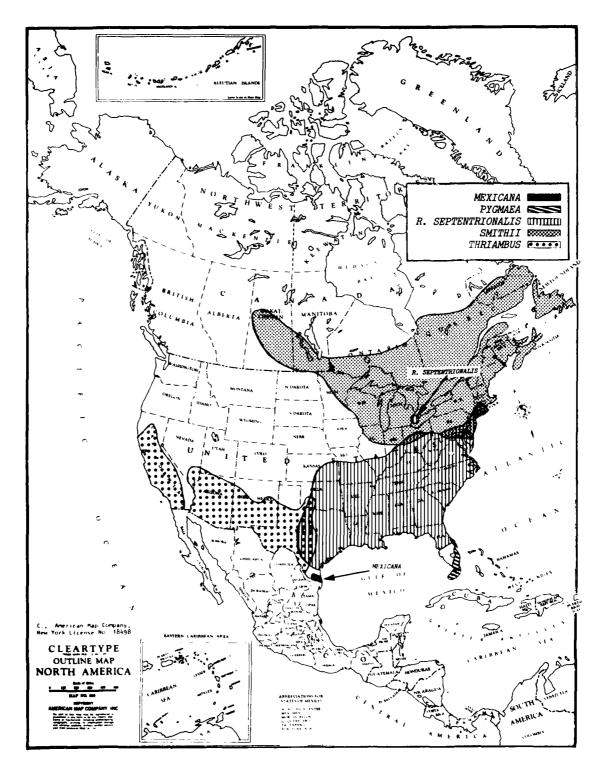


Plate 39. Distribution of Culey thriambus - USA: CA, OK, TX (106), AZ (385), NV (382), NM (225), UT (334); Tax. 57. Psorophora mexicana - USA: TX (241); Tax. 28. Psorophora pygmaca - USA: FL (106), Not in MS (95); Tax. 28. 34. Toxorhynchites r. septentrionalis - USA: AL, AR, DE, DC, FL, GA, IL, KS, KY, LA, MD, MS, MO, NJ, NC, OH, OK, PA, SC, TN, TX, VA, WV (106), CT (286), IN (218), NY (246); CANADA: ONT (351); Tax. 151, 476, 505. Wycomyra smithii - USA: CT, DE, H., ME, MA, MI, MN, NH, NJ, NY, OH, RI, WI (106), IN (411), MD (50), PA (492); CANADA: LAB, MAN, NS, ONT (106), NB (529), NFLD (360), PEI (505), PQ (288), SASK (77); Tax. 145, 505.

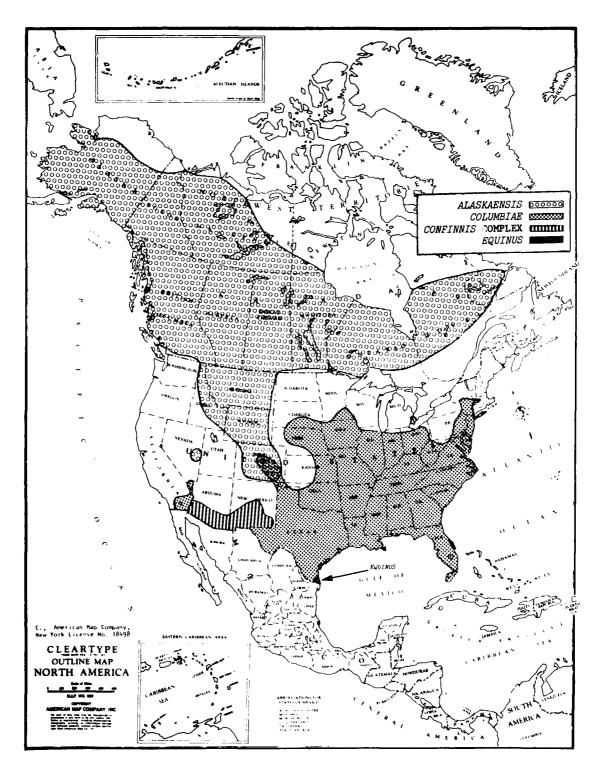


Plate 40. Distribution of Culiseta alaskaensis - USA; AK, CO, MT, WY (106), ID (146), NV (535); CANADA; ALTA, BC, LAB, MAN, NWT, PQ, YUK (106), SASK (380); Map modified after Hopla (227); Tax. 295, 505. Haemagogus equinus - USA; TX (469); Tax. 6, 28, 34. Psorophora columbiae - USA; AL, AR, CO, DE, DC, FL, GA, H., IN, IA, KS, KY, LA, MD, MA, MS, MO, NE, NJ, NM, NY, NC, OH, OK, PA, SC, SD, TN, TX, VA, WV (106, as Ps. confinnis), CA (54), MN (124), NV (112); CANADA; ONT (505); Tax. 34, 49, 54, 142, 505. Psorophora confinnis complex - USA; AZ, NM (106), CA (doubtful, 54); Tax. 34, 49, 54, 142.

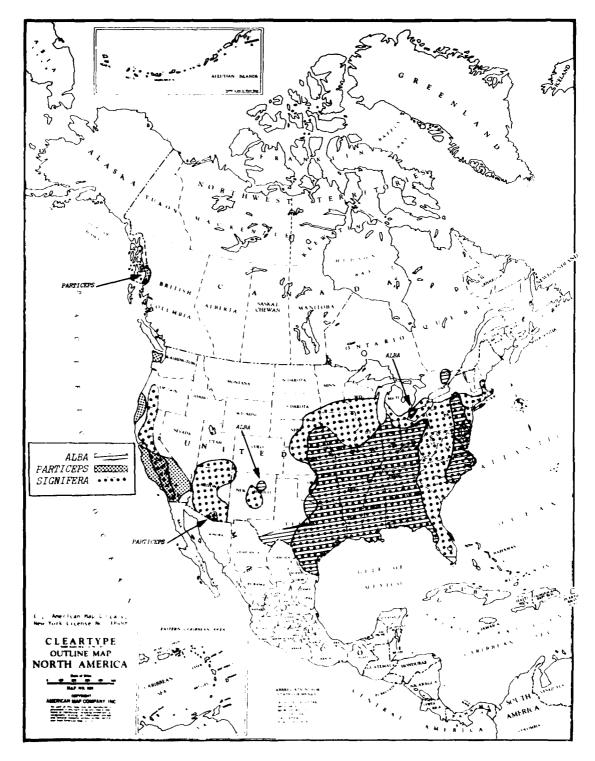


Plate 41. Distribution of Culiscia particeps - USA: CA, OR (106), AK (48), AZ (382), WA (325); Tax. 296, 448. Orthopodomyia alba - USA: AL, IL, KY, LA, MS, MO, NJ, NY, NC, TX, VA (106), AR, DC, MD, OH (508), DE (251), FL (Haeger in litt. 1967), GA (461), IN (67), IA (280), KS (525), MI (200), NM (315), OK (225), PA (495), NF (278), IN (60); CANADA: ON I (435), PQ (505); Tax. 505, 508. Orthopodomyia significa - USA: AL, AR, C1, DE, DC, FL, GA, IL, IN, IA, KS, KY, LA, MD, MA, MS, MO, NF, NJ, NM, NY, NC, OH, OK, PA, RI, SC, FN, TX, VA (106), AZ (382), CA, OR, UT (508), MI (200), MN (369), NH (Burger, in litt. 1977), SD (161), WV (3), WI (363); CANADA: ONT (435); Tax. 505, 508.

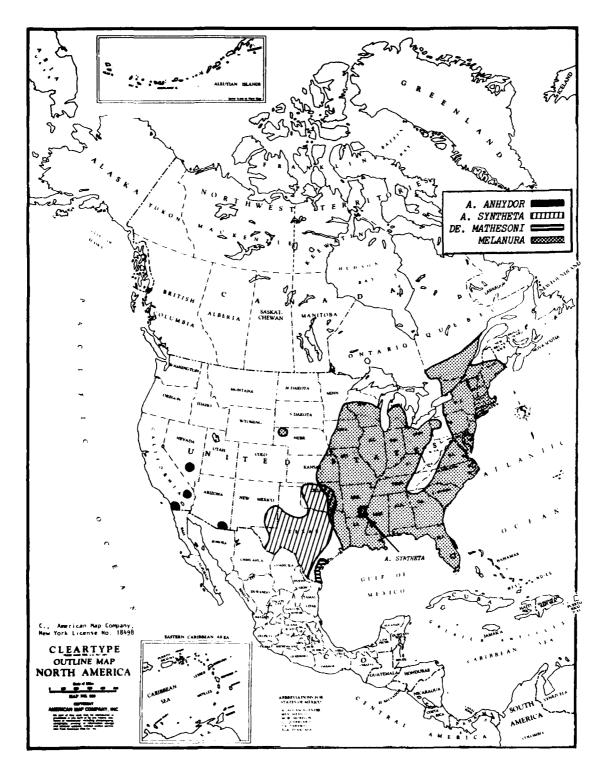


Plate 42. Distribution of Culrseta melanura - USA; AL, AR, DE, DC, FL, GA, IA, KY, LA, ME, MD, MA, MI, MN, MS, MO, NE, NH, NJ, NY, NC, OH, OK, PA, RI, SC, TN, TX, VA, WI (106), CI (482), II. (413), IN (406), KS (344), Not in CO (212); CANADA; ONT (108), PQ (168, 189); Tax. 505. Democerates mathesom - USA; TX (35); Tax. 1, 35, 359. Uranotaenia a. anhydor - USA; CA (106), AZ, NV (37); Tax. 37. Uranotaenia a. syntheta - USA; NM, OK, TX (106), AR (59); Tax. 37, 526.

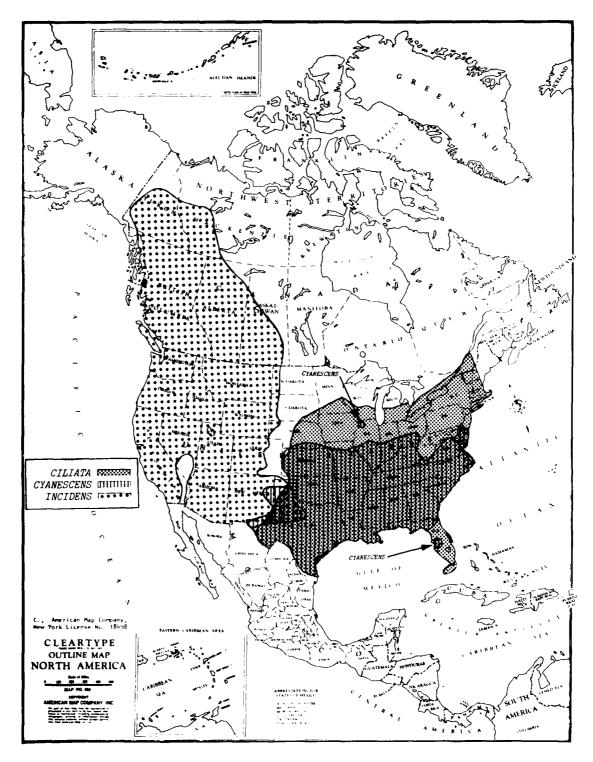


Plate 43. Distribution of Culiscia medens - USA: AK, AZ, CA, CO, ID, MT, NE, NV, NM, ND, OK, OR, TX, UT, WA, WY (106), SD (187); CANADA: ALTA, BC, NWT, YUK (106), SASK (310); Not in NS (177); Tax. 28, 296, 505, Psorophora ciliata - USA: AL, AR, CT, DE, DC, FU, GA, H., IN, IA, KS, KY, LA, MD, MA, MI, MS, MO, NE, NH, NJ, NY, NC, OH, OK, PA, RI, SC, SD, TN, TX, VA, WV, WI (106), MN (16), NM (316); CANADA: ON T, PQ (106); Tax. 28, 34, 39, 505, Psorophora cyanescens - USA: AL, AR, FL, GA, IL, IN, KS, KY, LA, MS, MO, NE, NM, NC, OH, OK, SC, TN, TX, VA (106), DE (254), MD (240), NJ (132).

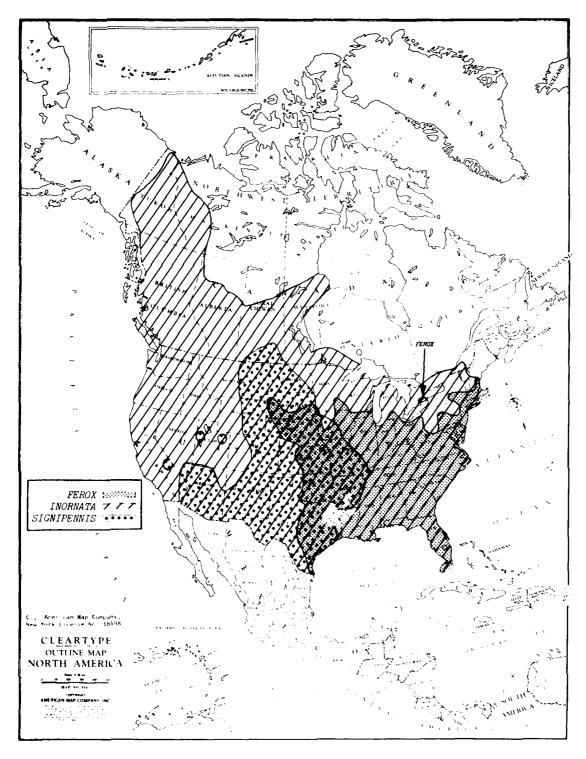
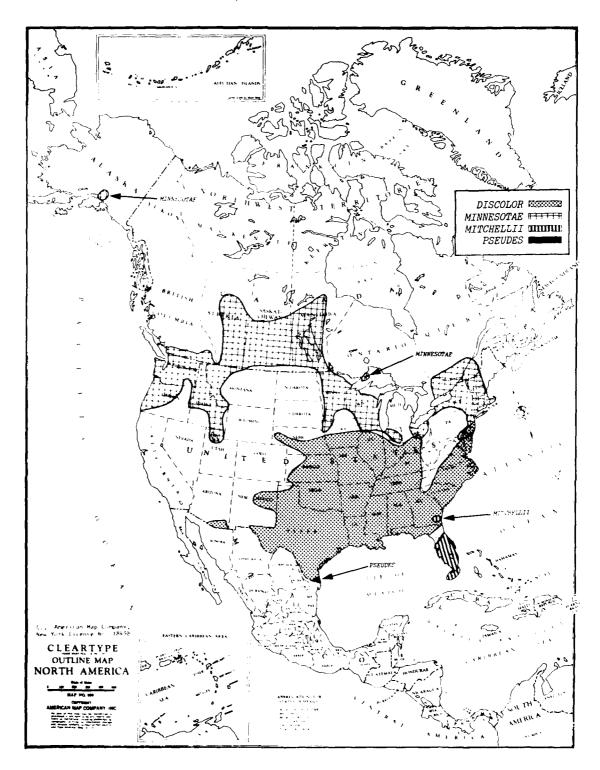
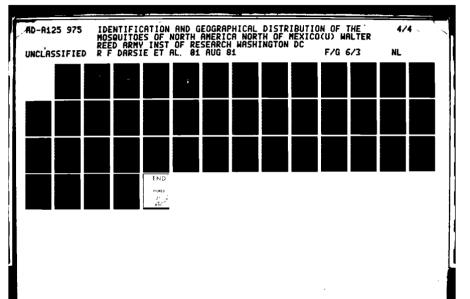
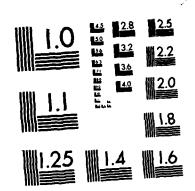


Plate 44. Distribution of Culiscia informata - USA: AL, AZ, AR, CA, CO, DE, DC, FL, GA, ID, IL, IN, IA, KS, KY, LA, MD, MA, MI, MN, MS, MO, MT, NE, NV, NH, NJ, NM, NY, NC, ND, OH, OK, OR, PA, SC, SD, TN, TX, UT, VA, WA, WI, WY (106), CT (483), WV (3); CANADA: ALTA, BC, MAN, NWT, ONT, SASK, YUK (106), PQ (265); Tax, 296, 505, Psorophora ferox - USA: AL, AR, CT, DE, DC, FL, GA, IL, IN, IA, KS, KY, LA, MA, MI, MN, MS, MO, NE, NH, NJ, NY, NC, OH, OK, PA, SC, SD, TN, TX, VA, WI (106), MD (46), WV (3); CANADA: ONT (505); Tax, 28, 34, 505, Psorophora significants - USA: AZ, AR, CO, IA, KS, KY, MO, MT, NE, NM, ND, OK, SD, TN, TX, WY (106), CA (120), NV (224), UT (382); CANADA: SASK (380); Tax, 505.







MICROCOPY RESOLUTION TEST CHART NATIONAL BUREAU OF STANDARDS-1963-A

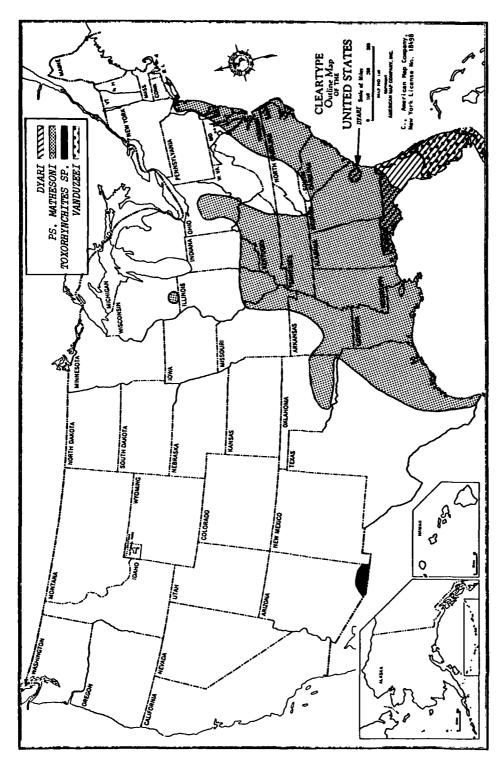


Plate 46. Distribution of Mansonia dyari - USA: FL, GA (106); Tax. 34, 389, 390. Psorophora mathesoni - USA: AL, AR, FL, GA, IL, IN, KY, LA, MS, MO, NY, NC, OH, OK, SC, TN, TX, VA (106), DE (254), MD (46), NJ (72), WI (466), Listed in all references as Ps. varipes; Tax. 33. Toxorhynchites sp. USA: AZ (512), Wyeomyia vanduzeei - USA: FL (106); Tax. 34.

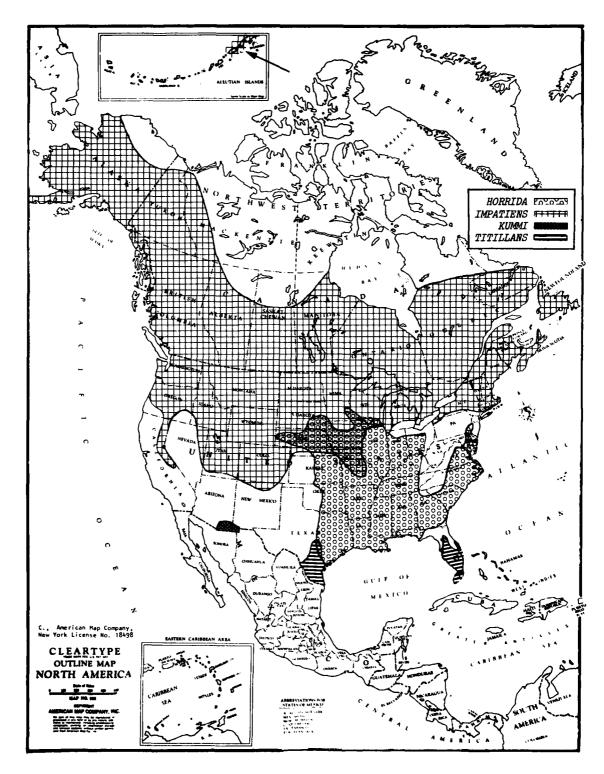


Plate 47. Distribution of Culiseta impatiens - USA: AK, CA, CO, ID, IA, ME, MA, MI, MO, MT, NE, NH, NY, OR, UT, VT, WA, WI, WY (106), CT (484), NV (382), NM (503), PA (Wills, in litt. 1979), SD (187); CANADA: ALTA, BC, LAB, MAN, NB, NWT, ON F, PQ, YUK (106), NFLD (360), SASK (380), Not in NS (505); Map modified after Hopla (227): Lax. 296, 505. Mansonia titillans - USA: FL, TX (106); Tax. 34, 389, 390. Orthopodomyta kummi-USA: AZ (307), NM (336); Tax. 508. Psorophora horrida - USA: AL, AR, DC, FL, GA, H., IN, IA, KS, KY, LA, MD, MS, MO, NE, NC, OH, OK, PA, SC, TN, TX, VA (106), DF (254), MI (Newson, in litt. 1977, no locality specified), MN (124), SD (187). WI (Dicke, in litt. 1979).

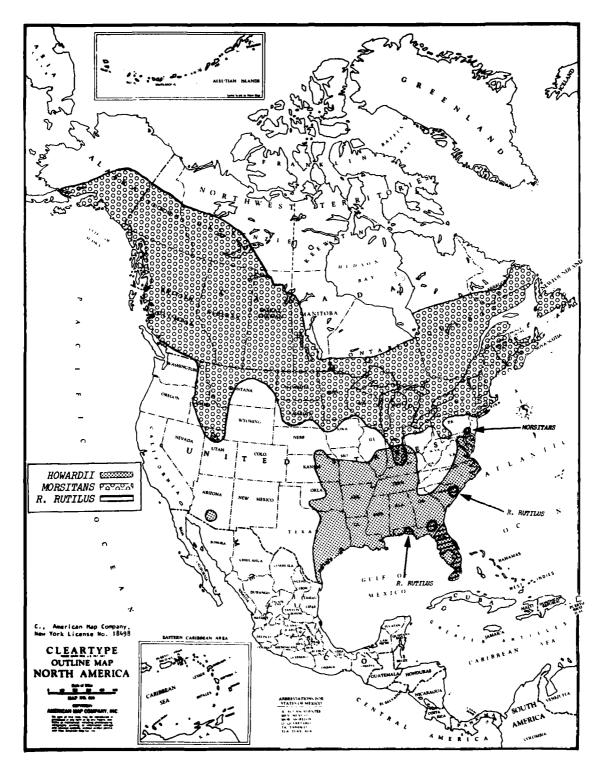


Plate 48. Distribution of Culiseta morsitans - USA: AK, CT, DE, ID, IL, IA, KY, ME, MA, MI, MN, NH, NJ, NY, ND, OH, PA, RI, SD, WI (106), IN (415), MD (50), MT (337), UT (330), VT (535), Not in CO (212); CANADA: AI TA, BC, LAB, MAN, NB, NWT, NS, ONT, PEI, PQ, SASK, YUK (106), NFLD (360); Tax. 15, 296, 505. Psorophora howardii - USA: AL, AR, DC, FL, GA, IL, IN, KS, KY, LA, MD, MS, MO, NE, NC, OK, SC, TN, TX, VA (106), AZ (382), DE (250), OH (Berry & Parsons, in litt. 1978). Toxorhynchites r. rutilus - USA: FL, GA, SC (106); Tax. 476.

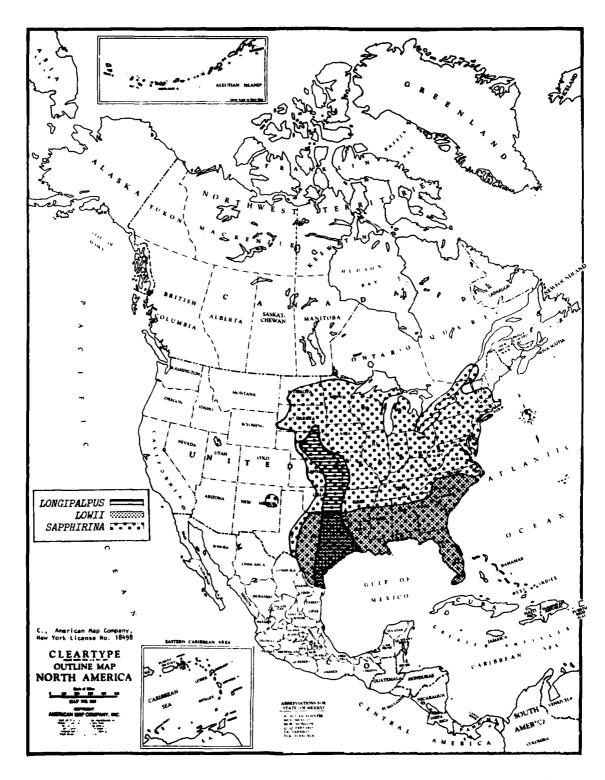


Plate 49. Distribution of *Psorophora longipalpus* - USA: AR, KS, LA, MO, OK, SD, TX (106), NE (376); Map after Roth (527); Tax. 246. *Uranotaenia lowii* - USA: AL, AR, FL, GA, LA, MS, NC, SC, TX (106), OK (353); Tax. 28, 34, 186. *Uranotaenia sapphirina* - USA: AL, AR, CT, DE, DC, FL, GA, IL, IN, IA, KS, KY, LA, MD, MA, MI, MN, MS, MO, NE, NH, NJ, NM, NY, NC, ND, OH, OK, PA, RI, SC, SD, TN, TX, VT, VA, WI (106), WV (3); CANADA: ONT, PQ (106); Tax. 34, 505.

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^{*} Includes mainly period from 1955 to 1978, i.e., since Carpenter & LaCasse (1955), but contains some references and distribution records before 1955 not cited by them.

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APPENDIX: LOCALITY DATA FOR MOSQUITO SPECIMENS USED TO PREPARE ILLUSTRATIONS FOR KEYS

For the benefit of mosquito taxonomists and other scientists interested in the localities from which the specimens were collected the following list is presented. Actually 96% of the specimens are from the USA and Canada, but for some of those species which are Neotropical in distribution and are found only in the extreme southern parts of USA, it was necessary to select specimens from the Caribbean islands, Mexico, Central America, Panama, and Colombia. Specimens were utilized from all states of the continental United States, except Iowa, Indiana, New Hampshire, South Dakota and West Virginia; and all provinces of Canada, except New Brunswick, Newfoundland, Nova Scotia, Prince Edward Island and Quebec. In all, adult females were selected from 38 states of the USA, 7 provinces of Canada and 8 foreign countries while larvae were from 35 states of the USA, 4 provinces of Canada and 9 foreign countries.

Since the mosquito fauna is better known in some states/provinces than in others, it is not surprising that specimens from only 11 states/provinces were used to prepare 50% of the adult illustrations, while larvae from 10 states/provinces accounted for 64.7% of the drawings used in the keys to immatures.

LOCALITY DATA FOR MOSQUITO SPECIMENS USED TO PREPARE ILLUSTRATIONS FOR KEYS

Figure Number	Species	Country	State/Province	County	Locality
1, 2	Tx. r. septentrionalis	USA	Delaware	Kent	Bombay Hool
3, 4	Ae. vexans	Canada	Ontario	Kenora	Dryden
5, 6	An. quadrimaculatus	USA	North Carolina	Robeson	Maxton
7, 8	Ae. vexans	Canada	Ontario	Kenora	Dryden
9, 10	Wy. smithii	USA	Massachusetts	Hampden	Westfield
11	Ae. vexans	Canada	Ontario	Kenora	Dryden
12	Ae. vexans	USA	North Dakota	Grand Forks	Grand Forks
13, 14	Ur. sapphirina	USA	Virginia	Fairfax	Falls Church
15	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
16	Ae. vexans	USA	North Dakota	Grand Forks	Grand Forks
7	Ps. ciliata	USA	Virginia	Accomack	Chincoteague
8	Cs. inornata	USA	Oregon	Portland	Portland
19, 20	Ma. titillans	Mexico	Tamaulipas		Tampico
21, 22	Ae. vexans	Canada	Ontario	Kenora	Dryden
23	Ps. ciliata	USA	Virginia	Accomack	Chincoteague
24	Ps. cyanescens	USA	Texas	Dallas	Dallas
25	Ae. vexans	USA	North Dakota	Grand Forks	Grand Forks
26	Ae. vexans	Canada	Ontario	Kenora	Dryden
27, 28	Cs. inornata	USA	Oregon	Portland	Portland
29, 30	Cx. pepiens	USA	New Jersey	Middlesex	Nixon
31	Hg. equinus	USA	Texas	Cameron	Brownsville
32	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
33, 34	Or. signifera	USA	Florida	Indian River	Vero Beach
35, 36	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
37	Cq. perturbans	USA	New York	Wayne	Fair Haven
8, 40	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
39	De. pseudes	USA	Texas	Cameron	Brownsville

Figure Numb >	Species	Country	State/Province	County	Locality
41, 42	Ae. purpureipes	USA	Arizona	Santa Cruz	Madera Canyon
43	Cx. pipiens	USA	New York	Brooklyn	Ft. Hamilton
44	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
45, 47	Ae. excrucians	USA	Montana	Glacier	Glacier Natl. Park
46	Ae. triseriatus	USA	Kentucky	Jefferson	Louisville
48	Ae. c. canadensis	USA	Minnesota	Roseau	Warroad
49	Ae. sollicitans	USA	Florida	Dade	Miami
50	Ae. vexans	Canada	Ontario	Kenora	Dryden
51, 52	Ae. taeniorhynchus	USA	Florida	Palm Beach	Palm Beach
53, 54	Ae. sollicitans	USA	Florida	Dade	Miami
55, 56	Ae. mitchellae	USA	Florida		
57, 60	Ae. sollicitans	USA	Florida	Dade	Miami
61, 62	Ac. nigromaculis	USA	Idaho	Ada	Boise
63, 64	Ae. papago	USA	Arizona	Pima	Mendoza Canyon
65	Ae. taeniorhynchus	USA	Florida	Palm Beach	Palm Beach
66	Ae. vexans	USA	North Dakota	Grand Forks	Grand Forks
67	Ae. aegypti	USA	Florida		
68	Ae. c. canadensis	USA	Minnesota	Roseau	Warroad
69	Ae. zoosophus	USA	Texas	Frio	
70	Ae. epactius	USA	Texas	Travis	
71, 73	Ae. vexans	Canada	Ontario	Kenora	Dryden
72	Ae. excrucians	USA	Montana	Glacier	Glacier Natl. Park
74	Ae. vexans	USA	North Dakota	Grand Forks	Grand Forks
75, 76	Ac. cantator	USA	New York	Long Island	
77	Ae. grossbecki	USA	Louisiana	Rapides	Alexandria
78	Ae. stimulans	Canada	Ontario	Carleton	Ottawa
79	Ae. squamiger	USA	California	Orange	Huntington
80	Ae. squamiger	USA	California	San Diego	San Diego
81, 82	Ac. grossbecki	USA	Louisiana	Rapides	Alexandria
83	Ae. n.gromaculis	USA	North Dakota	Ramsey	Devils Lake
84	Ae. nigromaculis	USA	Idaho	Ada	Boise
85, 86	Ae. increpitus	USA	Utah	Cache	River Heights
87	Ae. flavescens	Canada	Saskatchewan		Oxbow
88, 90	Ae. increpitus	USA	Utah	Cache	River Heights
89	Ae. excrucians	USA	Montana	Glacier	Glacier Natl. Park
91, 93	Ae. riparius	Canada	Alberta		Red Deer
92	Ae. stimulans	Canada	Ontario	Carleton	Ottawa
94	Ae. fitchii	Canada	Ontario	Algoma	White River
95, 96	Ae. riparius	Canada	Alberta		Red Deer
97, 98	Ae. aloponotum	USA	Oregon	Marion	Idanha
99-102	Ae. euedes	USA	Minnesota	Clearwater	Itasco State Park
103-106	Ae. fuchii	Canada	Ontario	Algoma	White River
107, 108	Ae. increpitus	USA	Utah	Cache	River Heights
109, 110	Ae. stimulans	Canada	Ontario	Carleton	Ottawa
111	Ae. fitchii	Canada	Ontario	Algoma	White River
112	Ae. mercurator	Canada	Yukon	Klondike	Dawson
113	Ae. stimulans	Canada	Ontario	Carleton	Ottawa
114	Ac. enedes	USA	Minnesota	Clearwater	Itasca State Park

Figure Number	Species	Country	State/Province	County	Locality
115-117	Ae. mercurator	Canada	Yukon	Klondike	Dawson
118-120	Ae. fitchii	Canada	Ontario	Algoma	White River
121, 122	Ae. stimulans	Canada	Ontario	Carleton	Ottawa
121, 122	Ae. euedes	USA	Minnesota	Clearwater	Itasca State Park
125, 124	Ae. dorsalis	USA	Oregon	Klamath	Klamath Falls
125, 126		USA	Massachusetts	Essex	Maillath Falls
129, 130	Ae. atropalpus Ae. melanimon	USA	Montana	Hill	Havre
	Ae. dorsalis	USA		Klamath	Klamath Falls
131-134		USA	Oregon	Elko	Carlin
135, 136	Ae. campestris	Taiwan	Nevada	EIKO	Carim
137	Ae. togoi		\(\(\text{C}_{\text{in}}\)	D	XA7
138-140	Ae. c. canadensis	USA	Minnesota	Roseau	Warroad
141, 142	Ae. atropalpus	USA	Massachusetts	Essex	,
143, 144	Ae. c. canadensis	USA	Minnesota	Roseau	Warroad
145, 146	Ae, c. mathesoni	USA	Florida	Clay	Camp Blanding
147-149	Ae. atropalpus	USA	Massachusetts	Essex	
150-152	Ae. sierrensis	USA	Washington	Mason	Lake Cushman
153-155	Ae. epactius	USA	Texas	Travis	
156-158	Ae. atropalpus	USA	Massachusetts	Essex	1
159	Ae. monticola	USA	Arizona	Pima	Sabino Basin
160	Ae. sierrensis	USA	Washington	Mason	Lake Cushman
161	Ae. varipalpus	USA	Arizona	Coconino	Williams
162,163	Ae, sierrensis	USA	California	Los Angeles	Pearblossom
164	Ae. sierrensis	USA	Washington	Mason	Lake Cushman
165, 166	Ae, deserticola	USA	California	Riverside	Joshua Tree Natl. Monument
167, 169, 170	Ae. f. pallens	USA	Louisiana	Rapides	Alexandria
168	Ae. triseriatus	USA	Kentucky	Jefferson	Louisville
171, 172	Ae. bimaculatus	USA	Texas	Cameron	Brownsville
173	Ae. purpureipes	USA	Arizona	Santa Cruz	Madera Canyon
174	Ae. hendersoni	USA	Colorado	Weld	Kuner
175	Ae. atlanticus	USA	North Carolina	Brunswick	Wilmington
176, 178	Ae. triseriatus	USA	Kentucky	Jefferson	Louisville
177	Ae. pullatus	USA	Colorado	Grand	Grand Lake
179	Ae. atlanticus	USA	North Carolina	Brunswick	Wilmington
180, 181	Ae, triseriatus	USA	Kentucky	Jefferson	Louisville
182, 184, 185	Ae. hendersoni	USA	Ohio	Portage	Ravenna
183	Ae. hendersoni	USA	Colorado	Weld	Kuner
186, 187	Ae. brelandi	USA	Texas	Brewster	Big Bend Natl. Park
188	Ae. trivittatus	USA	Missouri	Clay	Kansas City
189, 191	Ae, atlanticus	USA	North Carolina	Brunswick	Wilmington
190	Ae. infirmatus	USA	Florida	Gulf	
192, 193	Ae. scapularis	USA	Texas	Hidalgo	Mission
194, 195	Ae. infirmatus	USA	Florida	Gulf	
196, 197	Ae. burgeri	USA	Arizona	Santa Cruz	Bodie Canyon
198, 199	Ae. atlanticus	USA	North Carolina	Brunswick	Wilmington
200, 201	Ae. muelleri	USA	Arizona	Santa Cruz	Bodie Canyon
=VV, 4V1	Tree maraters	1 00/1	Allama	Sama Oluz	1 Marie Carryon

Figure Number	Specie.	Country	State/Province	County	Locality
202, 203, 205	Ae. atlanticus	USA	North Carolina	Brunswick	Wilmington
204	Ae. dupreei	USA	Louisiana	East Baton	Baton Rouge
				Rouge	
206	Ae. niphadopsis	USA	Utah	Salt Lake	Salt Lake City
207, 209	Ae. s. idahoensis	USA	Utah	Uintah	Ouray
208	Ae. pullatus	USA	Colorado	Grand	Grand Lake
210	Ae. niphadopsis	USA	Utah	Salt Lake	Salt Lake City
211, 212	Ae. s. spencerii	USA	North Dakota	Ramsey	Devils Lake
213, 214	Ae. s. idahoensis	USA	Utah	Uintah	Ouray
215, 216	Ae. ventrovittis	USA	Wyoming	Teton	Yellowstone Natl. Park
217	Ae, bicristatus	USA	California	Lake	Lower Lake
218	Ae. cataphylla	Canada	British Columbia		Cranbrook
219, 220	Ae. niphadopsis	USA	Utah	Salt Lake	Salt Lake City
221, 222	Ae, cataphylla	Canada	British Columbia	1	Cranbrook
223, 224	Ae. bicristatus	USA	California	Lake	Lower Lake
225, 226	Ae. cataphylla	Canada	British Columbia		Cranbrook
227, 229,	Ac. pullatus	USA	Colorado	Grand	Grand Lake
230					İ
228	Ae. diantaeus	USA	Michigan	Keweenaw	Copper Harbor
231, 232	Ac. implicatus	USA	Idaho	Kootenai	Athol
233, 234	Ae. intrudens	USA	Maine	Washington	Crawford
235, 236	Ac. pullatus	USA	Colorado	Grand	Grand Lake
237, 238	Ae, implicatus	USA	Idaho	Kootenai	Athol
239, 240	Ae, provocans	USA	Minnesota	Roseau	Warroad
241	Ac. diantaeus	USA	Michigan	Keweenaw	Copper Harbor
242	Ae. intrudens	USA	Maine	Washington	Crawford
243, 244	Ae. aurifer	USA	Delaware	New Castle	Glasgow
245-247	Ac. thibaulti	USA	Alabama	Lauderdale	Wilson Dam
248-250	Ae. decticus	USA	Massachusetts	Hampshire	Belchertown
251, 252	Ac, diantaeus	USA	Michigan	Keweenaw	Copper Harbor
253	Ae, sticticus	USA	Massachusetts	Hampshire	Northampton
254	Ae. punctor	USA	Massachusetts	Hampshire	Chesterfield
255	Ar, theleter	USA	Texas	Bexar	San Antonio
256-258	Ae, intrudens	USA	Maine	Washington	Crawford
259, 260	Ae, sticticus	USA	Massachusetts	Hampshire	Northampton
261, 262	Ae. cinereus	USA	Minnesota	Roseau	Warroad
263-265	Ae, intrudens	USA	Maine	Washington	Crawford
266	Ac. tortilis	Bahamas			
267, 268	Ac. rempeli	Canada	Northwest Territories		Baker Lakc
269-272	Ae, sticticus	USA	Massachusetts	Hampshire	Northampton
273-275	Ae, communis	USA	Michigan	Keweenaw	Copper Harbor
276, 277	Ac, nevadensis	USA	Nevada	Elko	Lamoille Canyon
278	Ae, churchillensis	Canada	Manitoba		Churchill
279, 280	Ac. ventrovittis	USA	Wyoming	Teton	Yellowstone Natl. Park
281	Ae. implicatus	USA	Idaho	Kootenai	Athol

Figure Number	Species	Country	State/Province	County	Locality
282	Ae. punctor	USA	Massachusetts	Hampshire	Chesterfield
283, 284	Ae. impiger	USA	Alaska	1	Nome
285	Ae. pionips	Canada	Ontario	Algoma	White River
286	Ae. implicatus	USA	Idaho	Kootenai	Athol
287, 288	Ae. impiger	USA	Alaska		Nome
289, 290	Ae. nigripes	Canada	Manitoba		Churchill
291, 292	Ae. schizopinax	USA	California	Nevada	Boca
293, 294	Ae. punctor	USA	Massachusetts	Hampshire	Chesterfield
295	Ae. implicatus	USA	Idaho	Kootenai	Athol
296	Ae. hexodontus	Canada	British Columbia		Prince Rupert
297-299	Ae. implicatus	USA	Idaho	Kootenai	Athol
300-302	Ae. punctor	USA	Massachusetts	Hampshire	Chesterfield
303, 304	Ae. pionips	Canada	Ontario	Algoma	White River
305-308	Ae, hexodontus	Canada	British Columbia		Prince Rupert
309, 310	Ae. punctor	USA	Massachusetts	Hampshire	Chesterfield
311	(Map)			'	
312	An. crucians	USA	Georgia	Baker	Newton
313	An. quadrimaculatus	USA	Arkansas	Arkansas	Stuttgart
314	An. earlei	USA	Minnesota	Ramsey	St. Paul
315	An, albimanus	Panama	Canal Zone	,	Gatun
316, 318-320	An. punctipennis	USA	Connecticut	Fairfield	Redding
317	An. crucians	USA	Georgia	Baker	Newton
321	An. pseudopunctipennis	USA	Texas	Cameron	Ft. Brown
322	An. pseudopunctipennis	USA	Texas	Travis	
323	An. punctipennis	USA	Connecticut	Fairfield	Redding
324	An. perplexens	USA	Florida		
325	An. pseudopunctipennis	USA	Texas	Travis	
326	An. pseudopunctipennis	USA	Texas	Cameron	Ft. Brown
327, 328	An. franciscanus	USA	New Mexico	Eddy	Artesia
329, 331	An. earlei	USA	Minnesota	Ramsey	St. Paul
330	An. quadrimaculatus	USA	Arkansas	Arkansas	Stuttgart
332	An. occidentalis	USA	California	Alameda	Palo Alto
333, 334	An, barberi	USA	Delaware	Kent	Bombay Hook
335	An, quadrimaculatus	USA	Arkansas	Arkansas	Stuttgart
336	An. freeborni	USA	California	Stanislaus	Modesto
337	An. barberi	USA	Delaware	Kent	Bombay Hook
338	An. judithae	USA	Arizona	Santa Cruz	Nogales
339	An. freeborni	USA	California	Stanislaus	Modesto
340	An. quadrimaculatus	USA	Arkansas	Arkansas	Stuttgart
341	An. walkeri	USA	Michigan	Livingston	''
342	An. atropos	USA	Louisiana	Plaquemines	Buras
343	An. freeborni	USA	California	Nevada	Auburn
344	An. quadrimaculatus	USA	Arkansas	Arkansas	Stuttgart
345-347	An. walkeri	USA	Michigan	Livingston	,"
348-350	An. atropos	USA	Louisiana	Plaquemines	Buras
351, 352	Cx. pipieus	USA	Oregon	Multnomah	Portland
353, 354	Cx. erraticus	USA	Florida	Dade	Miami
355	Cx. restuans	USA	Wisconsin	Dane	

Figure Number	Species	Country	State/Province	County	Locality
356	Cx. territans	USA	Virginia	Fairfax	Falls Church
357	Cx. tarsalis	USA	Texas	Victoria	Victoria
358	Cx. restuans	USA	Wisconsin	Dane	
359	Cx. tarsalis	USA	California		
360	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
361	Cx. bahamensis	USA	Florida	Monroe	Key Largo
362, 363	Cx. tarsalis	USA	Texas	Victoria	Victoria
364	Cx. tarsalis	USA	California		
365	Cx. peus	USA	California	Mariposa	
366	Cx. peus	USA	California	'	
367	Cx. thriambus	USA	Texas	Kerr	 Kerrville
368, 369	Cx. coronator	USA	Texas	Cameron	Weslaco
370	Cx. declarator	Costa Rica			Puerto Viejo
371, 372	Cx. erythrothorax	USA	California	San Luis Obispo	San Luis Obispo
373-375	Cx. nigripalpus	USA	Florida	'	'
	S. J				
376	Cx. restuans	USA	Wisconsin	Dane	
377, 378	Cx. nigripalpus	USA	Florida		ļ
379-381	Cx. salinarius	USA	Maryland	Calvert	 Chesapeake Beach
382	Cx. chidesteri	USA	Texas	Cameron	Brownsville
383	Cx. pipiens	USA	Tennessee	Campbell	Loyston
384	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
385-388	Cx. restuans	USA	Wisconsin	Dane	Madison
389, 390	Cx. interrogator	Panama	Canal Zone		
391	Cx. reevesi	Mexico	Baja California Norte		
392-394	Cx. territans	USA	Virginia	Fairfax	Falls Church
395	Cx. arizonensis	USA	Arizona	Yavapai	Prescott
396	Cx. apicalis	USA	Arizona	Cochise	Portal
397, 398	Cx. territans	USA	Virginia	Fairfax	Falls Church
399	Cx. boharti	USA	California	San Diego	San Diego
400	Cx. boharti	USA	California	Placer	Lake Tahoe
401	Cx. apicalis	USA	Arizona	Cochise	Portal
402	Cx. arizonensis	USA	Arizona	Yavapai	Prescott
403	Cx. latisquama	Colombia			
404	Cx. pipiens	USA	New Jersey	Middlesex	Nixon
405	Cx. erraticus	USA	Texas	Kinney	Brackettsville
406, 407	Cx. peccator	USA	Louisiana	La Salle	Olla
408	Cx. atratus	Cuba			Havana
409	Cx. opisthopus	USA	Florida	Dade	Miami
410, 412	Cx. peccator	USA	Louisiana	La Salle	Olla
411	Cx. abominator	USA	Texas	Bexar	
413	Cx. iolambdis	USA	Florida	Monroe	Key Largo
414, 415	Cx. atratus	Cuba		, , , , , , , , , , , , , , , , , , ,	Havana
416	Cx. pilosus	USA	Florida	Broward	Ft. Lauderdale
417	Cx. pilosus	USA	Florida	and the state of t	
418	Cx. paosis Cx. mulrennani	USA	Florida	Monroe	Big Pine Kev
419	CN, melanura	USA	Illinois	MONITOR	pig rine ixey

Figure Number	Species	Country	State/Province	County	Locality
420, 421	Cs. morsitans	USA	 Michigan	Livingston	
422	Cs. impatiens	USA	Colorado	Grand	Grand Lake
423, 424	Cs. particeps	USA	California	Humboldt	Arcata
425	Cs. morsitans	USA	Michigan	Livingston	1110444
426	Cs. impatiens	USA	Colorado	Grand	Grand Lake
427	Cs. particeps	USA	California	Humboldt	Arcata
428	Cs. alaskaensis	USA	Alaska		Anchorage
429	Cs. incidens	USA	Washington	Whatcom	Bellingham
430	Cs. impatiens	USA	Colorado	Grand	Grand Lake
431	Cs. minnesotae	USA	Minnesota	St. Louis	Virginia
432	Cs. morsitans	USA	Michigan	Livingston	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \
433, 434	Cs. inornata	USA	Missouri	St. Louis	W. St. Louis
435, 436	Cs. impatiens	USA	Colorado	Grand	Grand Lake
437	De. pseudes	Panama	Contracto	Grand	Orand Lake
438, 439	De. pseudes De. cancer	USA	 Florida	Indian River	Vero Beach
440	De. mathesoni	USA	Texas	Cameron	Brownsville
441	Ma. titillans	Cuba	1 CXas	Canicion	Brownsvine
442	Ma, titillans	USA	 Florida		ļ
443	Ma. dyari	USA	Florida	Indian River	Vero Beach
144 144	•	USA	Florida	Okeechobee	Okeechobee
445	Ma. dyari	Costa Rica	riorida	Океесновее	Okeechobee
	Or. kummi				1 TO 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
446, 447	Or. kummi	Panama		1	El Volcán
448 440	Or. alba	USA	Maryland	Anne Arundel	Patuxent
149	Or, signifera	USA	Louisiana	Chicot	Kilbourne
450	Or. alba	USA	Texas	Travis	Austin
451	Or, signifera	USA	Louisiana	Orleans	Camp Planché
452	Or, signifera	USA	Louisiana	Chicot	Kilbourne
453	Or. alba	USA	Maryland	Anne Arundel	Patuxent
454	Or, alba	USA	Texas	Travis	Austin
455, 456	Ps. columbiae	USA	Texas	Cameron	Brownsville
457	Ps. ciliata	USA	South Carolina	Beaufort	Parris Island
458	Ps. cyanescens	USA	New Jersey	Cumberland	Fairton
459	Ps. pygmaca	USA	Florida	Monroe	Key West
460-462	Ps. columbiae	USA	Texas	Cameron	Brownsville
463, 464, 466	Ps. discolor	USA	Georgia	Fulton	Ft. McPherson
465	Ps. signipennis	USA	Texas	Sutton	Sonora
467	Ps. ciliata	USA	South Carolina	Beaufort	Parris Island
468	Ps. cyanescens	USA	New Jersey	Cumberland	Fairton
469	Ps. ferox	USA	North Carolina	Columbia	Lake Waccamaw
470	Ps. ciliata	USA	South Carolina	Beaufort	Parris Island
171	Ps. ciliata	USA	South Carolina	Berkeley	McCellanville
472, 473	Ps. howardii	USA	Delaware	New Castle	Newport
474	Ps. cyanescens	USA	New Jersey	Cumberland	Fairton
475	Ps. cyanescens	USA	Texas	Dallas	Dallas
476, 477, 479	Ps. ferox	USA	North Carolina	Columbus	Lake Waccamaw
478	Ps. mathesoni	USA	Delaware	Sussex	Thompsonville
480	Ps. johnstonii	USA	Florida	Indian River	Vero Beach

Figure Number	Species	Country	State/Province	County	Locality
481, 482	Ps. mathesoni	USA	Delaware	Sussex	Thompsonville
483	Ps. varipes	Guatemala	Retalhuleu		Champerico
484	Ps. mexicana	USA	Texas	Cameron	Brownsville
485-487	Ps, ferox	USA	North Carolina	Columbus	Lake Waccamaw
488-491	Ps. horrida	USA	Louisiana	East Baton Rouge	Baton Rouge
492, 493	Ps. longipalpus	USA	Texas	Cameron	Brownsville
494	Ur, lowii	USA	Florida	Highlands	
495, 496	Ur. sapphirina	USA	District of Colombia		Washington
497, 498	Ur. a. anhydor	USA	California	Bernardino	Saratoga Springs
499	Ur. a. syntheta	USA	Texas	Cameron	
500, 501	Ur. vanduzeei	USA	Florida	Indian River	Vero Beach
502, 503	Wy. smithii	USA	New Jersey	Union	Rahway
504, 505	Wy, mitchellii	USA	Florida	Indian River	Vero Beach
506, 507, 509	Wy. smithii	USA	New Jersey	Union	Rahway
508	Wy, haynei	USA	South Carolina	Richland	Columbia
510	An. quadrimaculatus	USA	North Carolina		Camp Sutton
511, 513	Cx. pipiens	USA	Pennsylvania	Allegheny	Turtle Creek
512, 514, 515	Ma. dyari	USA	Florida	Palm Beach	W. Palm Beach
516	Cq. perturbans	USA	Minnesota	Clearwater	
517	Cq. perturbans	USA	Florida	Palm Beach	W. Palm Beach
518	Or, signifera	USA	Georgia	Fulton	Atlanta
519	Ae. aegypti	USA	Georgia	Chatham	Savannah
520	Tx. r. septentrionalis	USA	Georgia	Richmond	Augusta
521	Tx. r. rutilus	USA	Florida	Palm Beach	Boca Raton
522	Cx. pipiens	USA	Missouri	St. Louis	St. Louis
523	Ae. aegypti	USA	Georgia	Chatham	Savannah
524	Wy. smithii	USA	Maryland	Worchester	
525	Or, signifera	USA	Georgia	Fulton	Atlanta
526	Ur. sapphirina	USA	Florida	Palm Beach	Camp Murphy
527	Ur. sapphirina	USA	Georgia	Bryan	F ¹ . Stewart
528, 529	Ps. columbiae	USA	Delaware	New Castle	Summit Bridge
530, 531	De. pseudes	USA	Texas	Cameron	Brownsville
532	Ps. columbiae	USA	Delaware	New Castle	Summit Bridge
533, 535	Ac. aegypti	USA	Georgia	Fulton	Atlanta
534	Cs. inornata	USA	Louisiana	Rapides	Esler Field
536, 538	Cx. pipiens	USA	Pennsylvania	Allegheny	Turtle Creek
537	Ae. aegypti	USA	Georgia	Fulton	Atlanta
539	Ae. provocans	USA	New York	Tompkins	Ithaca
540	Ps. columbiae	USA	Delaware	New Castle	Summit Bridge
541, 542	Ac. atlanticus	USA	Georgia	Fulton	Atlanta
543, 545	Ae. aegypti	USA	Georgia	Chatham	Savannah
544	Hg. equinus	Guatemala			
546	Ae. provocans	USA	New York	Tompkins	Ithaca
547	Ac. aegypti	USA	Georgia	Chatham	Savannah
548	Ac. hemiteleus	USA	California	El Dorado	
549, 550	Ae, provocans	USA	New York	Tompkins	Ithaca
55 1	Ae, bicristatus	USA	California	Lake	
552	Ae. atlanticus	USA	Georgia	Fulton	Atlanta

Figure Number	Species	Country	State/Province	County	Locality
553	Ae. aegypti	USA	Georgia	Chatham	Savannah
554, 556, 557	Ae. nigromaculis	USA	California	Tulare	Visalia
555	Ae. abserratus	USA	New York	Tompkins	Ringwood
558, 559	Ae. f. pallens	USA	Louisiana	Rapides	Kiligwood
560	Ae. nigromaculis	USA	California	Tulare	Visalia
561	Ae. nigripes	Canada	Manitoba	Tulare	Visalia
562	Ae. f. pallens	USA	Louisiana	Rapides	
563	Ae. f. pallens	USA	South Carolina	Horry	Myrtle Beach
564, 565	Ae. thelcter	USA	Texas	Cameron	Brownsville
566	Ae. tormentor	USA	Kentucky	Bullitt	Ft. Knox
567	Ae. abserratus	USA	New York	Tompkins	Ringwood
568	Ae. bimaculatus	USA	Texas	Cameron	Brownsville
569	Ae. tormentor	USA	Kentucky	Bullitt	Ft. Knox
570	Ae. atlanticus	USA	Georgia	Fulton	Atlanta
571	Ae. sollicitans	USA	Illinois	St. Clair	Dupo
572	Ae. taeniorhynchus	USA	Florida	Palm Beach	, .
573	Ae. taeniorhynchus	USA	Florida	Highlands	Camp Murphy Avon Park
574, 575	Ae. abserratus	USA	New York	Tompkins	
576	Ae. taeniorhynchus	USA	Florida	Hillsborough	Ringwood MacDill Field
577	Ae. taeniorhynchus	USA	Florida	Palm Beach	
578	Ae. dupreei	USA	Georgia	Fulton	Camp Murphy Atlanta
579, 580	Ae. atlanticus	USA	Georgia	Fulton	Atlanta
581	Ae. sollicitans	USA	Illinois	St. Clair	· ·
582	Ae. atlanticus	USA	Georgia	Fulton	Dupo
583	Ae. hexodontus	USA	California	Tuolumne	Atlanta
584					Yosemite Natl. Park
585	Ae. punctor	USA	Maine	Penobscot	Orono
586, 587	Ae. sollicitans	USA	Illinois	St. Clair	Dupo
	Ac. mitchellae	USA	Mississippi	Harrison	Gulfport
588, 589 500, 500	Ae. sollicitans	USA	Illinois	St. Clair	Dupo
590, 592 591	Ae. infirmatus	USA	Florida	Highlands	Avon Park
	Ae. taeniorhynchus	USA	Florida	Palm Beach	Camp Murphy
593	Ae. trivittatus	USA	Illinois	Champaign	Champaign
594, 595	Ac. rempeli	Canada	Northwest Territories		Baker Lake
596	Ae. taeniorhynchus	USA	Florida	Hillsborough	MacDill Field
597	Ae. scapularis	Guatemala			
598	Ae. taeniorhynchus	USA	Florida	Hillsborough	Tampa
599	Ae. taeniorhynchus	USA	Florida	Hillsborough	MacDill Field
600, 601, 603 <u> </u>	Ae. scapularis	Guatemala			
502	Ae. scapularis	Dominican Republic			}
604, 605	Ac. tortilis	St. Lucia	1		
606	Ae. exerucians	USA	Massachusetts	Hampden	Springfield
507	Ae. melanimon	USA	California	Merced	, springing in
608, 610	Ae. cataphylla	USA	Oregon	Grant	Dixie Pass
509	Ae. excrucians	USA	Massachusetts	Hampden	Springfield
511	Ae. cataphylla	USA	California	Mono	, springricia

Figure Number	Species	Country	State/Province	County	Locality
612	Ae. atropalpus	USA	Maine	Hancock	Mt. Desert Is.
613-615	Ae. atropalpus	USA	Maryland	Montgomery	Bethesda
616, 617	Ae. epactius	USA	Texas	Comal	New Braunsfel
618, 620	Ae, diantaeus	USA	Vermont	Windham	Jacksonville
619	Ae. vexans	USA	Louisiana	Rapides	
621	Ae, diantaeus	USA	Michigan	Keweenaw	Isle Royale
622	Ae, aurifer	USA	Delaware	New Castle	New Castle
623	Ae, aurifer	USA	Maryland	Prince Georges	
624, 626, 627	Ae. s. spencerii	USA	Minnesota	Ramsey	
625	Ae. campestris	USA	Nevada	Churchill	
628	Ae, s. idahoensis	USA	Utah	Summit	Oakley
629	Ae. s. idahoensis	USA	Colorado	Grand	Grand Lake
630	Ae, excrucians	USA	Maryland	Cecil	Elkton
631	Ae. intrudens	USA	New York	Tompkins	Ringwood
632	Ae. excrucians	USA	Massachusetts	Hampden	Springfield
633	Ae. excrucians	USA	New York	Tompkins	MacLean
634-637	Ae. campestris	USA	Nevada	Churchill	
638	Ae, flavescens				
639	Ae, flavescens	W.Germany			Spandau
640	Ae, flavescens	USA	Alaska		Anchorage
641	Ae. flavescens	W. Germany			Spandau
642, 643	Ac. aloponotum	USA	Oregon	Marion	Idanha
644	Ae. intrudens	USA	New York	Tompkins	Ringwood
645	Ae, niphadopsis	USA	Utah	Tooele	Grantsville
646	Ae, vexans	USA	Georgia	Fulton	Atlanta
647	Ae. euedes	USA	Minnesota	Clearwater	Itasca State Park
648	Ae, intrudens	USA	Alaska		Steese Hwy.
649	Ae. intrudens	USA	New York	Tompkins	Ringwood
650, 651	Ae, euedes	USA	Minnesota	Clearwater	Itasca State Park
652	Ac. dections	USA	Massachusetts	Hampshire	Belchertown
653-655	Ae, niphadopsis	USA	Utah	Tooele	Grantsville
656	1e. niphadopsis	USA	Utah		
657-659	Ac, riparnis	USA	Minnesota	Ramsey	
660	Ac, cuedes	USA	Minnesota	Clearwater	Itasca State Park
661	Ac. ventrovittis	USA	California	Alpine	
662, 663	Ac, riparius	USA	Minnesota	Ramsey	
664, 665	Ac. ventrocuttis	USA	California	Alpine	
666	Ac. triscriatus	USA	Ohio	Portage	Ravenna
667	Ac. fitchin	USA	New York	Tompkins	Ringwood
668	Ac. purpurcipes	USA	Arizona	Santa Cruz	
669	Ac. triscriatus	USA	Ohio	Portage	Ravenna
670	Ac. papago	USA	Arizona	Pina	Mendoza Canyon
671	Ac, triseriatus	USA	Louisiana	Calcasie	Lake Charles
672,673	Ac. purpurcipes	USA	Arizona	Santa Cruz	
674-677	Ac. aegypti	USA	Georgia	Chatham	Savannah
678, 679	Ac. muelleri	USA	Arizona	Santa Cruz	Madera Canyon
680, 682	Ac, sierrensis	USA	California	San Diego	

Figure Number	Species	Country	State/Province	County	Locality
681	Ae. zoosophus	USA	Texas	Pecos	Sheffield
683, 685-687	Ae. monticola	USA	Arizona	Santa Cruz	
684	Ae. deserticola	USA	California	Los Angeles	
688	Ae. varipalpus	USA	Arizona	Coconino	
689	Ae. varipalpus	USA	Utah	Kane	
690, 691	Ae. burgeri	USA	Arizona	Santa Cruz	
692, 693, 695-697	Ae, triseriatus	USA	Ohio	Portage	Ravenna
694	Ae. zoosophus	USA	Texas	Pecos	Sheffield
698-700	Ae. hendersoni	USA	Colorado	Boulder	Boulder
701	Ae. brelandi	USA	Texas	Brewster	
702	Ae, impiger	USA	Alaska		Liberty Falls
703	Ae. cantator	USA	Rhode Island	Washington	Westerly
704	Ae. fitchii	USA	New York	Tompkins	Ringwood
705	Ae. c. canadensis	USA	Massachusetts	Hampshire	Belchertown
706	Ae. impiger	USA	Alaska	'	
707	Ae, stimulans	USA	Minnesota	Clearwater	
708	Ae. punctodes	USA	Alaska	1	Anchorage
709	Ae. impiger	USA	Alaska		Umiat
710	Ae, stimulans	USA	Minnesota	Clearwater	
711	Ae. aboriginis	USA	Oregon	Columbia	Vernonia
712	Ae. melanimon	USA	California	Kern	Bakersfield
713	Ae. sticticus	USA	Georgia	Bibb	Macon
714, 715	Ae. melanimon	USA	California	Merced	Macon
716, 717	Ae, stimulans	USA	Minnesota	Clearwater	
718, 719	Ae, nevadensis	USA	Nevada	Elko	Lamoille Canyo
710, 713 720, 721	Ae, stimulans	USA	.	Clearwater	Lamonic Canyo
720, 721 722, 723		Canada	Minnesota	Clearwater	D
	Ae, mercurator	Į.	Yukon Territory	D'I I	Dawson
724, 725	Ae. sticticus	USA	Georgia	Bibb	Macon
726 	Ae. flavescens	P.T.C. A	1	ł	1
727 799 - 790	Ae, flavescens	USA USA	Alaska	D'LL	Anchorage
728, 729	Ae, sticticus		Georgia	Bibb	Macon
730, 731	Ac. schizopinax	USA	California	Nevada	
732, 733	Ae. aboriginis	USA	Oregon	Columbia	Vernonia
734, 736, 737	Ae. pullatus	USA	Colorado	Larimer	Rocky Mt. Natl. Park
735	Ae. dorsalis	USA	Kansas	Stafford	D 11 .
738, 739	Ae. c. canadensis	USA	Massachusetts	Hampshire 	Belchertown
740, 741	Ac. pionips	USA	Michigan	Kewcenaw	Isle Royale
742	Ae. pullatus	USA	Alaska		Eklutna
743, 744	Ac. pullatus	USA	Colorado	Larimer	Rocky Mt. Natl. Park
745	Ae. pullatus	USA	Alaska		
746	Ar. cantator	USA	Maryland	Ann Arundel	Selby-on-Bay
747	Ae, cantator	USA	Rhode Island	Washington	Westerly
748	Ac. togoi	Canada	British Columbia		
749	Ae. c. canadensis	USA	Georgia	Rabun	1
750, 751	Ac. thibaulti	USA	Delaware	Sussex	Redden State Forest

Figure Number	Species	Country	State/Province	County	Locality
752, 753	Ae. c. canadensis	USA	Georgia	Rabun	
754	Ae. squaminger	USA	California	Marin	Richmond
755	4e. communis	USA	Minnesota	Clearwater	
756, 757	Ae. dorsalis	USA	Kansas	Stafford	
758, 759	Ae. increpitus	USA	California	Mariposa	
760	Ae. campestris	USA	Nevada	Churchill	
761-763	Ae. dorsalis	USA	Kansas	Stafford	
764, 765	Ae. grossbecki	USA	Louisiana	Rapides	Alexandria
766, 767	Ae. communis	USA	Alaska		Umiat
768-771	Ae. melanimon	USA	California	Merced	
772, 773	Ae. increpitus	USA	California	Mono	
774-776	Ae. implicatus	USA	Minnesota	Clearwater	
777-779	Ae. increpitus	USA	California	Mariposa	
780, 781	An. judithae	USA	Arizona	Cochise	Portal
782	An. albimanus	USA	Florida	Monroe	
783	An. albimanus	USA	Texas	Cameron	Cosmas
784	An. barberi	USA	Ohio	Stark	Canton
785	An. barberi	USA	Maryland	Montgomery	Cabin John
786	An. judithae	USA	Arizona	Cochise	Portal
787	An. judithae	USA	Arizona	Santa Cruz	Patagonia
788	An. albimanus	USA	Florida	Monroe	r atagoma
789	An, quadrimaculatus	USA	Tennessee	Dver	Dyersburg
790	An. albimanus	USA	Texas	Cameron	Cosmas
791	An. albimanus	USA	Florida	Monroe	Cosmas
792, 793	An. pseudopunctipennis	USA	Texas	Bell	Tomple
794	An, pseudopunctipennis	Dutch West	Curacao	Ben	Temple
795	An, pseudopunctipennis	Indies USA	Texas	 Hidalgo	 Edinburg
796, 797	An. franciscanus	USA	New Mexico	Eddy	Artesia
798	An. atropos	USA	Florida	Monroe	Key Largo
799	An, quadrimaculatus	USA	Tennessee	Dver	Dversburg
800	An, crucians	USA	Louisiana	Calcasie	Lake Charles
801	An. punctipennis	USA	Louisiana	Rapides	Esler Field
802	An. walkeri	USA	Tennessee	Obion	Walnut Log
803	An. walkeri	USA	Tennessee	Dver	Dversburg
804, 805	An, quadrimaculatus	USA	Tennessee	Dyer	Dyersburg Dyersburg
806, 808	An. bradleyi	USA	Mississippi	Harrison	Kessler Field
807	An, quadrimaculatus	USA	Louisiana	St. Charles	Narco
809	An, bradleyi	USA	Alabama	Mobile	Narco Mobile
810, 811		USA		Bibb	
812	An. georgianus		Georgia	·	Macon
	An. earlei	USA	Minnesota	Beltrami	Bemidji
813, 814	An, quadrimaculatus	USA	Tennessee	Dyer	Dyersburg
815, 817	An. punctipennis	USA	Louisiana	Rapides	Esler Field
816	An. occidentalis	USA	California	San Luis Obispo	
818	An. freeborni	USA	Utah	Salt Lake	Salt Lake City
819	An. freeborni	USA	Utah	Weber	Ogden
820	An, punctipennis	USA	Louisiana	Rapides	Alexandria
821	An, punctipennis	USA	California	Shasta	Tower House

Figure Number	Species	Country	State/Province	County	Locality
	Cx. pipiens	USA	Missouri	St. Louis	St. Louis
1	Cx, territans	1 - 1	Georgia	Fulton	Atlanta
,,,	Cx, bahamensis	USA	Florida	Monroe	Matecumbe Key
(/2 ((//2))	Cx. pipiens	USA	Pennsylvania	Allegheny	Turtle Creek
1	Cx. pipiens	USA	Missouri	St. Louis	St. Louis
l l	Cx. interrogator	USA	Texas	Cameron	Harlingen
•	Cx. tarsalis	USA	California	Contra Costa	Pittsburg
	Cx. restuans	USA	South Carolina	Richland	Columbia
,,,,,,	Cx. restuans	USA	North Carolina	Robeson	Maxton
833	Cx. thriambus	USA	California	Riverside	Coachella Valley
	Cx. coronator	USA	Texas	Cameron	Brownsville
835, 836, 838		USA	California	Contra Costa	Pittsburg
837	Cx. pipiens	USA	Pennsylvania	Allegheny	Turtle Creek
839	Cx. chidesteri	USA	Texas	Cameron	Brownsville
	Cx. declarator	USA	Texas	Caldwell	Luling
1	Cx. pipiens	USA	Pennsylvania	Allegheny	Turtle Creek
843	Cx. salinarius	USA	Maryland	Ann Arundel	Selby-on-Bay
1	Cx. peus	USA	California	Sacramento	Sacramento
	Cx. peus	USA	California	Marin	c
846	Cx. pipiens	USA	Missouri	St. Louis	St. Louis
847	Cx. pipiens	USA	Nebraska	Otoe	Dunbar Gulf Stream
848, 849	Cx. nigripalpus	USA	Florida	Palm Beach	Gulf Stream
850	Cx. salinarius	USA	Maryland	Ann Arundel	Selby-on-Bay
851, 853	Cx. salinarius	USA	Kansas	Douglas	
852	Cx. erythrothorax	USA	California	San Luis Obisp	tu l
854, 855	Cx. latisquama	Colombia	Tet · · ·	1	
856	Cx. peccator	USA	Florida	\ ₂ , ,	A.1
857	Cx. peccator	USA	Georgia	Fulton	Atlanta Ft. Gordon
858	Cx, territans	USA	Georgia	Richmond	Ft. Gordon
859	Cx. peccator	USA	Florida		Doggan
860, 861	Cx. arizonensis	USA	Arizona	Yavapai Clearwater	Prescott
862	Cx, territans	USA	Minnesota	Clearwater	Fe Comb
863, 865	Cx, territans	USA	Georgia	Richmond	Ft. Gordon Big Bend Natl.
864	Cx, apicalis	USA	Texas	Brewster	Big Bend Natl. Park
866	Cx. reevesi	USA	California	San Luis Obisp	po
866 867-869	Cx. recvesi Cx. territans	USA	Minnesota	Clearwater	
867-869 870, 871	Cx. writions Cx. boharti	USA	California	Benito	
870, 871 872	Cx. pilosus	USA	Louisiana	Orleans	Camp Villere
872 873	Cx. puosus Cx. atratus	Puerto Rico			Tortuguero
873 874	Cx. atratus Cx. pilosus	USA	Georgia	Fulton	Atlanta
874 875	Cx. pitosus Cx. erraticus	USA	Georgia	Baker	
875 876, 878	Cx, erraticus Cx, opisthopus	USA	Florida	Broward	Ft. Lauderdale
876, 878 877	Cx, opisthopus Cx, peccator	USA	Florida	1	
877 879	Cx. peccator Cx. opisthopus	USA	Florida	Dade	1
	Cx, opisthopus Cx, atratus	USA	Florida	Monroe	Vaca Key
880, 881 882	Cx. atratus Cx. abominator	USA USA	Fiorida Fexas	Comal	
882 883		USA USA	Florida	Martin	Jensen
883	Cx. iolambdis	1000	1 · manua	1	for a second

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885 Cx. iolambdis USA Florida Palm Beach Jupiter 886 Cx. aips Mexico Baja California Tijuana 887 Cx. petentor USA Florida Palm Beach Jupiter 888 Cx. iolambdis USA Florida Monroe Big Pine Key 890 Cx. mundua USA Florida Monroe Big Pine Key 891 Cx. mundua USA Colorado Larimer Estes Park 892 S93 Cx. mundua USA Minnesota Clearwater 895 Cx. inimucota USA Minnesota Clearwater 896 S97 Cx. inimucota USA Alaska Clearwater 900 Cx. inipatins USA Alaska Clearwater 901 Cx. inipatins USA Alaska Kernville 901 Cx. inipatins USA Colorado Larimer Estes Park 9012 903 Cx. inicidas USA	884	Cx. peccator	USA	Georgia	 Fulton	Atlanta
887	,	•	USA	Florida	Palm Beach	Jupiter
887	886		Mexico	Baja California		\
888 Cx. indambdis USA Florida Palm Beach Jupiter 889 Cx. melrenmani USA Florida Monroe Big Pine Key 890 Cx. melrenmani USA Florida Okaloosa Baker 891 Cx. mornata USA Colorado Larimer Estes Park 892 S95 Cx. mornata USA Minnesota Clearwater 896 S97 Cx. mornata USA Minnesota Clearwater 898 S96 Cx. mornata USA Minnesota Clearwater 890 Cx. mornata USA Minnesota Clearwater 890 Cx. mornata USA Minnesota Clearwater 890 Cx. funcrata USA California Kern Kerthikan 901 Cx. funcrata USA Delaware New Castle Newark 905 Ob Cx. funcidens USA California Kern Kernville 907 Cx. funcid		•	USA	.,		,
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907			1	 Delaware	New Castle	Newark
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957	Ps. ferox	USA	Georgia	Worth	
960	Ps. mathesoni	USA	Louisiana		
961	Ps. mathesoni	USA	Delaware	Sussex	Thompsonville
962, 963	Ps. ferox	USA	Georgia	Worth	
964	Ps. longipalpus	USA	Texas	Grayson	Denison
965	Ps. longipalpus	USA	Oklahoma	Tulsa	Tulsa
966	Ur. a. syntheta	USA	Texas	Bexar	San Antonio
967	Ur. sapphirina	USA	Georgia	Bryan	Ft. Stewart
968, 969	Ur. lowii	USA	Florida	Palm Beach	Boca Raton
970, 971	Ur. sapphirina	USA	Louisiana	Rapides	Alexandria
972, 973	Wy. mitchellii	USA	Florida	Palm Beach	Boca Raton
974	Wy. smithii	USA	Maryland	Prince Georges	Suitland
975	Wy. smithii	USA	Minnesota	Clearwater	
976	Wy. vanduzeei	USA	Florida	Palm Beach	Boca Raton
977	Wy. vanduzeei	USA	Florida	Dade	Miami
978	Wy. smithii	USA	Minnesota	Clearwater	}
979, 980	V. y. smithii	USA	Maryland	Prince Georges	Suitland
981	Wy. smithii	USA	Minnesota	Clearwater	
982, 983	Wy. haynei	USA	Georgia	Rabun	

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Identification and Geographical Distribution of the Mosquitoes of North America, North of Mexico

Supplement 1 to Mosquito Systematics

This volume contains keys for the identification of the adult females and fourth stage larvae of all 167 mosquito species and subspecies known to occur in North America, north of Mexico. Chapters on adult and larval morphology discuss the anatomical structures mentioned in the keys. accompanied by a series of full page plates. Separate generic keys for adult females and larvae are followed by keys to the species of each genus. All characters used in the keys are illustrated by 983 original drawings inserted between key couplets. In addition, in a separate chapter, the geographical distribution of each taxon is shown in a series of maps. Each is accompanied by a listing of the states (U.S.A.) and provinces (Canada) from which each taxon has been reported along with the relevant literature citations. Three tables of distribution provide a synopsis of the occurrence of mosquito species in the eastern United States, the western United States and Canada/Alaska. A bibliography of mosquito taxonomy and geographical distribution cites more than 500 references from 1955 to 1979.

It is anticipated that this text will be of value not only to mosquito control personnel, medical and aquatic entomologists, but also to introductory and advanced students of mosquitoes, due to full utilization of the standardization of morphological terms recommended by R. E. Harbach and K. L. Knight (1980) and the completely illustrated identification keys.

About the Authors:

Richard F. Darsie, Jr. is a research entomologist with the Medical Entomology Research and Training Unit, Guatemala City, where he is attached to the Universidad del Valle de Guatemala. He holds a B.A. from Bethany College (WV), a M.Sc. from the University of Pittsburgh and a Ph.D. from Cornell University. He was on the faculty of the Department of Entomology, University of Delaware for 12 years, worked for 3 years as a specialist in entomology in the malaria eradication program in Nepal with the Agency for International Development and since 1966 has been employed by the U.S.

Public Health Service, Center for Disease Control. Atlanta, GA. He was assigned to the Nepal malaria program for one additional year, then transferred to Manila where he served as entomologist in the Malaria Eradication Center for 4 years, following which he was Chief, Vector Borne Disease Training Activity in Atlanta for 5 years. During the subsequent 3 years he was assigned to the Central America Research Station, San Salvador. Dr. Darsie has authored some 60 scientific articles. He is a member of the Entomological Society of America, Entomological Society of Washington, and American Society of Tropical Medicine and Hygiene, besides the American Mosquito Control Association.

Ronald A. Ward has been a medical entomologist with the Walter Reed Army Institute of Research, Washington, DC, since 1958. He has a B.Sc. from Cornell University, a D.A.P. & E. from the London School of Hygiene and Tropical Medicine and a Ph.D. from the University of Chicago. Prior to joining the WRAIR, he was a biology instructor at Gonzaga University, Spokane, WA for 3 years. He has conducted field on malaria studies and trypanosomiasis in Southeast Asia, the Middle East and Africa. In 1966-67 he received the Secretary of the Army Research and Study Fellowship and worked in Professor P.C.C. Garnham's department at the London School. He has served on the Armed Forces Pest Management Board and is presently an Honorary Research Associate in entomology at the Smithsonian Institution. He is currently editor of Mosquito News, Journal of the American Mosquito Control Association and is on the editorial board of Mosquito Systematics. Dr. Ward has published more than 65 papers in medical entomology with emphasis on vector-parasite relationships. He is an active member of many scientific societies, including the American Mosquito Control Association, and is local Secretary (Washington, DC) of the Royal Society of Tropical Medicine and Hygiene.

